

Cardiovascular Biomarkers: Lessons of the Past and Prospects for the Future

Omran, F., Kyrou, I., Osman, F., Lim, V. G., Randeva, H. S. & Chatha, K.

Published PDF deposited in Coventry University's Repository

Original citation:

Omran, F., Kyrou, I., Osman, F., Lim, V.G., Randeva, H.S. & Chatha, K. 2022,
'Cardiovascular Biomarkers: Lessons of the Past and Prospects for the Future',
International Journal of Molecular Sciences, vol. 23, no. 10, 5680.

<https://dx.doi.org/10.3390/ijms23105680>

DOI 10.3390/ijms23105680

ISSN 1661-6596

ESSN 1422-0067

Publisher: MDPI

This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Review

Cardiovascular Biomarkers: Lessons of the Past and Prospects for the Future

Farah Omran ^{1,2,3} ID, Ioannis Kyrou ^{1,2,4,5,6} ID, Faizel Osman ^{1,7} ID, Ven Gee Lim ^{1,7}, Harpal Singh Randeva ^{1,2,3} and Kamaljit Chatha ^{1,8,*}

- ¹ Warwick Medical School, University of Warwick, Coventry CV4 7AL, UK; farah.omran87@hotmail.com (F.O.); kyrouj@gmail.com (I.K.); faizel.osman@uhcw.nhs.uk (F.O.); vengee.lim@uhcw.nhs.uk (V.G.L.); harpal.randeva@uhcw.nhs.uk (H.S.R.)
- ² Warwickshire Institute for the Study of Diabetes, Endocrinology and Metabolism (WISDEM), University Hospitals Coventry and Warwickshire NHS Trust, Coventry CV2 2DX, UK
- ³ Clinical Sciences Research Laboratories, University Hospitals Coventry and Warwickshire, Coventry CV2 2DX, UK
- ⁴ Centre of Applied Biological & Exercise Sciences, Faculty of Health & Life Sciences, Coventry University, Coventry CV1 5FB, UK
- ⁵ Aston Medical School, College of Health and Life Sciences, Aston University, Birmingham B4 7ET, UK
- ⁶ Laboratory of Dietetics and Quality of Life, Department of Food Science and Human Nutrition, School of Food and Nutritional Sciences, Agricultural University of Athens, 11855 Athens, Greece
- ⁷ Department of Cardiology, University Hospitals Coventry and Warwickshire NHS Trust, Coventry CV2 2DX, UK
- ⁸ Biochemistry and Immunology Department, University Hospitals Coventry and Warwickshire NHS Trust, Coventry CV2 2DX, UK
- * Correspondence: kamaljit.chatha@uhcw.nhs.uk



Citation: Omran, F.; Kyrou, I.; Osman, F.; Lim, V.G.; Randeva, H.S.; Chatha, K. Cardiovascular Biomarkers: Lessons of the Past and Prospects for the Future. *Int. J. Mol. Sci.* **2022**, *23*, 5680. <https://doi.org/10.3390/ijms23105680>

Academic Editor: Maria Luisa Balestrieri

Received: 30 April 2022

Accepted: 11 May 2022

Published: 19 May 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Cardiovascular diseases (CVDs) are a major healthcare burden on the population worldwide. Early detection of this disease is important in prevention and treatment to minimise morbidity and mortality. Biomarkers are a critical tool to either diagnose, screen, or provide prognostic information for pathological conditions. This review discusses the historical cardiac biomarkers used to detect these conditions, discussing their application and their limitations. Identification of new biomarkers have since replaced these and are now in use in routine clinical practice, but still do not detect all disease. Future cardiac biomarkers are showing promise in early studies, but further studies are required to show their value in improving detection of CVD above the current biomarkers. Additionally, the analytical platforms that would allow them to be adopted in healthcare are yet to be established. There is also the need to identify whether these biomarkers can be used for diagnostic, prognostic, or screening purposes, which will impact their implementation in routine clinical practice.

Keywords: cardiovascular diseases; biomarkers; future biomarkers

1. Cardiovascular Diseases: Overview

Cardiovascular diseases (CVDs) are the leading cause of death globally [1], including coronary and ischemic heart diseases, congestive heart failure, peripheral arterial diseases, deep vein or arterial thrombosis, pulmonary embolism (PE), and cerebrovascular diseases [2]. If detected early, many CVDs may be preventable by addressing relevant modifiable risk factors such as smoking, physical inactivity, obesity, diabetes, hypertension, and lipid disorders. Thus, it is important to have effective tools for screening, diagnosis, and prognosis. This is where biomarkers are increasingly recognized to play a key role. Aronson et al. defined a biomarker as a “biological observation that substitutes for and ideally predicts a clinically relevant endpoint or intermediate outcome that is more difficult to observe” [3]. Moreover, the National Institute of Health Consortium defined a biomarker as a “characteristic that is objectively measured and evaluated as an indicator

of normal biological processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention” [4,5]. Therefore, changes in biomarkers indicate physiological or pathological conditions, and may reflect responses to therapy. This wide-scope definition of biomarkers covers measurements of proteins, metabolites, and genetic factors [3]. In a broader view, imaging procedures used to identify and evaluate abnormal biological functions are also considered biomarkers [6]. Nevertheless, in 2009, the American Heart Association proposed an evaluation framework and defined criteria for how novel markers of cardiovascular risk should be evaluated in a standardized fashion before their clinical use can be recommended [7]. Depending on their role, biomarkers fall into three main classifications—namely screening, diagnostic, and prognostic biomarkers—and may have critical value in a corresponding clinical setting to evaluate risks associated with various factors related to health or disease. These factors can be the susceptibility to genetic traits or environmental factors, markers at points along the disease development, and/or progression, as well as subclinical or clinical or surrogate endpoints used in the evaluation of the safety and efficacy of therapeutic options. Moreover, focus has been increasingly placed on identifying biomarkers that can help to effectively target costlier resources (e.g., expensive treatments or invasive procedures) to the individuals/patients that would benefit most.

The present review will focus on circulating biomarkers, which can be measured in blood and are relevant to the clinical care of patients with CVDs.

2. Historic Overview: Early Biomarkers for CVDs

Creatine Kinase Myocardial Band (CK-MB) is one of the creatine kinase (CK) isoenzymes, representing the gold standard tests to detect and monitor cardiac injury in the 1980s [8]. CK is an intracellular enzyme that catalyses the reversible transformation of creatine and ATP to creatine phosphate and ADP, playing an important role in energy equilibrium of cells that sporadically needs high energy [9]. CK-MB is present in the myocardium, and it exists as two isoforms: namely CK-MB1 and CK-MB2, which are the plasma and tissue form, respectively [10,11]. Skeletal muscles also express CK-MB, albeit in lesser concentrations than the myocardium. [12]. As such, CK-MB is considered a cardiac marker and rises to detectable levels in the blood when there is damage to myocardium including ischemia or necrotic injury [9,13]. An increase in measured circulating CK-MB above the 99th percentile upper reference limit indicates a myocardial rather than muscular origin [14].

Elevated CK-MB values are often detected within three to four hours following the onset of acute myocardial infarction (MI), and are used to assist the diagnosis of MI [15,16] with the circulating concentration peaking within 24 h and returning to normal at 24–72 h [9,17]. Laboratory measurements of CK-MB simply illustrate the total of the isoforms CK-MB1 and CK-MB2 [18].

Apart from MI, other cardiovascular etiologies can cause elevation of CK-MB. These include myocarditis [19], pulmonary embolism [20,21], cardiac trauma [22], heart transplantation [23], chemotherapy-induced cardiotoxicity [24], and cardiac surgery [25]. However, CK-MB levels in these diseases/conditions do not add significant value to their diagnosis or the prognosis, and often cause confusion in the interpretation of the results [19,21–27]. Moreover, increased CK-MB levels do not specify the underlying etiology, and false-positive results can occur in a number of non-cardiovascular conditions, including end-stage renal failure [28], skeletal muscle trauma [22], muscular dystrophy [29], dermatomyositis [30], rhabdomyolysis [31], delirium tremens [32], and amyloidosis [33].

As an MI biomarker, CK-MB is now obsolete with the advent of Troponins (Tn), specifically high sensitivity Tn. As such, the CK-MB test now has limited value in the early and late diagnosis of acute MI and has been replaced by the Tn test, which is far more specific and sensitive [14,20]. However, its release kinetics can assist in diagnosing re-infarction if levels rise after initially declining following acute MI. Of note, it is reported that disparate troponin and CK-MB plasma levels are found in 28% of patients. In-hospital mortality rates are not significantly increased in patients with only CK-MB positive results

compared to patients with negative values of both biomarkers [15,34]. Thus, an isolated CK-MB elevation has limited prognostic as well as diagnostic value in patients with an acute coronary syndrome (ACS). In addition, although the newer assay of evaluating CK-MB2 is more sensitive, its major disadvantage is that it is relatively labour-intensive for the laboratory, which makes using it impractical. Accordingly, the expert consensus document on the Fourth Universal Definition of MI and myocardial injury now mentions only the cardiac Tn rise above the 99th percentile upper reference limit [14].

From a historic perspective, prior to CK and CK-MB being used as cardiac biomarkers, aspartate transaminase (AST) was the first used biomarker for acute MI diagnosis. Moreover, lactate dehydrogenase (LD) and its cardiac-specific iso-enzyme (LDH-1, and to a lesser extent LDH-2) were later introduced and used in the context of acute MI. In fact, CK-MB was typically requested alongside AST and LDH as a cardiac biomarker panel in most laboratories. It is noteworthy that myoglobin also rises after acute MI and can be used as useful cardiac biomarker in the differential diagnosis of suspected acute MI. However, the absence specificity and the large occurrence of false positive results diminished the usefulness of all these biomarkers, which are now almost completely replaced by more specific cardiac biomarkers; this will be discussed in the following sections.

3. Current CVDs Biomarkers in Clinical Practice

3.1. Troponins

Cardiac troponins (cTn), or the troponin complex, are composed of three regulatory proteins, namely troponin I (cTnI), troponin T (cTnT), and troponin C (cTnC). These are integral to the cardiac muscle thin filaments (actin protein) and play a critical role in regulating cardiac muscle activity including intracellular calcium concentrations throughout the contraction/relaxation process [35]. TnC is expressed in cardiac and slow skeletal myocytes [36], whilst cardiac isoforms of TnI and TnT are translated exclusively in the heart [37–40]. The proportion of both isoforms increase during human fetal development until they are solely expressed in the heart by the ninth month after birth [37–45]. Thus, both TnI and TnT seem optimal for investigating heart pathologies.

Currently, the evaluation of serum levels of TnT and TnI—the highly specific troponins of cardiac myocyte damage—is one of the most valuable biomarkers in the early diagnosis of MI associated with coronary vessel and ventricular remodeling [16,46]. Accordingly, a consensus statement by the: European Society of Cardiology (ESC), American College of Cardiology Foundation (ACCF), American Heart Foundation (AHA), and World Heart Federation (WHF) in 2018 defined a MI as a rise and/or fall in cTn with at least one value above the 99th centile upper reference limit in the context of symptoms or clinical evidence of myocardial ischaemia [14,47]. Different assays can be used to measure plasma concentrations of troponins including high and standard sensitivity methods. The Joint ESC/ACCF/AHA/WHF consensus recommend the detection of high-sensitivity (hs) troponin methods at the 99th percentile of an apparently healthy reference population at <10% variability [14,48]. This approach allows earlier detection of and increased diagnosis of MI, as well as a reduction of cases with unstable angina, improved risk stratification, better medication management, and selection of early invasive vs selective invasive strategy for patients with ACS [48–55]. Nevertheless, the ESC/ACCF/AHA/WHF consensus provides no recommendations for use of the 99th percentile value for risk assessment. Risk assessment is a separate issue and hs-Tn is used in this assessment. Importantly, interpretation of cTn levels should not be performed in isolation of clinical context, but with careful clinical evaluation in emergencies such as potential acute MI because cTn levels can be detected in apparently healthy populations, especially when measured by high-sensitivity methods [56,57]. Notably, triggers of cTn leakage not related to irreversible cardiomyocyte necrosis can include elevated troponin after prolonged physical activity, inducible MI, and prolonged episodes of tachyarrhythmia, even in presumably healthy individuals [58,59]. It is, however, not clear whether elevation of troponins in such conditions are similar to cells with membrane disruption following myocardial cell death. Furthermore, it is also not

known if an imbalance between oxygen demand and supply in patients with subclinical heart disease mediates such troponin secretion in apparently healthy individuals.

Overall, cTn have a short plasma half-life of around 2 h after release from injured myocardium. Of note, the released cTnT after MI exhibits discrete kinetics with two peaks; indeed, it starts to rise 3–4 h after the injury and slowly rises to a second lower peak over several days to even 2 weeks after the onset of injury [60–62]. cTnI is released quicker, within 1–2 h of MI [17,60,63]. As such, the pattern of cTn secretion assists with the MI diagnosis and prognosis. For instance, patients with large reperfused MI exhibit a classical biphasic time-release pattern of cTnT, while the release pattern of cTnT is different in non-reperfused MI as well as with small MIs [62,64]. It is reported that detecting the early peak may help in assessing the status of micro-vascular reperfusion, while the circulating cTn levels on day three or four represents MI size [62,65,66].

In addition, cTn are released into systemic circulation in response to cardiac myocyte injury such as in myocardial necrosis/ischemia, myocardial wall stress resulting from increased left-ventricular (LV) filling pressures, increased inflammatory cytokines, oxidative stress or catecholamine, and direct cellular destruction [62,67,68]. Thus, troponins have also gained a value as biomarkers of multiple CVDs other than MI. In this context, tako-tsubo cardiomyopathy (TC), which causes extensive acute regional wall motion dysfunction [69] and cannot be diagnosed based on ECG alone, results in a modest increase in cTn levels [69–73], with the peak occurring within 24 h and a more rapid decline in levels than seen in ACS. However, the elevation in levels of cTn do not reflect the dysfunction characteristic of TC [69]. Therefore, cTn release in TC appears to be unrelated to ischemic cardiomyocyte necrosis, which might explain why cTn have no prognostic value in TC [74,75]. In acute and chronic HF, the likely cause of troponin release is increased ventricular preload causing myocardial strain [76]. While the etiology is not specified through the rise in cTn [77–79], there is a strong prognostic value as the levels of increased cTn are closely linked with the severity of HF [80,81]. Furthermore, patients with aortic dissection display similar increases in cTn to that seen in MI associated with ACS, making cTn a poor diagnostic tool. Similarly, aortic stenosis and other valvular diseases display elevated TnT and TnI levels as reported in a significant body of literature [82]. For example, it was reported that hs-TnT plays a role as an independent prognostic factor for patients with aortic stenosis after valve replacement surgery [83]. Cardiac inflammatory conditions can also show increased cTn levels, most specifically in acute pericarditis through the involvement of the epicardium in the inflammatory process [84–86]. Furthermore, a stroke—both ischemic stroke and intracerebral haemorrhage—can cause an increase in cTn through secondary damage to the heart or a concomitant primary ACS [87–89]. Patients with acute pulmonary embolism (PE) also display increased cTn levels. Although the reason for this is not entirely understood, one explanation could be hypoxaemia resulted from perfusion-ventilation mismatch, hypoperfusion and paradoxical embolism from systemic veins to coronary arteries [90,91]. In patients with PE, cTnT peaks at lower levels and persists for a shorter period of time compared with cTnT values in acute MI [92]. Finally, elevated hs-cTnT precedes the advancement of hypertension and potentially identify people at future risk for hypertension and hypertensive end-organ damage [93–97]. However, there are no recommendations to measure hs-cTnT in screening for hypertension. Thus, more research is needed to establish the clinical use of troponins in the context of hypertension.

Disadvantages of Cardiac Troponins as a CVD Biomarker

Cardiac troponins are not specific to CVD as they can increase in various other conditions. These include sepsis, acute respiratory distress syndrome, chronic obstructive pulmonary disease, renal failure, diabetes, acute neurological event, chemotherapy, drugs, and toxins [60,98,99]. Additionally, cTn increase does not indicate a specific CVD etiology, e.g., whether it indicates MI, coronary artery disease, myocardial injury, or chronic dysfunction related myocardium or non-acute coronary syndrome ACS reasons. Therefore, the need for improved marker options yielding greater specificity remains obvious.

In addition, there are technical issues related to methods used to measure cTn. For instance, the use of high-sensitivity assays to measure troponins leads to the detection of cTn in the wider population with or without CVDs [100]. The efficiencies of cTn assays are also influenced by comorbidities, which cause measurement of inaccurate cTn circulation levels as seen in severe haemolysis [48,101–103]. Analytical issues related to heterophilic antibodies or interference of autoantibodies can cause false-positive or -negative results of cTn, which otherwise cannot be justified despite thorough clinical assessments [99,103,104]. There are many other analytical issues that interfere with the result and should be considered, such as non-specific binding, selection of matrix, and lot-to-lot variation. Thus, improving methods to measure troponins in order to avoid such issues is an important area for research.

3.2. Cardiac Natriuretic Peptides

Natriuretic peptides (NPs) are a group of hormones secreted from cardiomyocytes and are responsible for a spectrum of protective roles to the cardiovascular system including diuresis, natriuresis, and vasodilation, as well as opposition of the effects of the aldosterone-renin system [105,106]. Natriuretic peptides also influence metabolic processes such as lipolysis, weight loss, and improved insulin sensitivity [106]. Abnormalities of NPs are associated with multiple cardiovascular pathologies including HF, atrial fibrillation, systemic arterial hypertension, and inflammatory cardiac diseases, and therefore they are used as biomarkers to evaluate CVDs pathogenesis, diagnosis, prognosis, and therapy [107]. The main cardiac NPs in humans include atrial natriuretic peptide (ANP), B-type natriuretic peptide (BNP), and C-type natriuretic peptide (CNP) [108], which constitute a well-integrated regulatory system, and possess similar structural conformation but different potency [108,109].

The transcriptional regulation of ANP and BNP in humans is controlled by the heart cells [110,111]. C-type natriuretic peptide transcription is not controlled by any specific type of cardiomyocytes as it is derived primarily from endothelial cells and it presents within atrial and ventricular myocardium [112]. In pathological conditions, the key stimulant for a release of ANP and BNP from the heart is myocardial mechanical stretch and neurohormonal activation. Both BNP and ANP are elevated as a response to ventricle wall stress induced by stretched myocytes because of pressure overload or volume expansion of the ventricle, which are present in conditions of HF and MI [113–115]. The release of BNP or ANP leads to improved myocardial relaxation by inhibiting the vasoconstriction, fluid retention, and antidiuretic actions of the stimulated renin-angiotensin-aldosterone system [105,106,114,116].

All natriuretic peptides are synthesized as preprohormones, which are cleaved into an inactive N-terminal fragment (NTproANP, NT-proBNP) and the biologically active ANP [109,117–120] or BNP [116,121,122]. Both NT-proBNP and NT-proANP as well as biologically active BNP and ANP could be found in plasma [105,123,124]. The active ANP has a very short half-life of less than 5 min [125–127], which makes the use of NT-proANP more reliable as a biomarker because it is secreted correlatedly with ANP but has a markedly longer half-life of 60–120 min [117,128]. However, because NTproANP can be further cleaved into smaller fragments in vivo, the determination of mid-regional proANP (amino acids 53–90; MR-proANP) is the favorable detection site of ANP [117,129–131]. The half-life of BNP and NT-proBNP is 20 min and 90–120 min, respectively [124,132–134]. Circulating levels of BNP and NTproBNP can be used in patient care as they are correlated.

Elevation circulating levels of NPs' levels helps in the detection of CVDs and is associated with worse prognosis regardless of the etiology of the increase [105]. In fact, NPs are responsible for the classification of the severity, influencing care plans, and evaluating the prognosis of heart disease [132,135–141]. Both BNP and NT-proANP levels are reported to be independently predictive of the risk of death from major cardiovascular event, HF, stroke or transient ischemic attack, and atrial fibrillation [142]. Additionally, in apparently

healthy individuals, NTproBNP gives prognostic information of mortality and first major cardiovascular events that goes further than classical risk factors [143].

Natriuretic peptides are increased to very high blood levels in acute HF and to a lesser extent in chronic HF. For diagnosing acute and chronic HF, according to the guidelines of the ACCF/AHA/ESC, BNP and NT-proBNP are considered to be the most valuable and reliable biomarkers, with proportional increase to the severity of the disease giving information about HF prognosis [141,144–146]. This increase is because patients might actually manifest a state of BNP insufficiency, attributed to both a deficiency of active BNP and resistance to its actions [105,108]. However, ANP is also secreted in patients with chronic HF; its release is less marked than that of BNP and is suppressed in severe cases by BNP increase [144,147,148]. On the other hand, MR-proANP measurement can provide valuable additive information in HF assessment and it correlates with ventricular wall stress and HF severity [149,150].

As NPs affect multiple aspects of the cardiovascular system, their levels increase in various CVDs. Among these CVDs, acute ischemic heart diseases including MI are associated with an elevation of BNP and NT-proBNP levels, which is of value in predicting the prognosis and severity [105,151,152]. After a MI, NT-proBNP is increased for about 12 weeks and therefore might be more useful than BNP as a diagnostic and prognostic biomarker [113,153]. Furthermore, BNP and NT-proBNP are related to arrhythmias as both are increased in atrial fibrillation patients [154–156]. Parallel rise in the circulating levels of NPs and troponins were also reported in patients with various tachycardias [157]. Evidence from animal experiments suggests that BNP mRNA and its protein increase as early as 10 min after transient lethal ventricular arrhythmias, possibly mediated by myocardial stretch [158]. In addition, the levels of BNP are considerably elevated in TC, with a value of early BNP/cTnT and BNP/CK-MB ratios to distinguish TC from acute MI with higher accuracy than BNP alone [71]. This highlights that combining the determination of BNP levels with other biomarkers could provide additional benefit for the differential diagnosis of certain heart diseases. Patients with valvular diseases including aortic stenosis show increased NPs levels which are useful for clinically monitoring patients' pre- and post-valvular surgery. In general, levels of NPs are likely to rise with augmenting severity of the valvular dysfunction and with the consequential cardiac remodeling, giving significant prognostic details that could guide risk stratification and scheduling for surgery [159–165]. Elevated levels of NPs can also be seen in the pulmonary embolism, likely reflecting a right heart strain [166], although not usually to the same extent as in HF [166,167]. Additionally, patients with other pulmonary diseases can have elevated BNP levels but have much lower levels than in patients with HF [168–172]. Indeed, right heart abnormalities resulted from severe lung illness can lead to increased circulating levels of NPs [169,171]. Finally, the levels of NPs potentially present important information about the severity of hypertension [173]. One study reported a prognostic value for NT-proBNP to detect underlying cardiac structural damage in asymptomatic hypertensive patients [174], whilst another study suggested that BNP levels can aid in predicting the presence of ventricular wall dysfunction [126].

Disadvantages of Natriuretic Peptides as CVD Biomarkers

Increased circulating concentrations of NPs in circulation for CVD diagnosis are not adequately accurate or specific. In fact, expression of NPs is not exclusive to the heart. ANP is detected in the lungs, brain, liver, and in several cell types such as immune cells [175,176]. Similarly, BNP in humans is expressed in the brain, lungs, kidneys, aorta, and adrenal glands [176,177]. However, outside the heart, both ANP and BNP concentrations are much smaller. Overall, their diagnostic value is often complicated by increased levels due to non-cardiac conditions—some of which are potentially life-threatening. Such examples of non-cardiac pathologies include acute or chronic renal failure [178,179], ascitic liver cirrhosis [180,181], respiratory diseases [182], obesity, and endocrine disorders such as hyperaldosteronism [183,184], adrenal tumours [185], and hyperthyroidism [186,187]. Fur-

thermore, a number of physiological elements are associated with higher levels of NPs, including increased age and female gender [188–191].

4. Future: Prospective Biomarkers for CVDs

4.1. Heart-Type Fatty Acid-Binding Protein (H-FABP)

Heart-type fatty acid-binding protein (H-FABP) belongs to the FABP family and plays a role in multiple cellular activities including cardiac lipid transport, regulation of cell proliferation, and myocyte homeostasis [192–197]. The FABP family is detected in multiple tissues that demand increased activity of fatty-acids; H-FABP is mostly translated in the heart [198]. High amounts of H-FABP are found in muscle cells and are secreted very quickly after heart damage because of its low molecular size and free cytoplasmic location [199–201]. No cross-reaction was observed between H-FABP and other FABP types because of H-FABP's differences in morphology and immunology. Consequently, H-FABP displays a significant specificity for myocardial damage diagnosis [202].

Indeed, several reports have highlighted H-FABP as a specific former marker of myocardial damage. Compared to cardiac troponins, H-FABP circulation levels are elevated within three hours of the onset of acute MI, and reciprocate within 20 h [200,203–205]. Additionally, circulating H-FABP is significantly increased after 15 min of induced MI by transcoronary ablation of septal hypertrophy in patients with hypertrophic obstructive cardiomyopathy [199]. However, mixed data are available about its added value in MI. Some studies suggest incremental value of using H-FABP in conjunction with hs-troponins [206,207], while other studies do not indicate additional advantages of H-FABP levels with cTn for diagnosing acute MI [208–210].

Various reports have evaluated the function of H-FABP in patients with HF, suggesting an impartial association between H-FABP and HF outcomes, as well as the risk of detrimental cardiac injuries [141,211–215]. Higher levels of H-FABP are detected in patients with arrhythmia and HF [216]. Similarly, an impartial correlation of H-FABP with clinical outcomes has been documented in hypertensive patients with aortic valve disease [200]. Furthermore, cardiac remodeling characterised by ECG is found to be correlated with levels of H-FABP in patients that exhibited troponin levels below the current cut-offs [200]. It is worth mentioning that valvular replacement in patients with severe aortic valve stenosis is suggested to significantly reduce H-FABP plasma concentration, pointing at decreased ventricular wall stress and the prospective to ameliorate cardiac remodeling [217]. Moreover, H-FABP plays a role in risk assessment of PE because it indicates right-ventricular strain early [91,218–220]. Additionally, significant correlation between H-FABP and the risk of major adverse events and mortality was showed in PE patients, which explains why the 2019 ESC guidelines on the diagnosis and management of acute PE recommend the use of H-FABP for prognosis assessment [221], despite the fact that prospective trials are yet to be initiated.

Disadvantages of H-FABP as a CVD biomarker include unclear results in renal dysfunction caused by the fact that H-FABP elimination is completely dependent on the kidneys. In addition, H-FABP has low specificity in the presence of skeletal muscle injury as it increases in skeletal muscles pathologies [17].

Ultimately, no agreement for measuring H-FABP levels has been agreed upon in routine clinical practice in the context of CVDs. Future applications of H-FABP could involve initial investigation of ischemia and guidance of long-term treatment planning. In addition, H-FABP may play an essential role in diagnosis of HF, combined with clinical assessments, imaging, and a multi-biomarker approach.

4.2. Copeptin

Copeptin, a neurohormonal marker of stress, is the 39-amino acid C-terminal portion of arginine vasopressin (AVP) precursor peptide that is released from the posterior pituitary gland into the bloodstream upon appropriate stimuli [222]. The rate of release of copeptin is directly linked to the release of AVP, which is crucial to preserve the water balance of

the body. Indeed, AVP—also known as antidiuretic hormone (ADH)—controls the kidney reabsorption of free water, total volume of blood, osmolality of body fluid, vasoconstriction, and myocardial contraction, as well as cell proliferation [223]. AVP has a low molecular weight of nine amino acids and a very short half-life of 24 min [224], therefore its levels are better represented by circulating copeptin, which is more reliable to determine [225]. Indeed, copeptin has days of stability after blood withdrawal and is measured by a sandwich immunoluminometric assay within 3 h [225].

Copeptin is secreted at the onset of MI [226–228]. However, there are controversial data related to the usefulness of copeptin in MI diagnosis, with some data suggesting its value in identifying low-risk patients presenting to the emergency department with a potential acute MI [227,229]. Contrarily, other studies point out that if the rapid rule-out hs-cTn assays are available, copeptin has limited added value [72,230]. Further trials are needed to explore the potential benefits of the combination of copeptin with hs-cTn. Moreover, copeptin levels may provide prognostic information in combination with cTn [231,232]. Copeptin levels are also increased in several CVDs, including the development of HF where levels of circulating AVP are increased [233–236]. In fact, copeptin has been demonstrated to be a valuable prognosticator of the outcome and severity of HF, as well as independent prognosticator of mortality, generally indicating a poor prognosis [231,237–243]. Increased levels of copeptin are also seen in PE [91] and acute aortic syndrome [244], as well as hypertension [245–247].

Overall, copeptin is not a specific marker for CVD and it appears to be a complementary biomarker, which could be used as part of a multi-marker risk prognosis panel to better guide clinical decisions. However, more research is needed to confirm its application in routine practice.

Disadvantages of copeptin as a CVD biomarker include the large number of other disorders with which its elevated levels are associated, including respiratory disorders (e.g., acute exacerbation of chronic obstructive pulmonary disease, lower respiratory tract infections, and acute dyspnea), sepsis, hemorrhagic and septic shock, diabetes, metabolic syndrome, hyponatremia, vasodilatory shock, diabetes insipidus, polycystic kidney disease, intracerebral hemorrhage, ischemic stroke, and traumatic brain injury [223,248].

4.3. Adrenomedullin (ADM)

Adrenomedullin (ADM) is a peptide synthesised in many tissues—especially vasculature structural cells—and has numerous biologic actions, including being a renal vasodilator with natriuretic and diuretic properties [249–254]. ADM is derived from a precursor peptide (proADM) by post-translational processing, giving two cleaved inactive fragments (the mid-regional part of proADM (MR-proADM) and the C-terminus of the molecule) [255]. Titration of ADM secretion in the circulation is challenging because of the instant coupling of ADM to receptors in the proximity of its generation, inducing autocrine or paracrine activities. Additionally, its circulating levels are reduced by the short half-life of ADM and its instability [255,256]. Therefore, it is more practical to measure the levels of MR-proADM, which is more stable and directly reflects the ADM levels [255,257,258].

In acute MI, ADM levels were reported to increase in the early phase reaching a maximum after 2–3 days and returning to baseline after three weeks [259]. Furthermore, ADM is increased in HF [260–262], related to the severity and prognosis of HF, as well as to ischemic HF with ventricular dysfunction [251,258,261–268].

ADM is also far from being specific to CVDs since it is increased in various other diseases such as renal dysfunction, diabetes, and septic, haemorrhagic, or cardiogenic shock [269]. However, in patients with hypertension, ADM elevation directly correlates with the stages of hypertension as defined by the World Health Organization (WHO), as well as with the severity of target organ damage. This suggests a potential value for measuring ADM levels in hypertension, although the detailed relationship between plasma ADM levels and blood pressure is yet to be understood.

4.4. P-Selectin

P-selectin is a cell-surface adhesion molecule and is expressed on activated endothelial cells and platelets after stimulation by several inflammatory stimuli [270]. P-selectin has been suggested to participate in the pathology of atherosclerosis, a key risk factor for CVDs [271–274]. In fact, in patients with coronary artery disease or after acute MI, elevated levels of P-selectin are reported and hold an association with poor prognosis [275–278]. Furthermore, P-selectin was shown to provide incremental diagnostic value when used in combination with troponin and ECG to exclude acute MI with relatively high sensitivity [275]. Interestingly, increased circulating P-selectin levels are also linked to a higher risk of developing acute MI, stroke, and CVD-related death in healthy women [279]. Of note, P-selectin is also found to increase with exercise and age [280]. In this context, the value of P-selectin as a CVD biomarker remains to be clarified, independently of other established biomarkers that have demonstrated value for both screening and prognosis of atherosclerosis, i.e., cardiac troponins, natriuretic peptides, or C-reactive protein (CRP).

4.5. Soluble Urokinase -Type Plasminogen Activator Receptor (suPAR) and Plasminogen Activator Inhibitor-1 (PAI-1)

Soluble urokinase plasminogen activator receptor (suPAR) is a proinflammatory biomarker related to immune response and fibrinolysis inhibition, implicated in atherosclerosis [281–284]. Moreover, plasminogen activator inhibitor (PAI-1)—a potent inhibitor of fibrinolysis—is also linked with atherosclerosis, excessive fibrin accumulation and thrombus formation.

Increased circulating suPAR and PAI-1 levels are correlated with increased risk of major adverse CVD events, including death and MI in advanced coronary artery disease, as well as the early phase of atherosclerosis [285–288]. Indeed, in patients with MI, increased suPAR levels independently predict all-cause mortality and recurrent MI [289]. Similarly, increased PAI-1 concentrations were an independent risk factor for major CVD events after successful coronary stenting [290]. Moreover, elevated suPAR levels are linked to increased CVD risk in apparently healthy individuals [291,292]. However, association between suPAR and disease development and mortality in the general population was not specific to CVDs, since circulating levels of suPAR and PAI-1 were also associated with the development of inflammatory diseases, cancer, type 2 diabetes, and mortality in the general population [291,293].

4.6. Extracellular Matrix Remodelling

4.6.1. Galectin-3 (GAL-3)

Galectin-3 (GAL-3) is a member of the β -galactoside-binding lectins family, which is implicated in altering “cell-to-cell” and “cell-to-matrix” interactions, as well as regulating cellular functions at the cell surface [294–297]. GAL-3 is also involved in cell adhesion, proliferation, apoptosis, and angiogenesis. Notably, GAL-3 enhances fibrosis and its circulation levels are associated with cardiac remodelling and ventricular hypertrophy [298]. In fact, elevated GAL-3 levels aid in detecting early stage of myocardial dysfunction and HF, as well as other heart diseases, including ACS and acute myocarditis [299–315]. Moreover, this increase in GAL-3 levels is linked to a worse prognosis in these conditions [316,317], representing a potential sensitive biomarker for CVDs [301,302,313–315]. As a novel biomarker, GAL-3 has been recommended by the ACC/AHA guidelines for assessment of myocardial fibrosis in HF, whereas the ESC has not recommended the clinical use of GAL-3 [146]. Left ventricular remodelling following acute MI was found to hold no correlation with serum GAL-3 levels [302]. In addition, GAL-3 could not predict the mortality in patients with HF compared to other biomarkers, while it showed a distinctive advantage when used as in a member of multi-biomarker panel [318].

As with other potential CVD biomarkers, GAL-3 is also detected in many non-cardiac disorders, including kidney disease [319–322], diabetes [319,320,323], viral infections [306,324–326], autoimmune diseases [327–330], neurodegenerative disorders [331–335],

and tumour formation [336–345]. Collectively, the usefulness of GAL-3 to diagnose cardiac disorders is limited, and more research is needed to confirm its advantages as a prognostic CVD biomarker.

4.6.2. Matrix Metalloproteinases (MMPs) and Their Tissue Inhibitors (TIMPs)

Matrix metalloproteinases (MMPs) are enzymes that degenerate the formational constituents of the extracellular matrix and divert biologically active elements, such as growth factors, cytokines, and chemokines [346]. The actions of MMPs are closely regulated by the endogenous tissue inhibitors of metalloproteinases (TIMPs). The interaction's equilibrium between MMPs and TIMPs is responsible for maintaining tissue homeostasis and changes in this equilibrium are involved in progression of CVDs [347–354]. Furthermore, members of MMPs and TIMPs are proposed as promising biomarkers to predict future CVD events. For instance, in patients with coronary artery stenosis, serum levels of TIMP-1 were reported to exhibit a correlation with the incidence of major adverse cardiac events, and, thus, the prognosis in these patients [355–357]. Additionally, circulating levels of TIMP-1 rise in males more than in females, and elevate with well-known CVD risk factors, such as age, body mass index, the ratio of lipoprotein cholesterol, smoking, and diabetes [358]. Another example is MMP-3, which was shown to be associated with MI in one study [302]. Similarly, MMP-8 levels were associated with the risk for a coronary artery disease event, MI and death [359]. Of note, MMP-9 represents the most studied MMP in relationship to CVDs, with its levels reflecting the severity of the infarction and predicting mortality in MI patients [360–362], as well as the progression of HF [363,364]. Moreover, in hypertension, induced-MMP-9 at a very early stage enhances collagen breakdown and arterial destruction, with hypertensive patients presenting increased serum MMP-9 levels, which were related to aortic stiffness [365]. Further investigations are necessary to determine the advantages of the use of specific MMPs and TIMPs for diagnosis and prognosis of CVD in clinical practice.

4.7. Inflammatory Markers

4.7.1. Growth Differentiation Factor 15 (GDF-15)

Growth Differentiation Factor 15 (GDF-15) is a stress-responsive cytokine of the transforming growth factor - β (TGF- β) superfamily, constituting a biomarker of inflammation, as well as oxidative stress and hypoxia. Of note, CVDs are a main reason for GDF-15 up-regulation, which is hypothesized to reflect a defense mechanism in both acute and chronic cardiac injury [366,367]. Interestingly, the biological actions of GDF-15 are dependent on the underlying pathophysiology and may differ with the stage of the illness [366,368–372]. In fact, increased serum GDF-15 levels are observed in acute MI [366,373] and acute HF [374,375]. Furthermore, GDF-15 levels have been associated with adverse events in community-dwelling populations and higher risk to develop CVDs in patients with non-CVDs disorders [376–385], as well as adverse prognosis in patients with ACS [386–389] and HF [63,318,375,390–392]. Moreover, serial measurements of GDF-15 have been found to improve risk prediction models in patients with HF [390]. In addition, in patients with atrial fibrillation, increased GDF-15 levels are associated with major bleeding, mortality, and stroke [154,393–396]. Accordingly, GDF-15 has a potential valuable role in assessing the risk of major bleeding in patients treated with oral anticoagulant therapy [397]. Collectively, GDF-15 could be utilized for screening and may have prognostic value, especially as in the context of a composite biomarker panel [7,378,380,385,386,390,391,396,398–400]. Further research is necessary to evaluate the role of GDF-15 for guiding clinical management decisions and treatment options for CVD, in comparison to and in combination with other CVD biomarkers and clinical predictors.

However, GDF-15 expression is not specific to the myocardium, since this cytokine is also expressed in various other tissues/cells such as macrophages, endothelium, smooth muscle, and adipocytes. Thus, levels of GDF-15 may also rise in different conditions that are not

related to CVDs, such as advanced age, obesity, diabetes, and kidney dysfunction [401–405], limiting its diagnostic value for CVDs [404,405].

4.7.2. Endothelin-1 (ET-1)

ET-1 is a potent hormone with multiple actions regulating vasoconstriction and renal sodium excretion [406–409], and is produced by many organs, including the heart and kidneys. Of note, the major source of ET-1 is the vascular endothelium, with ET-1 being constantly synthesised in the blood vessels to preserve vascular resistance and pressure, as well as to regulate blood pressure [407,410]. As such, ET-1 levels have been shown to correlate with inflammation, vasoconstriction, vascular and cardiac hypertrophy, and with the development and progression of CVDs [406,411,412]. In addition, both ET-1 and C-terminal proET-1 levels have been shown to increase with age [413–417], lung function [418–420], chronic kidney disease [413,415,416,421], and cancer [422]. Moreover, elevated ET-1 levels are associated with HF, coronary artery disease, hypertrophic cardiomyopathy, hypertension, and cervical artery dissection [307,411,423,424]. Indeed, ET-1 has been demonstrated to have a potential use as a prognostic biomarker in many of these CVDs, including mortality in patients following acute MI and diagnosis of congestive HF. For example, in patients with HF, measuring ET-1 levels added prognostic information that was supplementary to that provided by NT-proBNP [425]. Interestingly, the ET-1 active form was initially thought to be unstable in serum with a short plasma half-life of 1.4–3.6 min [426,427], which restricts its clinical use. Later on, ET-1 was reported to have a large volume of wide distribution and an extended half-life of 7.5 h [428]. Therefore, the developed C-terminal segment of pro-Endothelin-1 (CTproET1) and other ET-1 surrogate peptides as the more stable form of ET-1 may not be as disadvantageous as has been initially suggested. Notably, in patients with chronic coronary artery disorder or acute MI, CT-proET-1 has been linked with cardiovascular death and HF independent of clinical variables, and demonstrated prognostic value comparable to BNP or NT-proBNP [267,429,430]. Additionally, increased blood levels of C-terminal proET-1 and ET-1 have been correlated with larger left atrial size, and all-cause mortality in samples of general populations [413,414], which potentially hold screening value. This, however, requires further study in diverse populations. Hence, the use of ET-1 levels to select patients for primary preventive strategies and to guide personalised treatment regimens are future directions to be explored.

4.7.3. Suppression of Tumorigenicity 2 (ST2)

Suppression of Tumorigenicity 2 (ST2) is a cellular receptor for interleukin-33 (IL-33) and belongs to the interleukin-1 receptor family [431]. ST2 has two variants, including the transmembrane (ST2L) and soluble (sST2) isoforms, which can be detected in circulation [432]. Notably, ST2 is transcribed by cardiomyocytes and vascular endothelial cells along with its agonist IL-33 following myocardial damage [431,433]. Coupling of IL-33 and ST2L potentially suppresses hypertrophy, fibrosis, and apoptosis of the heart, and alleviates detrimental cardiac remodelling [431,433]. However, sST2 competes with ST2L to bind IL-33, leading to suppression of the protective effects of the ST2L-IL33 pathway to the heart [431,433]. In fact, it was reported that blood levels of sST2 are markedly elevated in HF onset and worsening of chronic HF—as well as with MI—aortic valve impairments and hypertension [217,401,434–439]. Therefore, sST2 represents a potential predictor of CVDs prognosis [217,401,434–436,439]. Indeed, measuring sST2 is recommended as prognostic marker in HF according to ACCF/AHA guidelines [146,440].

Although sST2 can play an important role as a prognostic marker in the context of CVDs, it has limited diagnostic value because increased sST2 levels are not specific for any human disease. Indeed, such high sST2 levels may occur not only in CVD, but also in pulmonary diseases, burn injuries, and immune diseases [432]. Additionally, the exact origin of circulating sST2 is not clear, and studies did not prove that cardiac cells were solely responsible for serum sST2 changes in CVDs [441–444]. Indeed, sST2 is expressed by several tissues, such as the colon and many haematopoietic cells (e.g., basophiles,

CD4 lymphocytes, eosinophils, macrophages, and T-helper 2 cells) [432]. Overall, the contribution of non-cardiac secretion to the total circulating sST2 levels, as well as the pathophysiologic implications of sST2 in CVDs, is not well studied yet and represents an area for further research.

4.7.4. Lipoprotein-Associated Phospholipase A2 (Lp-PLA2)

Lipoprotein-associated phospholipase A2 (Lp-PLA2) is secreted by inflammatory cells that are involved in forming vulnerable plaques and developing atherosclerosis [445,446]. In fact, elevated Lp-PLA2 levels are associated with increased risk of coronary events [447], unrelated to the levels of total cholesterol [448]. In patients diagnosed with CVDs, levels of Lp-PLA2 also predict adverse outcomes indicating its prognostic value [449]. Further investigations are needed to confirm the potential value of Lp-PLA2 as a biomarker for CVDs.

4.7.5. Soluble CD40 Ligand

Soluble CD40 Ligand (sCD40L) is a mediator of vascular inflammation that is implicated in atherogenesis, activating CD40 receptor on various cells that contribute to atherosclerosis progression (e.g., on macrophages, endothelial cells and T-cells) [450,451]. Notably, CD40L antibodies and CD40L deficiency have been associated with reduced atherosclerosis [452]. Furthermore, sCD40L concentrations have been associated with prognosis in ACS [453,454]. However, the current knowledge about sCD40L is limited, and exploring its value for the clinical practice related to CVDs is needed.

4.8. MicroRNAs

Out of the three billion base pairs in the human genome, only about 1% directly code for proteins. The remainder were previously thought to be ‘junk’ DNA, but have now been recognised to play important roles in gene regulation and function. Some of these non-coding RNAs include short microRNAs (miRNAs) and longer, long non-coding RNAs (lncRNAs).

miRNAs are approximately 22 nucleotides in length and were first discovered in 1993. Since then, the field has expanded exponentially with 2000 miRNAs having been identified in humans (<http://www.mirbase.org>) (accessed on 14 April 2022). A key feature of the mechanism of action of miRNAs is that a single miRNA can regulate the expression of several genes, whilst, conversely, individual genes can be regulated by different miRNAs [455]. The factors affecting this regulatory mechanism are highly complex and are still not well-understood. Nevertheless, multiple basic science studies have found that miRNAs play a role in normal cardiac development, as well in CVDs, including (with key implicated miRNAs in parenthesis) HF (miR-133, miR-1, miR-25), atherosclerosis (close to 70 different miRNAs), arrhythmias (miR-1, miR-328, miR-223, miR-664), and hypertension (miR-181a, miR-663, miR-132, miR-212, miR-143/145).

Notably circulating miRNAs are highly stable and have been reported to be implicated in multiple cardiovascular conditions. In terms of clinical studies, the expression of each one of miR-21, miR-486-5p, miR-146a, miR-664a-3p, miR-195, miR-217, miR-126, miR-143, miR-146a, and miRNA-210 was altered in patients with severe vascular disease and AMI, as well as atherosclerosis. For example, in HF, altered circulating levels of miRNAs were reported for miR-122, miR-210, miR-423-5p, miR-499, miR-622, miR-16, miR-27a, miR-101, and miR-150. Such miRNAs are suggested to play a role as diagnostic and prognostic CVD biomarkers [456–459].

Although recent discoveries in miRNA research highlight their diagnostic, prognostic and perhaps even therapeutic value in CVDs, there is still lack of understanding regarding how exactly these miRNAs regulate gene expression. Additionally, the variability in miRNA-based phenotype regulation in CVD, the interactions between multiple miRNAs with their shared cognate mRNAs, and the effect of comorbidities on the circulating miRNAs levels are still important areas that need further investigations [456–459].

4.9. Other Biomarkers

Finally, traditional inflammatory biomarkers are involved in the pathology of CVDs, including serum amyloid A [460], osteoprotegerin (OPG) [307,434,461,462], myeloperoxidase, CRP, erythrocyte sedimentation rate (ESR), cytokines, neutrophils, and monocytes [463]. Furthermore, well-described biomarkers of diseases other than CVDs provide important information about the progression of a given CVD. For instance, since there is a close relationship between functional regulation of the kidneys and the heart, it is not surprising that markers related to kidney function such as cystatin C, uric acid, and albuminuria play a role in evaluating patients with various CVDs (e.g., in patients with HF). Similarly, markers of metabolic disorders are considered to have a prognostic information for the severity of CVDs, such as the lipid profile, vitamin D, fetuin-A, and diabetes-related biomarkers of, for example, blood glucose and haemoglobinA1c levels. These may be valuable additions in multi-marker approaches for effective screening, diagnosis, and prognosis of CVDs.

5. Conclusions

In conclusion, the contribution of measuring CVD biomarkers has become increasingly essential for the clinical practice, with an increasing number of established and emerging CVD biomarkers (Table 1). Such biomarkers are increasingly becoming more specific to cardiovascular pathophysiology, with enhanced sensitivity and specificity. For example, the National Institute for Health and Care Excellence (NICE) recommends measuring NT-proBNP in people with suspected HF to direct care plan. Urgent referral to have specialist assessment and transthoracic echocardiography is suggested when NT-proBNP level is above 2000 ng/L [464]. Additionally, according to the 2017 ACC/AHA/HFSA guidelines for the use of biomarkers in the management of HF, measurement of BNP or NT-proBNP was recommended to diagnose HF class I according to the New York Heart Association (NYHA) functional classification, as well as at hospital admission and discharge for added risk stratification in HF class I and IIa, respectively [465]. Furthermore, Tns are recommended to evaluate hospital admission prognosis in HF class I, whilst sST2 and GAL-3 received a class II recommendation for risk prediction in HF [465]. In addition, other novel biomarkers, such as MR-proANP and ADM, have been reported to be independent predictors of HF and to correctly reclassify HF patients [241,264]. Currently, the role of established CVD biomarkers has progressed from the simple retrospective confirmation of an already diagnosed condition to a central position in the screening/diagnostic/prognostic clinical algorithms. However, the use of many novel CVD biomarkers has yet to be employed in the clinical practice. This partly due to still insufficient knowledge regarding their exact clinical usefulness and their potential advantages over existing established biomarkers. Moreover, there are also technical issues (e.g., lack of readily commercial clinical assays) and challenges regarding the precise determination of any novel biomarker as an analyte. Therefore, intensive research is still required in this field to address the existing gaps in our knowledge regarding the clinical use of the aforementioned novel biomarkers in the context of CVD. Indeed, clear evidence is needed to prove the potential benefit of such novel biomarkers for CVD screening/diagnostics/prognostics, as well as potential benefits for personalized/biomarker-guided therapies. In this setting, multi-biomarker approaches, employing a patho-biologically diverse set of biomarkers, could have a significant impact regarding the diagnosis and/or management of CVDs.

Table 1. Details of selected key established and emerging biomarkers for cardiovascular diseases (CVD). For myocardial infarction (MI), troponins (Tns) are the main biomarker used in current clinical practice for diagnosis, along with natriuretic peptides (NPs) for prognosis. Candidate biomarkers that provide additional information for prognosis in MI include Adrenomedullin (ADM), Heart-Type Fatty Acid-Binding Protein (H-FABP), Copeptin, P-selectin, Soluble Urokinase -type Plasminogen Activator Receptor (suPAR), Plasminogen Activator Inhibitor-1 (PAI-1), Galectin-3 (GAL-3), Matrix metalloproteinases (MMPs) and their tissue inhibitors (TIMPs), suppression of tumorigenicity 2 (ST2), Growth Differentiation Factor 15 (GDF-15), and Endothelin-1 (ET-1). Indeed, diagnosis and prognosis of coronary syndromes are currently guided by Tns, NPs, whilst P-selectin, suPAR, PAI-1, and GAL-3 could be additionally useful in the context of acute coronary syndromes in the future. Moreover, NPs levels are also used for diagnosis and prognosis of heart failure (HF), whilst Tns, FABP, Copeptin, ADM, MMPs, TIMPs, ST2, GDF-15, ET-1, and GAL-3 could also be helpful as biomarkers for HF screening and prognosis. Similarly, for diagnosis and prognosis of arrhythmias, specifically atrial fibrillation, a number of biomarkers, including Tns, NPs, H-FABP, GDF-15, and pro-inflammatory cytokines are described below. Regarding pro-inflammatory cytokines, interleukin-6 (IL-6), interleukin-8 (IL-8), tumor necrosis factor- α , and intercellular adhesion molecule-1 impact arrhythmic activity and could be prognosis biomarkers [466–469]. Indeed, increased levels of IL-6 and interleukin-10 were predictive of the risk of atrial and ventricular arrhythmias in patients hospitalized with COVID-19 [470]. Interestingly, NPs levels could be also assessed for prognosis of heart surgical procedures or congenital heart disease. Furthermore, tako-tsubo cardiomyopathy or acute pericarditis diagnosis can be aided by Tns levels, whilst measuring Tns, NPs, copeptin, H-FABP, MMPs, TIMPs, and ST2 may be helpful for the diagnosis and prognosis of aortic dissection, acute aortic syndrome, aortic stenosis, and other valvular diseases. Finally, circulating levels of Tns, NPs, Copeptin, and P-selectin are utilized for the diagnosis of stroke, while Tns H-and FABP are used for pulmonary embolism diagnosis and prognosis, respectively.

CVD Biomarker	Use	Present	Future	S	D	P	Ref.
Troponins (Tns)	Myocardial infarction	✓			✓	✓	[14,51,471,472]
	Coronary syndromes	✓			✓	✓	[473–477]
	Heart failure	✓(?)		✓		✓	[80,81,478–483]
	Atrial fibrillation	✓(?)			✓	✓	[484–487]
	Tako-tsubo cardiomyopathy	✓(?)			✓		[69–75]
	Aortic dissection, Aortic stenosis and other valvular diseases	✓(?)			✓	✓	[82,83]
	Acute pericarditis	✓(?)			✓		[84–86]
	Stroke	✓(?)			✓	✓	[486–488]
	Pulmonary embolism	✓(?)			✓		[90–92]

Table 1. Cont.

CVD Biomarker	Use	Present	Future	S	D	P	Ref.
Natriuretic Peptides (NPs)	Heart failure	✓		✓	✓	✓	[142,258,264,489–495]
	Coronary syndromes	✓				✓	[267,491,496,497]
	Myocardial infarction	✓				✓	[263,473,498–500]
	Atrial fibrillation		✓		✓	✓	[142,501,502]
	Stroke		✓		✓		[142,503]
	Surgical procedures involving the heart		✓			✓	[145]
	Pulmonary embolism		✓			✓	[166,167]
	Left ventricular hypertrophy		✓		✓	✓	[126,182,383]
	Valvular heart disease		✓			✓	[142,146,160]
Heart-Type Fatty Acid-Binding Protein (H-FABP)	Congenital heart disease		✓			✓	[122,146,182]
	Myocardial infarction		✓		✓ (?)	✓	[504–507]
	Heart failure		✓			✓	[213,214,508]
	Arrhythmia		✓			✓	[211,216]
	Valvular heart disease		✓		✓		[200]
	Pulmonary embolism		✓			✓	[91,218–220]
Copeptin	Myocardial infarction		✓		✓ (?)	✓	[226,227,232,509]
	Heart failure		✓			✓	[238,239,510]
	Stroke		✓		✓	✓	[511–513]
	Pulmonary embolism		✓	-	-	-	[91]
	Acute aortic syndrome		✓			✓	[244]
Adrenomedullin (ADM)	Myocardial infarction	✓		✓	✓	✓	[266,268,514]
	Heart failure	✓				✓	[258,264]
Ischemia Modified Albumin (IMA)	Unstable angina	✓			✓		[515–518]

Table 1. Cont.

CVD Biomarker	Use	Present	Future	S	D	P	Ref.
P-selectin	Myocardial infarction		✓	✓		✓	[275–279]
	Acute coronary syndrome		✓	✓		✓	[275,276,278]
	Stroke		✓	✓			[279]
Soluble Urokinase -type Plasminogen Activator Receptor (suPAR) and Plasminogen Activator Inhibitor-1 (PAI-1)	Myocardial infarction		✓			✓	[285–288]
	Acute coronary syndrome		✓			✓	[285–288]
Galectin-3 (GAL-3)	Myocardial infarction		✓			✓	[302]
	Heart failure		✓			✓	[313,314,519,520]
	Acute coronary syndrome		✓			✓	[301,302,313–315]
	acute myocarditis		✓			✓	[306,521]
Matrix metalloproteinases (MMPs) and their tissue inhibitors (TIMPs)	Myocardial infarction		✓	✓		✓	[364,522]
	Coronary artery stenosis		✓	✓		✓	[361,523]
	Heart failure		✓	✓		✓	[363]
	Multiple CVDs		✓	✓			[522]
Suppression of Tumorigenicity 2 (ST2)	Myocardial infarction		✓			✓	[524–527]
	Heart failure		✓			✓	[526–528]
	Aortic valve impairments		✓			✓	[529]
Growth Differentiation Factor 15 (GDF-15)	Myocardial infarction		✓				[373]
	Heart failure		✓	✓		✓	[375,390]
	Atrial fibrillation		✓	✓		✓	[395]
	Multiple CVDs		✓	✓			[7,378,380,385,386,390,391,396,398–400]
Endothelin-1 (ET-1)	Myocardial infarction		✓			✓	[267,429,430]
	Heart failure		✓	✓	✓	✓	[267,429,430]
	Multiple CVDs		✓	✓			[307,411,413,414,423,424]

Table 1. *Cont.*

CVD Biomarker	Use	Present	Future	S	D	P	Ref.
Cytokines	Atrial fibrillation (interleukin-6, tumor necrosis factor- α and intercellular adhesion molecule-1)		✓		✓	✓	[467,468]
	Multiple CVDs		✓			✓	[530–533]
Lipoprotein-Associated Phospholipase A2 (Lp-PLA2)	Multiple CVDs		✓			✓	[447,448,534]
Soluble CD40 Ligand	Multiple CVDs		✓			✓	[453,535,536]
Serum Amyloid A	Multiple CVDs		✓			✓	[449,537]
Osteoprotegerin (OPG)	Multiple CVDs		✓			✓	[307,434,461,462]
Myeloperoxidase	Multiple CVDs		✓			✓	[63,359,504]
C-reactive protein (CRP)	Multiple CVDs		✓	✓	✓	✓	[467,533,537–548]
Erythrocyte sedimentation rate (ESR)	Multiple CVDs		✓			✓	[549]
Neutrophils and monocytes	Multiple CVDs		✓			✓	[463]
Cystatin C	Multiple CVDs		✓			✓	[550–552]

S: Screening, D: Diagnosis, P: Prognosis. (?) = biomarker is recommended to use for this purpose in some reports.

Author Contributions: Conceptualization K.C. and F.O. (Farah Omran); Writing—Original Draft Preparation, F.O. (Farah Omran); Writing—Review & Editing, F.O. (Farah Omran), I.K., F.O. (Faizel Osman), V.G.L., H.S.R. and K.C.; Visualization, F.O. (Farah Omran), I.K., F.O. (Faizal Osman), H.S.R. and K.C.; Supervision, K.C. All authors listed have made a substantial, direct and intellectual contribution to the work. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. World Health Organization (WHO) Cardiovascular Diseases (CVDs). 2021. Available online: [https://www.who.int/news-room/fact-sheets/detail/cardiovascular-diseases-\(cvds\)](https://www.who.int/news-room/fact-sheets/detail/cardiovascular-diseases-(cvds)) (accessed on 11 June 2021).
2. Behera, S.; Pramanik, K.; Nayak, M. Recent Advancement in the Treatment of Cardiovascular Diseases: Conventional Therapy to Nanotechnology. *Curr. Pharm. Des.* **2015**, *21*, 4479–4497. [CrossRef] [PubMed]
3. Aronson, J.K.; Ferner, R.E. Biomarkers—A general review. *Curr. Protoc. Pharmacol.* **2017**, *2017*, 9.23.1–9.23.17. [CrossRef] [PubMed]
4. Biomarkers Definitions Working Group. Biomarkers and surrogate endpoints: Preferred definitions and conceptual framework. *Clin. Pharmacol. Ther.* **2001**, *69*, 89–95. [CrossRef] [PubMed]
5. Strimbu, K.; Tavel, J.A. What are biomarkers? *Curr. Opin. HIV AIDS* **2010**, *5*, 463–466. [CrossRef]
6. Smith, J.J.; Sorensen, A.G.; Thrall, J.H. Biomarkers in imaging: Realizing radiology’s future. *Radiology* **2003**, *227*, 633–638. [CrossRef]
7. Hlatky, M.A.; Greenland, P.; Arnett, D.K.; Ballantyne, C.M.; Criqui, M.H.; Elkind, M.S.V.; Go, A.S.; Harrell, F.E., Jr.; Hong, Y.; Howard, B.V.; et al. Criteria for evaluation of novel markers of cardiovascular risk: A scientific statement from the American heart association. *Circulation* **2009**, *119*, 2408–2416. [CrossRef]
8. Danese, E.; Montagnana, M. An historical approach to the diagnostic biomarkers of acute coronary syndrome. *Ann. Transl. Med.* **2016**, *4*, 194. [CrossRef]
9. Aydin, S.; Ugur, K.; Aydin, S.; Sahin, İ.; Yardim, M. Biomarkers in acute myocardial infarction: Current perspectives. *Vasc. Health Risk Manag.* **2019**, *15*, 1–10. [CrossRef]
10. McLeish, M.J.; Kenyon, G.L. Relating structure to mechanism in creatine kinase. *Crit. Rev. Biochem. Mol. Biol.* **2005**, *40*, 1–20. [CrossRef]
11. Kemp, M.; Donovan, J.; Higham, H.; Hooper, J. Biochemical markers of myocardial injury. *Br. J. Anaesth.* **2004**, *93*, 63–73. [CrossRef]
12. Kurapati, R.; Soos, M. “CPK-MB”. In *StatPearls*; StatPearls Publishing: Treasure Island, FL, USA, 2020.
13. Dolci, A.; Panteghini, M. The exciting story of cardiac biomarkers: From retrospective detection to gold diagnostic standard for acute myocardial infarction and more. *Clin. Chim. Acta* **2006**, *369*, 179–187. [CrossRef] [PubMed]
14. Thygesen, K.; Alpert, J.S.; Jaffe, A.S.; Chaitman, B.R.; Bax, J.J.; Morrow, D.A.; White, H.D. Fourth Universal Definition of Myocardial Infarction (2018). *J. Am. Coll. Cardiol.* **2018**, *72*, 2231–2264. [CrossRef] [PubMed]
15. Goodman, S.G.; Steg, P.G.; Eagle, K.A.; Fox, K.A.A.; López-Sendón, J.; Montalescot, G.; Budaj, A.; Kennelly, B.M.; Gore, J.M.; Allegrone, J.; et al. The diagnostic and prognostic impact of the redefinition of acute myocardial infarction: Lessons from the Global Registry of Acute Coronary Events (GRACE). *Am. Heart J.* **2006**, *151*, 654–660. [CrossRef] [PubMed]
16. The Joint European Society of Cardiology/ American College of Cardiology Committee. Myocardial infarction redefined—A consensus document of The Joint European Society of Cardiology/American College of Cardiology Committee for the redefinition of myocardial infarction. *J. Am. Coll. Cardiol.* **2000**, *36*, 959–969. [CrossRef]
17. Morrow, D.A.; Cannon, C.P.; Jesse, R.L.; Newby, L.K.; Rakvilde, J.; Storrow, A.B.; Wu, A.H.; Christenson, R.H. National Academy of Clinical Biochemistry Laboratory Medicine Practice Guidelines: Clinical characteristics and utilization of biochemical markers in acute coronary syndromes. *Circulation* **2007**, *115*, 356–375.
18. Christenson, R.H.; Vaidya, H.; Landt, Y.; Bauer, R.S.; Green, S.F.; Apple, F.A.; Jacob, A.; Magneson, G.R.; Nag, S.; Wu, A.H.; et al. Standardization of Creatine Kinase-MB (CK-MB) Mass Assays: The Use of Recombinant CK-MB as a Reference Material. *Clin. Chem.* **1999**, *45*, 1414–1423. [CrossRef]
19. Lauer, B.; Niederau, C.; Kühl, U.; Schannwell, M.; Pauschinger, M.; Strauer, B.E.; Schultheiss, H.P. Cardiac troponin T in patients with clinically suspected myocarditis. *J. Am. Coll. Cardiol.* **1997**, *30*, 1354–1359. [CrossRef]
20. Collinson, P.O.; Garrison, L.; Christenson, R.H. Cardiac biomarkers—A short biography. *Clin. Biochem.* **2015**, *48*, 197–200.
21. Singh, G.; Baweja, P.S. Creatine Kinase-MB. *Am. J. Clin. Pathol.* **2014**, *141*, 415–419. [CrossRef]
22. Ruppert, M.; Van Hee, R. Creatinine-kinase-MB determination in non-cardiac trauma: Its difference with cardiac infarction and its restricted use in trauma situations. *Eur. J. Emerg. Med.* **2001**, *8*, 177–179. [CrossRef]
23. Ladowski, J.S.; Sullivan, M.; Schatzlein, M.H.; Peterson, A.; Underhill, D.J.; Scheeringa, R.H. Cardiac isoenzymes following heart transplantation. *Chest* **1992**, *102*, 1520–1521. [CrossRef] [PubMed]
24. de Iuliis, F.; Salerno, G.; Taglieri, L.; De Biase, L.; Lanza, R.; Cardelli, P.; Scarpa, S. Serum biomarkers evaluation to predict chemotherapy-induced cardiotoxicity in breast cancer patients. *Tumor Biol.* **2016**, *37*, 3379–3387. [CrossRef] [PubMed]

25. Jo, M.S.; Lee, J.; Kim, S.Y.; Kwon, H.J.; Lee, H.K.; Park, D.J.; Kim, Y. Comparison between creatine kinase MB, heart-type fatty acid-binding protein, and cardiac troponin T for detecting myocardial ischemic injury after cardiac surgery. *Clin. Chim. Acta* **2019**, *488*, 174–178. [[CrossRef](#)] [[PubMed](#)]
26. Adams, J.E.; Siegel, B.A.; Goldstein, J.A.; Jaffe, A.S. Elevations of CK-MB following pulmonary embolism; A manifestation of occult right ventricular infarction. *Chest* **1992**, *101*, 1203–1206. [[CrossRef](#)]
27. Bozbay, M.; Uyarel, H.; Avsar, S.; Oz, A.; Keskin, M.; Tanik, V.O.; Bakhshaliyev, N.; Ugur, M.; Pehlivanoglu, S.; Eren, M. Creatinine kinase isoenzyme-MB: A simple prognostic biomarker in patients with pulmonary embolism treated with thrombolytic therapy. *J. Crit. Care* **2015**, *30*, 1179–1183. [[CrossRef](#)]
28. Wang, S.; Qin, L.; Wu, T.; Deng, B.; Sun, Y.; Hu, D.; Mohan, C.; Zhou, X.J.; Peng, A. Elevated cardiac markers in chronic kidney disease as a consequence of hyperphosphatemia-induced cardiac myocyte injury. *Med. Sci. Monit.* **2014**, *20*, 2043–2053.
29. Matsumura, T.; Saito, T.; Fujimura, H.; Shinno, S. Cardiac troponin I for accurate evaluation of cardiac status in myopathic patients. *Brain Dev.* **2007**, *29*, 496–501. [[CrossRef](#)]
30. Volochayev, R. Laboratory Test Abnormalities are Common in Polymyositis and Dermatomyositis and Differ Among Clinical and Demographic Groups. *Open Rheumatol. J.* **2012**, *6*, 54–63. [[CrossRef](#)]
31. Benoist, J.F.; Cosson, C.; Mimoz, O.; Edouard, A. Serum cardiac troponin I, creatine kinase (CK), and CK-MB in early posttraumatic rhabdomyolysis. *Clin. Chem.* **1997**, *43*, 416–417. [[CrossRef](#)]
32. Osborn, L.A.; Rossum, A.; Standefer, J.; Jackson, J.; Skipper, B.; Beeson, C.; Crawford, M.H. Evaluation of CK and CK-MB in Alcohol Abuse Subjects with Recent Heavy Consumption. *Cardiology* **1995**, *86*, 130–134. [[CrossRef](#)]
33. Nguyen, H.T.; Nguyen, C.T.H. Cardiac amyloidosis mimicking acute coronary syndrome: A case report and literature review. *Eur. Heart J. Case Rep.* **2020**, *4*, 1–7. [[CrossRef](#)] [[PubMed](#)]
34. Newby, L.K.; Roe, M.T.; Chen, A.Y.; Ohman, E.M.; Christenson, R.H.; Pollack, C.V.; Hoekstra, J.W.; Peacock, W.F.; Harrington, R.A.; Jesse, R.L.; et al. Frequency and clinical implications of discordant creatine kinase-MB and troponin measurements in acute coronary syndromes. *J. Am. Coll. Cardiol.* **2006**, *47*, 312–318. [[CrossRef](#)] [[PubMed](#)]
35. Marín-García, J. Cardiomyopathies: A Comparative Analysis of Phenotypes and Genotypes. In *Post-Genomic Cardiology*, 2nd ed.; Elsevier: Amsterdam, The Netherlands, 2014; pp. 363–426.
36. Li, M.X.; Hwang, P.M. Structure and function of cardiac troponin C (TNNC1): Implications for heart failure, cardiomyopathies, and troponin modulating drugs. *Gene* **2015**, *571*, 153–166. [[CrossRef](#)] [[PubMed](#)]
37. Dellow, K.A.; Bhavsar, P.K.; Brand, N.J.; Barton, P.J.R. Identification of novel, cardiac-restricted transcription factors binding to a CACC-box within the human cardiac troponin I promoter. *Cardiovasc. Res.* **2001**, *50*, 24–33. [[CrossRef](#)]
38. Bhavsar, P.K.; Dellow, K.A.; Yacoub, M.H.; Brand, N.J.; Barton, P.J.R. Identification of cis-acting DNA elements required for expression of the human cardiac troponin I gene promoter. *J. Mol. Cell. Cardiol.* **2000**, *32*, 95–108. [[CrossRef](#)] [[PubMed](#)]
39. Anderson, P.A.W.; Malouf, N.N.; Oakeley, A.E.; Pagani, E.D.; Allen, P.D. Troponin T Isoform Expression in Humans. *Circ. Res.* **1991**, *69*, 1226–1233. [[CrossRef](#)]
40. Karlén, J.; Karlsson, M.; Eliasson, H.; Bonamy, A.K.E.; Halvorsen, C.P. Cardiac Troponin T in Healthy Full-Term Infants. *Pediatr. Cardiol.* **2019**, *40*, 1645–1654. [[CrossRef](#)]
41. Sasse, S.; Brand, N.J.; Kyriyanou, P.; Dhoot, G.K.; Wade, R.; Arai, M.; Periasamy, M.; Yacoub, M.H.; Barton, P.J. Troponin I gene expression during human cardiac development and in end—Stage heart failure. *Circ. Res.* **1993**, *72*, 932–938. [[CrossRef](#)]
42. Anderson, P.A.W.; Oakeley, A.E. Immunological identification of five troponin T isoforms reveals an elaborate maturational troponin T profile in rabbit myocardium. *Circ. Res.* **1989**, *65*, 1087–1093. [[CrossRef](#)]
43. Mesnard, L.; Logeart, D.; Taviaux, S.; Diriong, S.; Mercadier, J.-J.; Samson, F. Human Cardiac Troponin T: Cloning and Expression of New Isoforms in the Normal and Failing Heart. *Circ. Res.* **1995**, *76*, 687–692. [[CrossRef](#)]
44. Barton, P.J.R.; Felkin, L.E.; Koban, M.U.; Cullen, M.E.; Brand, N.J.; Dhoot, G.K. The slow skeletal muscle troponin T gene is expressed in developing and diseased human heart. *Mol. Cell. Biochem.* **2004**, *263*, 91–97. [[CrossRef](#)] [[PubMed](#)]
45. Samson, F.; Mesnard, L.; Mihovilovic, M.; Potter, T.G.; Mercadier, J.J.; Roses, A.D.; Gilbert, J.R. A New Human Slow Skeletal Troponin T (TnTs) mRNA Isoform Derived from Alternative Splicing of a Single Gene. *Biochem. Biophys. Res. Commun.* **1994**, *199*, 841–847. [[CrossRef](#)] [[PubMed](#)]
46. Thygesen, K.; Alpert, J.S.; Jaffe, A.S.; Simoons, M.L.; Chaitman, B.R.; White, H.D. Third universal definition of myocardial infarction. *J. Am. Coll. Cardiol.* **2012**, *60*, 1581–1598. [[CrossRef](#)] [[PubMed](#)]
47. Bagai, A.; Alexander, K.P.; Berger, J.S.; Senior, R.; Sajeev, C.; Pracon, R.; Mavromatis, K.; Lopez-Sendón, J.L.; Gosselin, G.; Diaz, A.; et al. Use of troponin assay 99th percentile as the decision level for myocardial infarction diagnosis. *Am. Heart J.* **2017**, *190*, 135–139. [[CrossRef](#)] [[PubMed](#)]
48. Giannitsis, E.; Kurz, K.; Hallermayer, K.; Jarausch, J.; Jaffe, A.S.; Katus, H.A. Analytical validation of a high-sensitivity cardiac troponin T assay. *Clin. Chem.* **2010**, *56*, 254–261. [[CrossRef](#)]
49. Melanson, S.E.F.; Morrow, D.A.; Jarolim, P. Earlier detection of myocardial injury in a preliminary evaluation using a new troponin I assay with improved sensitivity. *Am. J. Clin. Pathol.* **2007**, *128*, 282–286. [[CrossRef](#)]
50. Mueller, C.; Twerenbold, R.; Reichlin, T. Early diagnosis of myocardial infarction with sensitive cardiac troponin assays. *Clin. Chem.* **2019**, *65*, 490–491. [[CrossRef](#)]
51. Keller, T.; Zeller, T.; Peetz, D.; Tzikas, S.; Roth, A.; Czyz, E.; Bickel, C.; Baldus, S.; Warnholtz, A.; Fröhlich, M. Sensitive Troponin I Assay in Early Diagnosis of Acute Myocardial Infarction. *N. Engl. J. Med.* **2009**, *361*, 868–877. [[CrossRef](#)]

52. James, S.; Armstrong, P.; Califf, R.; Simoons, M.L.; Venge, P.; Wallentin, L.; Lindahl, B. Troponin T levels and risk of 30-day outcomes in patients with the acute coronary syndrome: Prospective verification in the GUSTO-IV trial. *Am. J. Med.* **2003**, *115*, 178–184. [CrossRef]
53. Venge, P.; Lagerqvist, B.O.; Diderholm, E.; Lindahl, B.; Wallentin, L. Clinical performance of three cardiac troponin assays in patients with unstable coronary artery disease (a FRISC II substudy). *Am. J. Cardiol.* **2002**, *89*, 1035–1041. [CrossRef]
54. Morrow, D.A.; Rifai, N.; Sabatine, M.S.; Ayanian, S.; Murphy, S.A.; De Lemos, J.A.; Braunwald, E.; Cannon, C.P. Evaluation of the AccuTnI cardiac troponin I assay for risk assessment in acute coronary syndromes. *Clin. Chem.* **2003**, *49*, 1396–1398. [CrossRef] [PubMed]
55. Morrow, D.A.; Cannon, C.P.; Rifai, N.; Frey, M.J.; Vicari, R.; Lakkis, N.; Robertson, D.H.; Hille, D.A.; DeLuca, P.T.; DiBattiste, P.M.; et al. Ability of minor elevations of troponins I and T to predict benefit from an early invasive strategy in patients with unstable angina and non-ST elevation myocardial infarction: Results from a randomized trial. *J. Am. Med. Assoc.* **2001**, *286*, 2405–2412. [CrossRef] [PubMed]
56. Doust, J.; Glasziou, P. High-Sensitivity Troponin Highlights the Need for New Methods to Evaluate Diagnostic Tests. *Circ. Cardiovasc. Qual. Outcomes* **2018**, *11*, 9–12. [CrossRef] [PubMed]
57. Aw, T.C.; Huang, W.T.; Le, T.T.; Pua, C.J.; Ang, B.; Phua, S.K.; Yeo, K.K.; Cook, S.A.; Chin, C.W. High-Sensitivity cardiac Troponins in Cardio-Healthy Subjects: A Cardiovascular Magnetic Resonance Imaging Study. *Sci. Rep.* **2018**, *8*, 15409. [CrossRef]
58. Sabatine, M.S.; Morrow, D.A.; de Lemos, J.A.; Jarolim, P.; Braunwald, E. Detection of acute changes in circulating troponin in the setting of transient stress test-induced myocardial ischaemia using an ultrasensitive assay: Results from TIMI 35. *Eur. Heart J.* **2009**, *30*, 162–169. [CrossRef]
59. Gresslien, T.; Agewall, S. Troponin and exercise. *Int. J. Cardiol.* **2016**, *221*, 609–621. [CrossRef]
60. Thygesen, K.; Mair, J.; Katus, H.; Plebani, M.; Venge, P.; Collinson, P.; Lindahl, B.; Giannitsis, E.; Hasin, Y.; Galvani, M.; et al. Recommendations for the use of cardiac troponin measurement in acute cardiac care. *Eur. Heart J.* **2010**, *31*, 2197–2204. [CrossRef]
61. Garg, P.; Morris, P.; Fazlanie, A.L.; Vijayan, S.; Dancso, B.; Dastidar, A.G.; Plein, S.; Mueller, C.; Haaf, P. Cardiac biomarkers of acute coronary syndrome: From history to high-sensitivity cardiac troponin. *Intern. Emerg. Med.* **2017**, *12*, 147–155. [CrossRef]
62. Park, K.C.; Gaze, D.C.; Collinson, P.O.; Marber, M.S. Cardiac troponins: From myocardial infarction to chronic disease. *Cardiovasc. Res.* **2017**, *113*, 1708–1718. [CrossRef]
63. Savic-Radojevic, A.; Pljesa-Ercegovac, M.; Matic, M.; Simic, D.; Radovanovic, S.; Simic, T. *Novel Biomarkers of Heart Failure*, 1st ed.; Elsevier Inc.: Amsterdam, The Netherlands, 2017; Volume 79.
64. Katus, H.A.; Remppis, A.; Scheffold, T.; Diederich, K.W.; Kuebler, W. Intracellular compartmentation of cardiac troponin T and its release kinetics in patients with reperfused and nonreperfused myocardial infarction. *Am. J. Cardiol.* **1991**, *67*, 1360–1367. [CrossRef]
65. Hallén, J. Troponin for the estimation of infarct size: What have we learned? *Cardiology* **2012**, *121*, 204–212. [CrossRef] [PubMed]
66. Giannitsis, E.; Steen, H.; Kurz, K.; Ivandic, B.; Simon, A.C.; Futterer, S.; Schild, C.; Isfort, P.; Jaffe, A.S.; Katus, H.A. Cardiac Magnetic Resonance Imaging Study for Quantification of Infarct Size Comparing Directly Serial VS. Single Time-Point Measurements of Cardiac Troponin T. *J. Am. Coll. Cardiol.* **2008**, *51*, 307–314. [CrossRef] [PubMed]
67. Rahman, A.; Broadley, S.A. Review article: Elevated troponin: Diagnostic gold or fool's gold? *EMA Emerg. Med. Australas.* **2014**, *26*, 125–130. [CrossRef] [PubMed]
68. Hickman, P.E.; Potter, J.M.; Aroney, C.; Koerbin, G.; Southcott, E.; Wu, A.H.B.; Roberts, M.S. Cardiac troponin may be released by ischemia alone, without necrosis. *Clin. Chim. Acta* **2010**, *411*, 318–323. [CrossRef] [PubMed]
69. Ramaraj, R.; Sorrell, V.L.; Movahed, M.R. Levels of troponin release can aid in the early exclusion of stress-induced (takotsubo) cardiomyopathy. *Exp. Clin. Cardiol.* **2009**, *14*, 6–8.
70. Novo, G.; Giambanco, S.; Bonomo, V.; Sutera, M.R.; Giambanco, F.; Rotolo, A.; Evola, S.; Assennato, P.; Novo, S. Troponin I/ejection fraction ratio: A new index to differentiate Takotsubo cardiomyopathy from myocardial infarction. *Int. J. Cardiol.* **2015**, *180*, 255–257. [CrossRef]
71. Randhawa, M.S.; Dhillon, A.S.; Taylor, H.C.; Sun, Z.; Desai, M.Y. Diagnostic utility of cardiac biomarkers in discriminating takotsubo cardiomyopathy from acute myocardial infarction. *J. Card. Fail.* **2014**, *20*, 2–8. [CrossRef]
72. Burgdorf, C.; Schubert, A.; Schunkert, H.; Kurowski, V.; Radke, P.W. Release patterns of copeptin and troponin in Tako-Tsubo cardiomyopathy. *Peptides* **2012**, *34*, 389–394. [CrossRef]
73. Sharkey, S.W.; Lesser, J.R.; Menon, M.; Parpart, M.; Maron, M.S.; Maron, B.J. Spectrum and Significance of Electrocardiographic Patterns, Troponin Levels, and Thrombolysis in Myocardial Infarction Frame Count in Patients with Stress (Tako-tsubo) Cardiomyopathy and Comparison to Those in Patients with ST-Elevation Anterior Wall Myoc. *Am. J. Cardiol.* **2008**, *101*, 1723–1728. [CrossRef]
74. Nef, H.M.; Möllmann, H.; Kostin, S.; Troidl, C.; Voss, S.; Weber, M.; Dill, T.; Rolf, A.; Brandt, R.; Hamm, C.W.; et al. Tako-Tsubo cardiomyopathy: Intraindividual structural analysis in the acute phase and after functional recovery. *Eur. Heart J.* **2007**, *28*, 2456–2464. [CrossRef]
75. Looi, J.L.; Wong, C.W.; Khan, A.; Webster, M.; Kerr, A.J. Clinical Characteristics and Outcome of Apical Ballooning Syndrome in Auckland, New Zealand. *Heart Lung Circ.* **2012**, *21*, 143–149. [CrossRef] [PubMed]
76. Feng, J.; Schaus, B.J.; Fallavollita, J.A.; Lee, T.-C.; Carty, J.M. Preload Induces Troponin I Degradation Independently of Myocardial Ischemia. *Circulation* **2001**, *103*, 2035–2037. [CrossRef] [PubMed]

77. Parissis, J.T.; Ikonomidou, I.; Rafouli-Stergiou, P.; Mebazaa, A.; Delgado, J.; Farmakis, D.; Vilas-Boas, F.; Paraskevaidis, I.; Anastasiou-Nana, M.; Follath, F. Clinical characteristics and predictors of in-hospital mortality in acute heart failure with preserved left ventricular ejection fraction. *Am. J. Cardiol.* **2011**, *107*, 79–84. [CrossRef] [PubMed]
78. Santhanakrishnan, R.; Chong, J.P.C.; Ng, T.P.; Ling, L.H.; Sim, D.; Toh GLeong, K.; Shuan, D.; Yeo, P.; Ong, H.Y. Growth differentiation factor 15, ST2, high-sensitivity troponin T, and N-terminal pro brain natriuretic peptide in heart failure with preserved vs. reduced ejection fraction. *Eur. J. Heart Fail.* **2012**, *14*, 1338–1347. [CrossRef] [PubMed]
79. Kociol, R.D.; Pang, P.S.; Gheorghiade, M.; Fonarow, G.C.; O'Connor, C.M.; Felker, G.M. Troponin elevation in heart failure: Prevalence, mechanisms, and clinical implications. *J. Am. Coll. Cardiol.* **2010**, *56*, 1071–1078. [CrossRef] [PubMed]
80. Peacock, W.F.; De Marco, T.; Fonarow, G.C.; Diercks, D.; Wynne, J.; Apple, F.S.; Wu, A.H. Cardiac Troponin and Outcome in Acute Heart Failure. *N. Engl. J. Med.* **2008**, *358*, 2117–2126. [CrossRef] [PubMed]
81. Latini, R.; Masson, S.; Anand, I.S.; Missov, E.; Carlson, M.; Vago, T.; Angelici, L.; Barlera, S.; Parrinello, G.; Maggioni, A.P.; et al. Prognostic value of very low plasma concentrations of troponin T in patients with stable chronic heart failure. *Circulation* **2007**, *116*, 1242–1249. [CrossRef]
82. Nunes, J.P.L. Elevated troponin and aortic valve disease. *J. Am. Coll. Cardiol.* **2013**, *61*, 1467. [CrossRef]
83. Martin, A.; Malhotra, A.; Sullivan, B.; Ramakrishna, H. Troponin elevations in patients with chronic cardiovascular disease: An analysis of current evidence and significance. *Ann. Card. Anaesth.* **2016**, *19*, 321–327.
84. Brandt, R.R.; Filzmaier, K.; Hanrath, P. Circulating cardiac troponin I in acute pericarditis. *Am. J. Cardiol.* **2001**, *87*, 1326–1328. [CrossRef]
85. Imazio, M. Pericarditis with troponin elevation: Is it true pericarditis and a reason for concern? *J. Cardiovasc. Med.* **2014**, *15*, 73–77. [CrossRef] [PubMed]
86. McNamara, N.; Ibrahim, A.; Satti, Z.; Ibrahim, M.; Kiernan, T.J. Acute pericarditis: A review of current diagnostic and management guidelines. *Future Cardiol.* **2019**, *15*, 119–126. [CrossRef] [PubMed]
87. Sandhu, R.; Aronow, W.S.; Rajdev, A.; Sukhija, R.; Amin, H.; D'aquila, K.; Sangha, A. Relation of Cardiac Troponin I Levels with In-Hospital Mortality in Patients with Ischemic Stroke, Intracerebral Hemorrhage, and Subarachnoid Hemorrhage. *Am. J. Cardiol.* **2008**, *102*, 632–634. [CrossRef] [PubMed]
88. Kerr, G.; Ray, G.; Wu, O.; Stott, D.J.; Langhorne, P. Elevated troponin after stroke: A systematic review. *Cerebrovasc. Dis.* **2009**, *28*, 220–226. [CrossRef]
89. Jespersen, C.M.; Hansen, J.F. Myocardial stress in patients with acute cerebrovascular events. *Cardiology* **2008**, *110*, 123–128. [CrossRef]
90. Coma-canella, I.; Gamallo, C.; Onsurbe, P.M.; Lopez-sendon, J. Acute right ventricular infarction secondary to massive pulmonary embolism. *Eur. Heart J.* **1988**, *9*, 534–540. [CrossRef]
91. Giannitsis, E.; Katus, H.A. Biomarkers for clinical decision-making in the management of pulmonary embolism. *Clin. Chem.* **2017**, *63*, 91–100. [CrossRef]
92. Müller-Bardorff, M.; Weidtmann, B.; Giannitsis, E.; Kurowski, V.; Katus, H.A. Release kinetics of cardiac troponin T in survivors of confirmed severe pulmonary embolism. *Clin. Chem.* **2002**, *48*, 673–675. [CrossRef]
93. de Lemos, J.A.; Drazner, M.H.; Omland, T.; Ayers, C.R.; Khera, A.; Rohatgi, A.; Hashim, I.; Berry, J.D.; Das, S.R.; Morrow, D.A.; et al. Association of Troponin T Detected with a Highly Sensitive Assay and Cardiac Structure and Mortality Risk in the General Population. *JAMA* **2010**, *304*, 2503. [CrossRef]
94. Beatty, A.L.; Ku, I.A.; Christenson, R.H.; DeFilippi, C.R.; Schiller, N.B.; Whooley, M.A. High-Sensitivity Cardiac Troponin T Levels and Secondary Events in Outpatients with Coronary Heart Disease From the Heart and Soul Study. *JAMA Intern. Med.* **2013**, *173*, 763. [CrossRef]
95. Sato, Y.; Yamamoto, E.; Sawa, T.; Toda, K.; Hara, T.; Iwasaki, T.; Fujiwara, H.; Takatsu, Y. High-sensitivity cardiac troponin T in essential hypertension. *J. Cardiol.* **2011**, *58*, 226–231. [CrossRef] [PubMed]
96. Uçar, H.; Gür, M.; Kivrak, A.; Koyunsever, N.Y.; Şeker, T.; Akilli, R.E.; Türkoğlu, C.; Kaypaklı, O.; Şahin, D.Y.; Elbasan, Z.; et al. High-sensitivity cardiac troponin T levels in newly diagnosed hypertensive patients with different left ventricle geometry. *Blood Press.* **2014**, *23*, 240–247. [CrossRef] [PubMed]
97. McEvoy, J.W.; Chen, Y.; Nambi, V.; Ballantyne, C.M.; Sharrett, A.R.; Appel, L.J.; Post, W.S.; Blumenthal, R.S.; Matsushita, K.; Selvin, E. High-sensitivity cardiac Troponin T and risk of hypertension. *Circulation* **2015**, *132*, 825–833. [CrossRef] [PubMed]
98. Giannitsis, E.; Katus, H.A. Cardiac troponin level elevations not related to acute coronary syndromes. *Nat. Rev. Cardiol.* **2013**, *10*, 623–634. [CrossRef]
99. Agewall, S.; Giannitsis, E.; Jernberg, T.; Katus, H. Troponin elevation in coronary vs. non-coronary disease. *Eur. Heart J.* **2011**, *32*, 404–411. [CrossRef]
100. Klinkenberg, L.J.J. High-sensitivity cardiac troponins in health and disease. *Ned. Tijdschr. Voor Klin. Chemie Lab.* **2016**, *41*, 235–241.
101. Bais, R. The Effect of Sample Hemolysis on Cardiac Troponin, I. and T. Assays. *Clin. Chem.* **2010**, *56*, 1357–1359. [CrossRef]
102. Florkowski, C.; Wallace, J.; Walmsley, T.; George, P. The Effect of Hemolysis on Current Troponin Assays—A Confounding Preanalytical Variable? *Clin. Chem.* **2010**, *56*, 1195–1197. [CrossRef]
103. Panteghini, M. Assay-related issues in the measurement of cardiac troponins. *Clin. Chim. Acta* **2009**, *402*, 88–93. [CrossRef]

104. Göser, S.; Andrassy, M.; Buss, S.J.; Leuschner, F.; Volz, C.H.; Öttl, R.; Zitrich, S.; Blaudeck, N.; Hardt, S.E.; Pfitzer, G.; et al. Cardiac troponin I but not cardiac troponin T induces severe autoimmune inflammation in the myocardium. *Circulation* **2006**, *114*, 1693–1702. [[CrossRef](#)]
105. Daniels, L.B.; Maisel, A.S. Natriuretic Peptides. *J. Am. Coll. Cardiol.* **2007**, *50*, 2357–2368. [[CrossRef](#)] [[PubMed](#)]
106. Gupta, D.K.; Wang, T.J. Natriuretic Peptides and Cardiometabolic Health. *Circ. J.* **2015**, *79*, 1647–1655. [[CrossRef](#)] [[PubMed](#)]
107. Pemberton, C.J.; Charles, C.J.; Richards, A.M. Cardiac Natriuretic Peptides. In *Endocrinology of the Heart in Health and Disease: Integrated, Cellular, and Molecular Endocrinology of the Heart*; Schisler, J.C., Lang, C.H., Willis, M.S., Eds.; Academic Press: Cambridge, MA, USA, 2016; pp. 1–354.
108. del Ry, S.; Cabiati, M.; Clerico, A. Natriuretic peptide system and the heart. *Cardiovasc. Issues Endocrinol.* **2014**, *43*, 134–143.
109. Potter, L.R.; Yoder, A.R.; Flora, D.R.; Antos, L.K.; Dickey, D.M. Natriuretic peptides: Their structures, receptors, physiologic functions and therapeutic applications. *Handb. Exp. Pharmacol.* **2009**, *191*, 341–366.
110. LaPointe, M.C. Molecular regulation of the brain natriuretic peptide gene. *Peptides* **2005**, *26*, 944–956. [[CrossRef](#)] [[PubMed](#)]
111. Houweling, A.C.; van Borren, M.M.; Moorman, A.F.M.; Christoffels, V.M. Expression and regulation of the atrial natriuretic factor encoding gene Nppa during development and disease. *Cardiovasc. Res.* **2005**, *67*, 583–593. [[CrossRef](#)]
112. Kalra, P.R.; Clague, J.R.; Bolger, A.P.; Anker, S.D.; Poole-Wilson, P.A.; Struthers, A.D.; Coats, A.J. Myocardial production of C-type natriuretic peptide in chronic heart failure. *Circulation* **2003**, *107*, 571–573. [[CrossRef](#)] [[PubMed](#)]
113. Hama, N.; Itoh, H.; Shirakami, G.; Nakagawa, O.; Suga, S.; Ogawa, Y.; Masuda, I.; Nakanishi, K.; Yoshimasa, T.; Hashimoto, Y. Rapid Ventricular Induction of Brain Natriuretic Peptide Gene Expression in Experimental Acute Myocardial Infarction. *Circulation* **1995**, *92*, 1558–1564. [[CrossRef](#)]
114. Nakagawa, O.; Ogawa, Y.; Itoh, H.; Suga, S.I.; Komatsu, Y.; Kishimoto, I.; Nishino, K.; Yoshimasa, T.; Nakao, K. Rapid transcriptional activation and early mRNA turnover of brain natriuretic peptide in cardiocyte hypertrophy: Evidence for brain natriuretic peptide as an ‘emergency’ cardiac hormone against ventricular overload. *J. Clin. Investig.* **1995**, *96*, 1280–1287. [[CrossRef](#)]
115. Wang, D.; Oparil, S.; Feng, J.A.; Li, P.; Perry, G.; Chen, L.B.; Dai, M.; John, S.W.; Chen, Y.F. Effects of pressure overload on extracellular matrix expression in the heart of the atrial natriuretic peptide-null mouse. *Hypertension* **2003**, *42*, 88–95. [[CrossRef](#)]
116. Cao, Z.; Jia, Y.; Zhu, B. BNP and NT-proBNP as diagnostic biomarkers for cardiac dysfunction in both clinical and forensic medicine. *Int. J. Mol. Sci.* **2019**, *20*, 1820. [[CrossRef](#)] [[PubMed](#)]
117. Morgenthaler, N.G.; Struck, J.; Thomas, B.; Bergmann, A. Immunoluminometric Assay for the Midregion of Pro-Atrial Natriuretic Peptide in Human Plasma. *Clin. Chem.* **2004**, *50*, 234–236. [[CrossRef](#)] [[PubMed](#)]
118. Morgenthaler, N.G.; Struck, J.; Christ-Crain, M.; Bergmann, A.; Müller, B. Pro-atrial natriuretic peptide is a prognostic marker in sepsis, similar to the APACHE II score: An observational study. *Crit. Care* **2004**, *9*, R37. [[CrossRef](#)] [[PubMed](#)]
119. Vesely, D.L. Atrial natriuretic peptide prohormone gene expression: Hormones and diseases that upregulate its expression. *IUBMB Life* **2002**, *53*, 153–159. [[CrossRef](#)]
120. Lee, C.S.; Tkacs, N.C. Current concepts of neurohormonal activation in heart failure: Mediators and mechanisms. *AACN Adv. Crit. Care* **2008**, *19*, 364–385. [[CrossRef](#)]
121. Kerkelä, R.; Ulvila, J.; Magga, J. Natriuretic peptides in the regulation of cardiovascular physiology and metabolic events. *J. Am. Heart Assoc.* **2015**, *4*, e002423. [[CrossRef](#)]
122. de Lemos, J.A.; McGuire, D.K.; Drazner, M.H. B-type natriuretic peptide in cardiovascular disease. *Lancet* **2003**, *362*, 316–322. [[CrossRef](#)]
123. Yamanouchi, S.; Kudo, D.; Endo, T.; Kitano, Y.; Shinozawa, Y. Blood N-terminal proBNP as a potential indicator of cardiac preload in patients with high volume load. *Tohoku J. Exp. Med.* **2010**, *221*, 175–180. [[CrossRef](#)]
124. Vanderheyden, M.; Bartunek, J.; Goethals, M. Brain and other natriuretic peptides: Molecular aspects. *Eur. J. Heart Fail.* **2004**, *6*, 261–268. [[CrossRef](#)]
125. Nawarskas, J.; Rajan, V.; Frishman, W.H. Vasopeptidase inhibitors, neutral endopeptidase inhibitors, and dual inhibitors of angiotensin-converting enzyme and neutral endopeptidase. *Heart Dis.* **2001**, *3*, 378–385. [[CrossRef](#)]
126. Conen, D.; Zeller, A.; Pfisterer, M.; Martina, B. Usefulness of B-Type Natriuretic Peptide and C-Reactive Protein in Predicting the Presence or Absence of Left Ventricular Hypertrophy in Patients with Systemic Hypertension. *Am. J. Cardiol.* **2006**, *97*, 249–252. [[CrossRef](#)] [[PubMed](#)]
127. Yandle, T.G.; Richards, A.M.; Nicholls, M.G.; Cuneo, R.; Espiner, E.A.; Livesey, J.H. Metabolic clearance rate and plasma half life of alpha-human atrial natriuretic peptide in man. *Life Sci.* **1986**, *38*, 1827–1833. [[CrossRef](#)]
128. Buckley, M.G.; Marcus, N.J.; Yacoub, M.H. Cardiac peptide stability, aprotinin and room temperature: Importance for assessing cardiac function in clinical practice. *Clin. Sci.* **1999**, *97*, 689–695. [[CrossRef](#)]
129. Seidler, T.; Pemberton, C.; Yandle, T.; Espiner, E.; Nicholls, G.; Richards, M. The amino terminal regions of proBNP and proANP oligomerise through leucine zipper-like coiled-coil motifs. *Biochem. Biophys. Res. Commun.* **1999**, *255*, 495–501. [[CrossRef](#)]
130. Baertschi, A.J.; Monnier, D.; Schmidt, U.; Levitan, E.S.; Fakan, S.; Roatti, A. Acid prohormone sequence determines size, shape, and docking of secretory vesicles in atrial myocytes. *Circ. Res.* **2001**, *89*, e23–e29. [[CrossRef](#)]
131. Cappellin, E.; Gatti, R.; Spinella, P.; De Palo, C.B.; Woloszczuk, W.; Maragno, I. Plasma atrial natriuretic peptide (ANP) fragments proANP (1–30) and proANP (31–67) measurements in chronic heart failure: A useful index for heart transplantation? *Clin. Chim. Acta* **2001**, *310*, 49–52. [[CrossRef](#)]

132. Fu, S.; Ping, P.; Wang, F.; Luo, L. Synthesis, secretion, function, metabolism and application of natriuretic peptides in heart failure. *J. Biol. Eng.* **2018**, *12*, 2. [[CrossRef](#)]
133. Potter, L.R. Natriuretic peptide metabolism, clearance and degradation. *FEBS J.* **2011**, *278*, 1808–1817. [[CrossRef](#)]
134. Maalouf, R.; Bailey, S. A review on B-type natriuretic peptide monitoring: Assays and biosensors. *Heart Fail. Rev.* **2016**, *21*, 567–578. [[CrossRef](#)]
135. Rodriguez, R.M.G.; Spada, R.; Pooya, S.; Jeannesson, E.; Moreno Garcia, M.A.; Anello, G. Homocysteine predicts increased NT-pro-BNP through impaired fatty acid oxidation. *Int. J. Cardiol.* **2013**, *167*, 768–775. [[CrossRef](#)]
136. Khanam, S.S.; Son, J.W.; Lee, J.W.; Youn, Y.J.; Yoon, J.; Lee, S.H.; Kim, J.Y.; Ahn, S.G.; Ahn, M.S.; Yoo, B.S. Prognostic value of short-term follow-up BNP in hospitalized patients with heart failure. *BMC Cardiovasc. Disord.* **2017**, *17*, 215. [[CrossRef](#)] [[PubMed](#)]
137. Wieczorek, S.J.; Wu, A.H.B.; Christenson, R.; Krishnaswamy, P.; Gottlieb, S.; Rosano, T. A rapid B-type natriuretic peptide assay accurately diagnoses left ventricular dysfunction and heart failure: A multicenter evaluation. *Am. Heart J.* **2002**, *144*, 834–839. [[CrossRef](#)] [[PubMed](#)]
138. Tapanainen, J.M.; Lindgren, K.S.; Mäkkilä, T.H.; Vuolteenaho, O.; Leppäläluoto, J.; Huikuri, H.V. Natriuretic peptides as predictors of non-sudden and sudden cardiac death after acute myocardial infarction in the beta-blocking era. *J. Am. Coll. Cardiol.* **2004**, *43*, 757–763. [[CrossRef](#)] [[PubMed](#)]
139. Ponikowski, P.; Voors, A. 2016 Esc guidelines for the diagnosis and treatment of acute and chronic heart failure: The Task Force for the diagnosis and treatment of acute and chronic heart failure of the European society of cardiology (ESC): Developed with the special contribution. *Russ. J. Cardiol.* **2017**, *141*, 7–81. [[CrossRef](#)]
140. Shao, M.; Huang, C.; Li, Z.; Yang, H.; Feng, Q. Effects of glutamine and valsartan on the brain natriuretic peptide and N-terminal pro-B-type natriuretic peptide of patients with chronic heart failure. *Pakistan J. Med. Sci.* **2014**, *31*, 82. [[CrossRef](#)]
141. Sun, Y.P.; Wei, C.P.; Ma, S.C.; Zhang, Y.F.; Qiao, L.Y.; Li, D.H. Effect of carvedilol on serum heart-type fatty acid-binding protein, brain natriuretic peptide, and cardiac function in patients with chronic heart failure. *J. Cardiovasc. Pharmacol.* **2015**, *65*, 480–484. [[CrossRef](#)] [[PubMed](#)]
142. Wang, T.J.; Larson, M.G.; Levy, D.; Benjamin, E.J.; Leip, E.P.; Omland, T.; Wolf, P.A.; Vasan, R.S. Plasma Natriuretic Peptide Levels and the Risk of Cardiovascular Events and Death. *N. Engl. J. Med.* **2004**, *350*, 655–663. [[CrossRef](#)]
143. Kistorp, C.; Faber, J. Levels as Predictors of Mortality and Cardiovascular Events in Older Adults. *JAMA* **2005**, *293*, 1609–1616. [[CrossRef](#)]
144. Mukoyama, M.; Nakao, K.; Hosoda, K.; Suga, S.I.; Saito, Y.; Ogawa, Y.; Shirakami, G.; Jougasaki, M.; Obata, K.; Yasue, H.; et al. Brain natriuretic peptide as a novel cardiac hormone in humans: Evidence for an exquisite dual natriuretic peptide system, atrial natriuretic peptide and brain natriuretic peptide. *J. Clin. Investig.* **1991**, *87*, 1402–1412. [[CrossRef](#)]
145. Ponikowski, P.; Voors, A.A.; Anker, S.D.; Bueno, H.; Cleland, J.G.F.; Coats, A.J.S.; Falk, V.; González-Juanatey, J.R.; Harjola, V.P.; Jankowska, E.A.; et al. 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure. *Eur. Heart J.* **2016**, *37*, 2129–2200. [[CrossRef](#)]
146. Yancy, C.W.; Jessup, M.; Bozkurt, B.; Butler, J.; Casey, D.E.; Drazner, M.H.; Fonarow, G.C.; Geraci, S.A.; Horwich, T.; Januzzi, J.L.; et al. 2013 ACCF/AHA guideline for the management of heart failure: A report of the American college of cardiology foundation/american heart association task force on practice guidelines. *J. Am. Coll. Cardiol.* **2013**, *62*, e147–e239. [[CrossRef](#)] [[PubMed](#)]
147. Hosoda, K.; Nakao, K.; Mukoyama, M.; Saito, Y.; Jougasaki, M.; Shirakami, G.; Suga, S.I.; Ogawa, Y.; Yasue, H.; Imura, H. Expression of brain natriuretic peptide gene in human heart: Production in the ventricle. *Hypertension* **1991**, *17* (Suppl. S2), 1152–1156. [[CrossRef](#)] [[PubMed](#)]
148. Mukoyama, M.; Nakao, K.; Saito, Y.; Ogawa, Y.; Hosoda, K.; Suga, S.; Shirakami, G.; Jougasaki, M.; Imura, H. Increased Human Brain Natriuretic Peptide in Congestive Heart Failure. *N. Engl. J. Med.* **1990**, *323*, 757–758. [[PubMed](#)]
149. Januzzi, J.L. Natriuretic peptides as biomarkers in heart failure. *J. Investig. Med.* **2013**, *61*, 950–955. [[CrossRef](#)]
150. Cui, K.; Huang, W.; Fan, J.; Lei, H. Midregional pro-atrial natriuretic peptide is a superior biomarker to N-terminal pro-B-type natriuretic peptide in the diagnosis of heart failure patients with preserved ejection fraction. *Medicine* **2018**, *97*, e12277. [[CrossRef](#)]
151. Morita, E.; Yasue, H.; Yoshimura, M.; Ogawa, H.; Jougasaki, M.; Matsumura, T.; Mukoyama, M.; Nakao, K. Increased plasma levels of brain natriuretic peptide in patients with acute myocardial infarction. *Circulation* **1993**, *88*, 82–91. [[CrossRef](#)]
152. Chang, H.; Min, J.K.; Rao, S.V.; Patel, M.R.; Simonetti, O.P.; Ambrosio, G.; Raman, S.V. Non-ST-segment elevation acute coronary syndromes targeted imaging to refine upstream risk stratification. *Circ. Cardiovasc. Imaging* **2012**, *5*, 536–546. [[CrossRef](#)]
153. Gill, D.; Seidler, T.; Troughton, R.W.; Yandle, T.G.; Frampton, C.M.; Richards, M.; Lainchbury, J.G.; Nicholls, G. Vigorous response in plasma N-terminal pro-brain natriuretic peptide (NT-BNP) to acute myocardial infarction. *Clin. Sci.* **2004**, *106*, 135–139. [[CrossRef](#)]
154. Chang, K.W.; Hsu, J.C.; Toomu, A.; Fox, S.; Maisel, A.S. Clinical Applications of Biomarkers in Atrial Fibrillation. *Am. J. Med.* **2017**, *130*, 1351–1357. [[CrossRef](#)]
155. Ellinor, P.T.; Low, A.F.; Patton, K.K.; Shea, M.A.; MacRae, C.A. Discordant atrial natriuretic peptide and brain natriuretic peptide levels in lone atrial fibrillation. *J. Am. Coll. Cardiol.* **2005**, *45*, 82–86. [[CrossRef](#)]

156. Knudsen, C.W.; Omland, T.; Clopton, P.; Westheim, A.; Wu, A.H.B.; Duc, P.; McCord, J.; Nowak, R.M.; Hollander, J.E.; Storrow, A.B.; et al. Impact of atrial fibrillation on the diagnostic performance of B-type natriuretic peptide concentration in dyspneic patients: An analysis from the breathing not properly multinational study. *J. Am. Coll. Cardiol.* **2005**, *46*, 838–844. [CrossRef] [PubMed]
157. Ocak, T.; Erdem, A.; Duran, A.; Tekelioğlu, Ü.Y.; Öztürk, S.; Ayhan, S.S.; Özlu, M.F.; Tosun, M.; Koçoğlu, H.; Yazici, M. The diagnostic significance of NT-proBNP and troponin I in emergency department patients presenting with palpitations. *Clinics* **2013**, *68*, 543–547. [CrossRef]
158. Qi, W.; Kjekshus, H.; Klinge, R.; Kjekshus, J.K.; Hall, C. Cardiac natriuretic peptides and continuously monitored atrial pressures during chronic rapid pacing in pigs. *Acta Physiol. Scand.* **2000**, *169*, 95–102. [CrossRef] [PubMed]
159. Detaint, D.; Detaint, D.; Messika-Zeitoun, D.; Avierinos, J.F.; Scott, C.; Chen, H.; Burnett, J.C.; Enriquez-Sarano, M. B-type natriuretic peptide in organic mitral regurgitation: Determinants and impact on outcome. *Circulation* **2005**, *111*, 2391–2397. [CrossRef] [PubMed]
160. Sutton, T.M.; Stewart, R.A.H.; Gerber, I.L.; West, T.M.; Richards, A.M.; Yandle, T.G.; Kerr, A.J. Plasma natriuretic peptide levels increase with symptoms and severity of mitral regurgitation. *J. Am. Coll. Cardiol.* **2003**, *41*, 2280–2287. [CrossRef]
161. Tharaux, P.L.; Dussaule, J.C.; Hubert-Briere, J.; Vahanian, A.; Acar, J.; Ardaillou, R. Plasma atrial and brain natriuretic peptides in mitral stenosis treated by valvulotomy. *Clin. Sci.* **1994**, *87*, 671–677. [CrossRef] [PubMed]
162. Nessmith, M.G.; Fukuta, H.; Brucks, S.; Little, W.C. Usefulness of an elevated B-type natriuretic peptide in predicting survival in patients with aortic stenosis treated without surgery. *Am. J. Cardiol.* **2005**, *96*, 1445–1448. [CrossRef]
163. Bergler-Klein, J.; Klaar, U.; Heger, M.; Rosenhek, R.; Mundigler, G.; Gabriel, H.; Binder, T.; Pacher, R.; Maurer, G.; Baumgartner, H. Natriuretic peptides predict symptom-free survival and postoperative outcome in severe aortic stenosis. *Circulation* **2004**, *109*, 2302–2308. [CrossRef]
164. Arat-Özkan, A.; Kaya, A.; Yiğit, Z.; Balci, H.; Öküçün, B.; Yazıcıoğlu, N.; Küçüköğlu, S. Serum N-terminal pro-BNP levels correlate with symptoms and echocardiographic findings in patients with mitral stenosis. *Echocardiography* **2005**, *22*, 473–478. [CrossRef]
165. Gerber, I.L.; Stewart, R.A.H.; Legget, M.E.; West, T.M.; French, R.L.; Sutton, T.M.; Yandle, T.G.; French, J.K.; Richards, A.M.; White, H.D. Increased plasma natriuretic peptide levels reflect symptom onset in aortic stenosis. *Circulation* **2003**, *107*, 1884–1890. [CrossRef]
166. Kucher, N.; Printzen, G.; Goldhaber, S.Z. Prognostic role of brain natriuretic peptide in acute pulmonary embolism. *Circulation* **2003**, *107*, 2545–2547. [CrossRef] [PubMed]
167. Kucher, N.; Printzen, G.; Doernhoefer, T.; Windecker, S.; Meier, B.; Hess, O.M. Low pro-brain natriuretic peptide levels predict benign clinical outcome in acute pulmonary embolism. *Circulation* **2003**, *107*, 1576–1578. [CrossRef] [PubMed]
168. Morrison, L.K.; Harrison, A.; Krishnaswamy, P.; Kazanegra, R.; Clopton, P.; Maisel, A. Utility of a rapid B-natriuretic peptide assay in differentiating congestive heart failure from lung disease in patients presenting with dyspnea. *J. Am. Coll. Cardiol.* **2002**, *39*, 202–209. [CrossRef]
169. Klinger, J.R.; Arnal, F.; Warburton, R.; Ou, L.C.; Hill, N.S. Downregulation of pulmonary atrial natriuretic peptide receptors in rats exposed to chronic hypoxia. *J. Appl. Physiol.* **1994**, *77*, 1309–1316. [CrossRef]
170. Bando; Ishii, Y.; Sugiyama, Y.; Kitamura, S. Elevated plasma brain natriuretic peptide levels in chronic respiratory failure with cor pulmonale. *Respir. Med.* **1999**, *93*, 507–514. [CrossRef]
171. Nagaya, N.; Nishikimi, T.; Uematsu, M.; Satoh, T.; Kyotani, S.; Sakamaki, F.; Kakishita, M.; Fukushima, K.; Okano, Y.; Nakanishi, N.; et al. Plasma brain natriuretic peptide as a prognostic indicator in patients with primary pulmonary hypertension. *Circulation* **2000**, *102*, 865–870. [CrossRef]
172. Nagaya, N.; Nishikimi, T.; Okano, Y.; Uematsu, M.; Satoh, T.; Kyotani, S.; Kurabayashi, S.; Hamada, S.; Kakishita, M.; Nakanishi, N.; et al. Plasma brain natriuretic peptide levels increase in proportion to the extent of right ventricular dysfunction in pulmonary hypertension. *J. Am. Coll. Cardiol.* **1998**, *31*, 202–208. [CrossRef]
173. Rubattu, S.; Volpe, M. Natriuretic peptides in the cardiovascular system: Multifaceted roles in physiology, pathology and therapeutics. *Int. J. Mol. Sci.* **2019**, *20*, 3991. [CrossRef]
174. Mueller, T.; Gegenhuber, A.; Dieplinger, B.; Poelz, W.; Haltmayer, M. Capability of B-type natriuretic peptide (BNP) and amino-terminal proBNP as indicators of cardiac structural disease in asymptomatic patients with systemic arterial hypertension. *Clin. Chem.* **2005**, *51*, 2245–2251. [CrossRef]
175. de Vito, P. Atrial natriuretic peptide: An old hormone or a new cytokine? *Peptides* **2014**, *58*, 108–116. [CrossRef]
176. Gerbes, A.L.; Dagnino, L.; Nguyen, T.; Nemer, M. Transcription of brain natriuretic peptide and atrial natriuretic peptide genes in human tissues. *J. Clin. Endocrinol. Metab.* **1994**, *78*, 1307–1311. [PubMed]
177. Szabó, G. Biology of the B-Type Natriuretic Peptide: Structure, Synthesis and Processing. *Biochem. Anal. Biochem.* **2012**, *1*, 8–10. [CrossRef]
178. Safley, D.M.; Wad, A.; Sullivan, R.A.; Sandberg, K.R.; Mourad, I.; Boulware, M.; Merhi, W.; McCullough, P.A. Changes in B-type natriuretic peptide levels in hemodialysis and the effect of depressed left ventricular function. *Adv. Chronic Kidney Dis.* **2005**, *12*, 117–124. [CrossRef] [PubMed]
179. Khalifeh, N.; Haider, D.; Hörl, W.H. Natriuretic peptides in chronic kidney disease and during renal replacement therapy: An update. *J. Investig. Med.* **2009**, *57*, 33–39. [CrossRef] [PubMed]

180. Yildiz, R.; Yildirim, B.; Karincaoglu, M.; Harputluoglu, M.; Hilmiooglu, F. Brain natriuretic peptide and severity of disease in non-alcoholic cirrhotic patients. *J. Gastroenterol. Hepatol.* **2005**, *20*, 1115–1120. [[CrossRef](#)]
181. Henriksen, J.H. Increased circulating pro-brain natriuretic peptide (proBNP) and brain natriuretic peptide (BNP) in patients with cirrhosis: Relation to cardiovascular dysfunction and severity of disease. *Gut* **2003**, *52*, 1511–1517. [[CrossRef](#)]
182. Lok, B.Y.; Mukerjee, D.; Timms, P.M.; Ashrafian, H.; Coghlan, J.G. Natriuretic peptides, respiratory disease, and the right heart. *Chest* **2004**, *126*, 1330–1336.
183. Kato, J.; Etoh, T.; Kitamura, K.; Eto, T. Atrial and brain natriuretic peptides as markers of cardiac load and volume retention in primary aldosteronism. *Am. J. Hypertens.* **2005**, *18*, 354–357. [[CrossRef](#)]
184. Lee, Y.J.; Lin, S.R.; Shin, S.J.; Lai, Y.H.; Lin, Y.T.; Tsai, J.H. Brain natriuretic peptide is synthesized in the human adrenal medulla and its messenger ribonucleic acid expression along with that of atrial natriuretic peptide are enhanced in patients with primary aldosteronism. *J. Clin. Endocrinol. Metab.* **1994**, *79*, 1476–1482.
185. Totsune, K.; Takahashi, K.; Murakami, O.; Satoh, F.; Sone, M.; Ohneda, M.; Miura, Y.; Mouri, T. Immunoreactive brain natriuretic peptide in human adrenal glands and adrenal tumors. *Eur. J. Endocrinol.* **1996**, *135*, 352–356. [[CrossRef](#)]
186. Schultz, M.; Faber, J.; Kistorp, C.; Jarlov, A.; Pedersen, F.; Wiinberg, N.; Hildebrandt, P. N-terminal-pro-B-type natriuretic peptide (NT-pro-BNP) in different thyroid function states. *Clin. Endocrinol.* **2004**, *60*, 54–59. [[CrossRef](#)] [[PubMed](#)]
187. Wei, T.; Zeng, C.; Tian, Y.; Chen, Q.; Wang, L. B-type natriuretic peptide in patients with clinical hyperthyroidism. *J. Endocrinol. Investig.* **2005**, *28*, 8–11. [[CrossRef](#)] [[PubMed](#)]
188. Redfield, M.M.; Rodeheffer, R.J.; Jacobsen, S.J.; Mahoney, D.W.; Bailey, K.R.; Burnett, J.C. Plasma brain natriuretic peptide concentration: Impact of age and gender. *J. Am. Coll. Cardiol.* **2002**, *40*, 976–982. [[CrossRef](#)]
189. Wang, T.J.; Larson, M.G.; Levy, D.; Leip, E.P.; Benjamin, E.J.; Wilson, P.W.F.; Sutherland, P.; Omland, T.; Vasan, R.S. Impact of age and sex on plasma natriuretic peptide levels in healthy adults. *Am. J. Cardiol.* **2002**, *90*, 254–258. [[CrossRef](#)]
190. Kawai, K.; Hata, K.; Tanaka, K.; Kubota, Y.; Inoue, R.; Masuda, E.; Miyazaki, T.; Yokoyama, M. Attenuation of biologic compensatory action of cardiac natriuretic peptide system with aging. *Am. J. Cardiol.* **2004**, *93*, 719–723. [[CrossRef](#)]
191. Costello-Boerrigter, L.C.; Boerrigter, G.; Redfield, M.M.; Rodeheffer, R.J.; Urban, L.H.; Mahoney, D.W.; Jacobsen, S.J.; Heublein, D.M.; Burnett, J.C. Amino-Terminal Pro-B-Type Natriuretic Peptide and B-Type Natriuretic Peptide in the General Community. *J Am Coll Cardiol.* **2006**, *47*, 345–353. [[CrossRef](#)]
192. Das, U.N. Heart-type fatty acid-binding protein (H-FABP) and coronary heart disease. *Indian Heart J.* **2016**, *68*, 16–18. [[CrossRef](#)]
193. van der Vusse, G.J.; Glatz, J.F.C.; Stam, H.C.G.; Reneman, R.S. Fatty acid homeostasis in the normoxic and ischemic heart. *Physiol. Rev.* **1992**, *72*, 881–940. [[CrossRef](#)]
194. Wang, S.; Zhou, Y.; Andreyev, O.; Hoyt, R.F.; Singh, A.; Hunt, T.; Horvath, K.A. Overexpression of FABP3 inhibits human bone marrow derived mesenchymal stem cell proliferation but enhances their survival in hypoxia. *Exp. Cell Res.* **2014**, *323*, 56–65. [[CrossRef](#)]
195. Zhu, C.; Hu, D.L.; Liu, Y.Q.; Zhang, Q.J.; Chen, F.K.; Kong, X.Q.; Cao, K.J.; Zhang, J.S.; Qian, L.M. Fabp3 Inhibits Proliferation and Promotes Apoptosis of Embryonic Myocardial Cells. *Cell Biochem. Biophys.* **2011**, *60*, 259–266. [[CrossRef](#)]
196. Shen, Y.; Song, G.; Liu, Y.; Zhou, L.; Liu, H.; Kong, X.; Sheng, Y.; Cao, K.; Qian, L. Silencing of FABP3 Inhibits Proliferation and Promotes Apoptosis in Embryonic Carcinoma Cells. *Cell Biochem. Biophys.* **2013**, *66*, 139–146. [[CrossRef](#)] [[PubMed](#)]
197. Binas, B.; Erol, E. FABPs as determinants of myocellular and hepatic fuel metabolism. *Mol. Cell. Biochem.* **2007**, *299*, 75–84. [[CrossRef](#)] [[PubMed](#)]
198. Chmurzyńska, A. The multigene family of fatty acid-binding proteins (FABPs): Function, structure and polymorphism. *J. Appl. Genet.* **2006**, *47*, 39–48. [[CrossRef](#)] [[PubMed](#)]
199. Liebtrau, C.; Nef, H.M.; Dörr, O.; Gaede, L.; Hoffmann, J.; Hahnel, A.; Rolf, A.; Troidl, C.; Lackner, K.J.; Keller, T.; et al. Release kinetics of early ischaemic biomarkers in a clinical model of acute myocardial infarction. *Heart* **2014**, *100*, 652–657. [[CrossRef](#)] [[PubMed](#)]
200. Iida, M.; Yamazaki, M.; Honjo, H.; Kodama, I.; Kamiya, K. Predictive value of heart-type fatty acid-binding protein for left ventricular remodelling and clinical outcome of hypertensive patients with mild-to-moderate aortic valve diseases. *J. Hum. Hypertens.* **2007**, *21*, 551–557. [[CrossRef](#)] [[PubMed](#)]
201. Fischer, T.A.; McNeil, P.L.; Khakee, R.; Finn, P.; Kelly, R.A.; Pfeffer, M.A.; Pfeffer, J.M. Cardiac Myocyte Membrane Wounding in the Abruptly Pressure-Overloaded Rat Heart Under High Wall Stress. *Hypertension* **1997**, *30*, 1041–1046. [[CrossRef](#)] [[PubMed](#)]
202. Colli, A.; Josa, M.; Pomar, J.L.; Mestres, C.A.; Gherli, T. Heart fatty acid binding protein in the diagnosis of myocardial infarction: Where do we stand today? *Cardiology* **2007**, *108*, 4–10. [[CrossRef](#)]
203. Kleine, A.H.; Glatz, J.F.C.; van Nieuwenhoven, F.A.; van der Vusse, G.J. Release of heart fatty acid-binding protein into plasma after acute myocardial infarction in man. *Mol. Cell. Biochem.* **1992**, *116*, 155–162. [[CrossRef](#)]
204. Cappellini, F.; Da Molin, S.; Signorini, S.; Avanzini, F.; Saltafossi, D.; Falbo, R.; Brambilla, P. Heart-type fatty acid-binding protein may exclude acute myocardial infarction on admission to emergency department for chest pain. *Acute Card. Care* **2013**, *15*, 83–87. [[CrossRef](#)]
205. Garcia-Valdecasas, S.; Ruiz-Alvarez, M.J.; Garcia De Tena, J.; De Pablo, R.; Huerta, I.; Barrionuevo, M.; Coca, C.; Arribas, I. Diagnostic and prognostic value of heart-type fatty acid-binding protein in the early hours of acute myocardial infarction. *Acta Cardiol.* **2011**, *66*, 315–321. [[CrossRef](#)]

206. Dupuy, A.M.; Cristol, J.P.; Kuster, N.; Reynier, R.; Lefebvre, S.; Badiou, S.; Jreige, R.; Sebbane, M. Performances of the heart fatty acid protein assay for the rapid diagnosis of acute myocardial infarction in ED patients. *Am. J. Emerg. Med.* **2015**, *33*, 326–330. [[CrossRef](#)] [[PubMed](#)]
207. Okamoto, F.; Sohmiya, K.; Ohkaro, Y.; Kawamura, K.; Asayama, K.; Kimura, H.; Nishimura, S.; Ishii, H.; Sunahara, N.; Tanaka, T. Human heart-type cytoplasmic fatty acid-binding protein (H-FABP) for the diagnosis of acute myocardial infarction. Clinical evaluation of H-FABP in comparison with myoglobin and creatine kinase isoenzyme MB. *Clin. Chem. Lab. Med.* **2000**, *38*, 231–238. [[CrossRef](#)] [[PubMed](#)]
208. Xu, L.Q.; Yang, Y.M.; Tong, H.; Xu, C.F. Early Diagnostic Performance of Heart-Type Fatty Acid Binding Protein in Suspected Acute Myocardial Infarction: Evidence From a Meta-Analysis of Contemporary Studies. *Heart Lung Circ.* **2018**, *27*, 503–512. [[CrossRef](#)] [[PubMed](#)]
209. Schoenenberger, A.W.; Stallone, F.; Walz, B.; Bergner, M.; Twerenbold, R.; Reichlin, T.; Zogg, B.; Jaeger, C.; Erne, P.; Mueller, C. Incremental value of heart-type fatty acid-binding protein in suspected acute myocardial infarction early after symptom onset. *Eur. Heart J. Acute Cardiovasc. Care* **2016**, *5*, 185–192. [[CrossRef](#)]
210. Bivona, G.; Agnello, L.; Bellia, C.; Sasso, B.L.; Ciaccio, M. Diagnostic and prognostic value of H-FABP in acute coronary syndrome: Still evidence to bring. *Clin. Biochem.* **2018**, *58*, 1–4. [[CrossRef](#)]
211. Ho, S.K.; Wu, Y.W.; Tseng, W.K.; Leu, H.B.; Yin, W.H.; Lin, T.H.; Chang, K.C.; Wang, J.H.; Yeh, H.I.; Wu, C.C.; et al. The prognostic significance of heart-type fatty acid binding protein in patients with stable coronary heart disease. *Sci. Rep.* **2018**, *8*, 14410. [[CrossRef](#)]
212. Ishino, M.; Shishido, T.; Arimoto, T.; Takahashi, H.; Miyashita, T.; Miyamoto, T.; Nitobe, J.; Watanabe, T.; Kubota, I. Heart-Type Fatty Acid Binding Protein (H-FABP) in Acute Decompensated Heart Failure. *J. Card. Fail.* **2010**, *16*, S166. [[CrossRef](#)]
213. Hoffmann, U.; Espeter, F.; Weiß, C.; Ahmad-Nejad, P.; Lang, S.; Brueckmann, M.; Akin, I.; Neumaier, M.; Borggrefe, M.; Behnes, M. Ischemic biomarker heart-type fatty acid binding protein (hFABP) in acute heart failure—Diagnostic and prognostic insights compared to NT-proBNP and troponin I. *BMC Cardiovasc. Disord.* **2015**, *15*, 50. [[CrossRef](#)]
214. Niizeki, T.; Takeishi, Y.; Arimoto, T.; Nozaki, N.; Hirono, O.; Watanabe, T.; Nitobe, J.; Miyashita, T.; Miyamoto, T.; Koyama, Y.; et al. Persistently increased serum concentration of heart-type fatty acid-binding protein predicts adverse clinical outcomes in patients with chronic heart failure. *Circ. J.* **2008**, *72*, 109–114. [[CrossRef](#)]
215. Niizeki, T.; Takeishi, Y.; Arimoto, T.; Takahashi, T.; Okuyama, H.; Takabatake, N.; Nozaki, N.; Hirono, O.; Tsunoda, Y.; Shishido, T.; et al. Combination of heart-type fatty acid binding protein and brain natriuretic peptide can reliably risk stratify patients hospitalized for chronic heart failure. *Circ. J.* **2005**, *69*, 922–927. [[CrossRef](#)]
216. Otaki, Y.; Arimoto, T.; Takahashi, H.; Kadokawa, S.; Ishigaki, D.; Narumi, T.; Honda, Y.; Iwayama, T.; Nishiyama, S.; Shishido, T.; et al. Prognostic value of myocardial damage markers in patients with chronic heart failure with atrial fibrillation. *Intern. Med.* **2014**, *53*, 661–668. [[CrossRef](#)] [[PubMed](#)]
217. Mirna, M.; Wernly, B.; Paar, V.; Jung, C.; Jirak, P.; Figulla, H.R.; Kretzschmar, D.; Franz, M.; Hoppe, U.C.; Lichtenauer, M.; et al. Multi-biomarker analysis in patients after transcatheter aortic valve implantation (TAVI). *Biomarkers* **2018**, *23*, 773–780. [[CrossRef](#)] [[PubMed](#)]
218. Qian, H.Y.; Huang, J.; Yang, Y.J.; Yang, Y.M.; Li, Z.Z.; Zhang, J.M. Heart-type Fatty Acid Binding Protein in the Assessment of Acute Pulmonary Embolism. *Am. J. Med. Sci.* **2016**, *352*, 557–562. [[CrossRef](#)] [[PubMed](#)]
219. Dellas, C.; Lobo, J.L.; Rivas, A.; Ballaz, A.; Portillo, A.K.; Nieto, R.; del Rey, J.M.; Zamorano, J.L.; Lankeit, M.; Jiménez, D. Risk stratification of acute pulmonary embolism based on clinical parameters, H-FABP and multidetector CT. *Int. J. Cardiol.* **2018**, *265*, 223–228. [[CrossRef](#)]
220. Bajaj, A.; Rathor, P.; Sehgal, V.; Shetty, A.; Kabak, B.; Hosur, S. Risk stratification in acute pulmonary embolism with heart-type fatty acid-binding protein: A meta-analysis. *J. Crit. Care* **2015**, *30*, 1151.e1–1151.e7. [[CrossRef](#)]
221. Konstantinides, S.V.; Konstantinides, S.V.; Meyer, G.; Bueno, H.; Galie, N.; Gibbs, J.S.R.; Ageno, W.; Agewall, S.; Almeida, A.G.; Andreotti, F.; et al. 2019 ESC Guidelines for the diagnosis and management of acute pulmonary embolism developed in collaboration with the European respiratory society (ERS). *Eur. Heart J.* **2020**, *41*, 543–603. [[CrossRef](#)]
222. Acher, R.; Chauvet, J.; Rouille, Y. Dynamic processing of neuropeptides: Sequential conformation shaping of neurohypophysial preprohormones during intraneuronal secretory transport. *J. Mol. Neurosci.* **2002**, *18*, 223–228. [[CrossRef](#)]
223. Christ-Crain, M. Vasopressin and Copeptin in health and disease. *Rev. Endocr. Metab. Disord.* **2019**, *20*, 283–294. [[CrossRef](#)]
224. Dietzen, D.J. Amino Acids, Peptides, and Proteins. In *Principles and Applications of Molecular Diagnostics*; Rifai, N., Horvath, A.R., Wittwer, C.T., Park, J.Y., Eds.; Elsevier: Amsterdam, The Netherlands, 2018; pp. 345–380.
225. Morgenthaler, N.G.; Struck, J.; Alonso, C.; Bergmann, A. Assay for the measurement of copeptin, a stable peptide derived from the precursor of vasopressin. *Clin. Chem.* **2006**, *52*, 112–119. [[CrossRef](#)]
226. Khan, S.Q.; Dhillon, O.S.; O'Brien, R.J.; Struck, J.; Quinn, P.A.; Morgenthaler, N.G.; Squire, I.B.; Davies, J.E.; Bergmann, A.; Ng, L.L. C-terminal provasopressin (copeptin) as a novel and prognostic marker in acute myocardial infarction: Leicester acute myocardial infarction peptide (LAMP) study. *Circulation* **2007**, *115*, 2103–2110. [[CrossRef](#)]
227. Reichlin, T.; Hochholzer, W.; Stelzig, C.; Laule, K.; Freidank, H.; Morgenthaler, N.G.; Bergmann, A.; Potocki, M.; Noveanu, M.; Breidthardt, T.; et al. Incremental Value of Copeptin for Rapid Rule Out of Acute Myocardial Infarction. *J. Am. Coll. Cardiol.* **2009**, *54*, 60–68. [[CrossRef](#)] [[PubMed](#)]

228. Lotze, U.; Lemm, H.; Heyer, A.; Müller, K. Combined determination of highly sensitive troponin T and copeptin for early exclusion of acute myocardial infarction: First experience in an emergency department of a general hospital. *Vasc. Health Risk Manag.* **2011**, *7*, 509–515. [CrossRef] [PubMed]
229. Narayan, H.; Dhillon, O.S.; Quinn, P.A.; Struck, J.; Squire, I.B.; Davies, J.E.; NG, L.L. C-terminal provasopressin (copeptin) as a prognostic marker after acute non-ST elevation myocardial infarction: Leicester acute myocardial infarction peptide II (LAMP II) study. *Clin. Sci.* **2011**, *121*, 79–89. [CrossRef] [PubMed]
230. Balmelli, C.; Meune, C.; Twerenbold, R.; Reichlin, T.; Rieder, S.; Drexler, B.; Rubini, M.G.; Mosimann, T.; Reiter, M.; Haaf, P.; et al. Comparison of the performances of cardiac troponins, including sensitive assays, and copeptin in the diagnostic of acute myocardial infarction and long-term prognosis between women and men. *Am. Heart J.* **2013**, *166*, 30–37. [CrossRef] [PubMed]
231. Kelly, D.; Squire, I.B.; Khan, S.Q.; Quinn, P.; Struck, J.; Morgenthaler, N.G.; Davies, J.E.; NG, L.L. C-Terminal Provasopressin (Copeptin) is Associated with Left Ventricular Dysfunction, Remodeling, and Clinical Heart Failure in Survivors of Myocardial Infarction. *J. Card. Fail.* **2008**, *14*, 739–745. [CrossRef] [PubMed]
232. Keller, T.; Tzikas, S.; Zeller, T.; Czyz, E.; Lillpopp, L.; Ojeda, F.M.; Roth, A.; Bickel, C.; Baldus, S.; Sinning, C.R.; et al. Copeptin Improves Early Diagnosis of Acute Myocardial Infarction. *J. Am. Coll. Cardiol.* **2010**, *55*, 2096–2106. [CrossRef]
233. Schrier, R.W.; Abraham, W.T. Mechanisms of disease: Hormones and hemodynamics in heart failure. *N. Engl. J. Med.* **1999**, *340*, 577–585. [CrossRef]
234. Thibonnier, M. Vasopressin receptor antagonists in heart failure. *Curr. Opin. Pharmacol.* **2003**, *3*, 683–687. [CrossRef]
235. Goldsmith, S.R. Congestive heart failure: Potential role of arginine vasopressin antagonists in the therapy of heart failure. *Congest. Heart Fail.* **2002**, *8*, 251–256. [CrossRef]
236. Gegenhuber, A.; Struck, J.; Dieplinger, B.; Poelz, W.; Pacher, R.; Morgenthaler, N.G.; Bergmann, A.; Haltmayer, M.; Mueller, T. Comparative Evaluation of B-Type Natriuretic Peptide, Mid-Regional Pro-A-type Natriuretic Peptide, Mid-Regional Pro-Adrenomedullin, and Copeptin to Predict 1-Year Mortality in Patients with Acute Destabilized Heart Failure. *J. Card. Fail.* **2007**, *13*, 42–49. [CrossRef]
237. Neuhold, S.; Huelsmann, M.; Strunk, G.; Struck, J.; Adlbrecht, C.; Gouya, G.; Elhenicky, M.; Pacher, R. Prognostic value of emerging neurohormones in chronic heart failure during optimization of heart failure-specific therapy. *Clin. Chem.* **2010**, *56*, 121–126. [CrossRef] [PubMed]
238. Neuhold, S.; Huelsmann, M.; Strunk, G.; Stoiser, B.; Struck, J.; Morgenthaler, N.G.; Bergmann, A.; Moertl, D.; Berger, R.; Pacher, R. Comparison of Copeptin, B-Type Natriuretic Peptide, and Amino-Terminal Pro-B-Type Natriuretic Peptide in Patients with Chronic Heart Failure. Prediction of Death at Different Stages of the Disease. *J. Am. Coll. Cardiol.* **2008**, *52*, 266–272. [CrossRef] [PubMed]
239. Stoiser, B.; Mörtl, D.; Hülsmann, M.; Berger, R.; Struck, J.; Morgenthaler, N.G.; Bergmann, A.; Pacher, R. Copeptin, a fragment of the vasopressin precursor, as a novel predictor of outcome in heart failure. *Eur. J. Clin. Investig.* **2006**, *36*, 771–778. [CrossRef] [PubMed]
240. Alehagen, U.; Dahlström, U.; Rehfeld, J.F.; Goetze, J.P. Association of copeptin and N-terminal proBNP concentrations with risk of cardiovascular death in older patients with symptoms of heart failure. *JAMA J. Am. Med. Assoc.* **2011**, *305*, 2088–2095. [CrossRef] [PubMed]
241. Masson, S.; Latini, R.; Carbonieri, E.; Moretti, L.; Rossi, M.G.; Cricugno, S.; Milani, V.; Marchioli, R.; Struck, J.; Bergmann, A.; et al. The predictive value of stable precursor fragments of vasoactive peptides in patients with chronic heart failure: Data from the GISSI-heart failure (GISSI-HF) trial. *Eur. J. Heart Fail.* **2010**, *12*, 338–347. [CrossRef] [PubMed]
242. Tentzeris, I.; Jarai, R.; Farhan, S.; Perkmann, T.; Schwarz, M.A.; Jakl, G.; Wojta, J.; Huber, K. Complementary role of copeptin and high-sensitivity troponin in predicting outcome in patients with stable chronic heart failure. *Eur. J. Heart Fail.* **2011**, *13*, 726–733. [CrossRef] [PubMed]
243. Pozsonyi, Z.; Förhész, Z.; Gombos, T.; Karádi, I.; Jánoskuti, L.; Prohászka, Z. Copeptin (C-terminal pro Arginine-Vasopressin) is an independent long-term prognostic marker in heart failure with reduced ejection fraction. *Heart Lung Circ.* **2015**, *24*, 359–367. [CrossRef]
244. Morello, F.; Oddi, M.; Cavalot, G.; Ianniello, A.; Giachino, F.; Nazerian, P.; Battista, S.; Magnino, C.; Tizzani, M.; Settanni, F.; et al. Prospective diagnostic and prognostic study of copeptin in suspected acute aortic syndromes. *Sci. Rep.* **2018**, *8*, 16713. [CrossRef]
245. Bosch, A.; Ott, C.; Schmid, A.; Kannenkeril, D.; Karg, M.; Ditting, T.; Veelken, R.; Uder, M.; Schmieder, R.E. Copeptin As a Research Marker in Cardiovascular Disease. *J. Hypertens.* **2018**, *36*, e35. [CrossRef]
246. Afsar, B. Pathophysiology of copeptin in kidney disease and hypertension. *Clin. Hypertens.* **2017**, *23*, 13. [CrossRef]
247. Tenderenda-Banasiuk, E.; Wasilewska, A.; Filonowicz, R.; Jakubowska, U.; Waszkiewicz-Stojda, M. Serum copeptin levels in adolescents with primary hypertension. *Pediatr. Nephrol.* **2014**, *29*, 423–429. [CrossRef]
248. Dobša, L.; Edozien, C.K. Copeptin and its potential role in diagnosis and prognosis of various diseases. *Biochem. Med.* **2012**, *23*, 172–190.
249. Jougasaki, M.; Burnett, J.C. Adrenomedullin: Potential in physiology and pathophysiology. *Life Sci.* **2000**, *66*, 855–872. [CrossRef]
250. Voors, A.A.; Kremer, D.; Geven, C.; ter Maaten, J.M.; Struck, J.; Bergmann, A.; Pickkers, P.; Metra, M.; Mebazaa, A.; Düngen, H.D.; et al. Adrenomedullin in heart failure: Pathophysiology and therapeutic application. *Eur. J. Heart Fail.* **2019**, *21*, 163–171. [CrossRef] [PubMed]

251. Nishikimi, T.; Yoshihara, F.; Mori, Y.; Kangawa, K.; Matsuoka, H. Cardioprotective effect of adrenomedullin in heart failure. *Hypertens. Res.* **2003**, *26*, 121–127. [[CrossRef](#)]
252. Sugo, S.; Minamino, N.; Kangawa, K.; Miyamoto, K.; Kitamura, K.; Sakata, J.; Eto, T.; Matsu, H. Endothelial Cells Actively Synthesize and Secrete Adrenomedullin. *Biochem. Biophys. Res. Commun.* **1994**, *201*, 1160–1166. [[CrossRef](#)]
253. Kitamura, K.; Matsui, E.; Kato, J.; Katoh, F.; Kita, T.; Tsuji, T.; Kangawa, K.; Eto, T. Adrenomedullin (11–26): A novel endogenous hypertensive peptide isolated from bovine adrenal medulla. *Peptides* **2001**, *22*, 1713–1718. [[CrossRef](#)]
254. Nishikimi, T.; Kuwahara, K.; Nakagawa, Y.; Kangawa, K.; Nakao, K. *Adrenomedullin*; Elsevier Inc.: Amsterdam, The Netherlands, 2017.
255. Morgenthaler, N.G.; Struck, J.; Alonso, C.; Bergmann, A. Measurement of midregional proadrenomedullin in plasma with an immunoluminometric assay. *Clin. Chem.* **2005**, *51*, 1823–1829. [[CrossRef](#)]
256. Meeran, K.; O’Shea, D.; Upton, P.D.; Small, C.J.; Ghatei, M.A.; Byfield, P.H.; Bloom, S.R. Circulating adrenomedullin does not regulate systemic blood pressure but increases plasma prolactin after intravenous infusion in humans: A pharmacokinetic study. *J. Clin. Endocrinol. Metab.* **1997**, *82*, 95–100. [[CrossRef](#)]
257. Caruhel, P.; Mazier, C.; Kunde, J.; Morgenthaler, N.G.; Darbouret, B. Homogeneous time-resolved fluoroimmunoassay for the measurement of midregional proadrenomedullin in plasma on the fully automated system B.R.A.H.M.S. KRYPTOR®. *Clin. Biochem.* **2009**, *42*, 725–728. [[CrossRef](#)]
258. Maisel, A.; Mueller, C.; Nowak, R.; Peacock, W.F.; Landsberg, J.W.; Ponikowski, P.; Mockel, M.; Hogan, C.; Wu, A.H.B.; Richards, M.; et al. Mid-Region Pro-Hormone Markers for Diagnosis and Prognosis in Acute Dyspnea. Results From the BACH (Biomarkers in Acute Heart Failure) Trial. *J. Am. Coll. Cardiol.* **2010**, *55*, 2062–2076. [[CrossRef](#)] [[PubMed](#)]
259. Bunton, D.C.; Petrie, M.C.; Hillier, C.; Johnston, F.; McMurray, J.J.V. The clinical relevance of adrenomedullin: A promising profile? *Pharmacol. Ther.* **2004**, *103*, 179–201. [[CrossRef](#)] [[PubMed](#)]
260. Jougasaki, M.; Rodeheffer, R.J.; Redfield, M.M.; Yamamoto, K.; Wei, C.M.; McKinley, L.J.; Burnett, J.C. Cardiac secretion of adrenomedullin in human heart failure. *J. Clin. Investig.* **1996**, *97*, 2370–2376. [[CrossRef](#)] [[PubMed](#)]
261. Yu, C.M.; Cheung, B.M.Y.; Leung, R.; Wang, Q.; Lai, W.H.; Lau, C.P. Increase in plasma adrenomedullin in patients with heart failure characterised by diastolic dysfunction. *Heart* **2001**, *86*, 155–160.
262. Pousset, F.; Masson, F.; Chavirovskaia, O.; Isnard, R.; Carayon, A.; Golmard, J.L.; Lechat, P.; Thomas, D.; Komajda, M. Plasma adrenomedullin, a new independent predictor of prognosis in patients with chronic heart failure. *Eur. Heart J.* **2000**, *21*, 1009–1014. [[CrossRef](#)]
263. Richards, A.M.; Doughty, R.; Nicholls, M.G.; MacMahon, S.; Sharpe, N.; Murphy, J.; Espiner, E.A.; Frampton, C.; Yandle, T.G. Plasma N-terminal pro-brain natriuretic peptide and adrenomedullin: Prognostic utility and prediction of benefit from carvedilol in chronic ischemic left ventricular dysfunction. *J. Am. Coll. Cardiol.* **2001**, *37*, 1781–1787. [[CrossRef](#)]
264. Shah, R.V.; Truong, Q.A.; Gaggin, H.K.; Pfannkuche, J.; Hartmann, O.; Januzzi, J.L. Mid-regional pro-atrial natriuretic peptide and pro-adrenomedullin testing for the diagnostic and prognostic evaluation of patients with acute dyspnoea. *Eur. Heart J.* **2012**, *33*, 2197–2205. [[CrossRef](#)]
265. Nishikimi, T.; Saito, Y.; Kitamura, K.; Ishimitsu, T.; Eto, T.; Kangawa, K.; Matsu, H.; Omae, T.; Matsuoka, H. Increased plasma levels of adrenomedullin in patients with heart failure. *J. Am. Coll. Cardiol.* **1995**, *26*, 1424–1431. [[CrossRef](#)]
266. Klip, I.; Voors, A.A.; Anker, S.D.; Hillege, H.L.; Struck, J.; Squire, I.; Van Veldhuisen, D.J.; Dickstein, K. Prognostic value of mid-regional pro-adrenomedullin in patients with heart failure after an acute myocardial infarction. *Heart* **2011**, *97*, 892–898. [[CrossRef](#)]
267. Sabatine, M.S.; Morrow, D.A.; de Lemos, J.A.; Omland, T.; Sloan, S.; Jarolim, P.; Solomon, S.D.; Pfeffer, M.A.; Braunwald, E. Evaluation of Multiple Biomarkers of Cardiovascular Stress for Risk Prediction and Guiding Medical Therapy in Patients with Stable Coronary Disease. *Circulation* **2012**, *125*, 233–240. [[CrossRef](#)]
268. Khan, S.Q.; O’Brien, R.J.; Struck, J.; Quinn, P.; Morgenthaler, N.; Squire, I.; Davies, J.; Bergmann, A.; Ng, L.L. Prognostic Value of Midregional Pro-Adrenomedullin in Patients with Acute Myocardial Infarction. The LAMP (Leicester Acute Myocardial Infarction Peptide) Study. *J. Am. Coll. Cardiol.* **2007**, *49*, 1525–1532. [[CrossRef](#)] [[PubMed](#)]
269. Ishimitsu, T.; Nishikimi, T.; Saito, Y.; Kitamura, K.; Eto, T.; Kangawa, K.; Matsu, H.; Omae, T.; Matsuoka, H. Plasma levels of adrenomedullin, a newly identified hypotensive peptide, in patients with hypertension and renal failure. *J. Clin. Investig.* **1994**, *94*, 2158–2161. [[CrossRef](#)] [[PubMed](#)]
270. McEver, R.P. Selectins: Initiators of leucocyte adhesion and signalling at the vascular wall. *Cardiovasc. Res.* **2015**, *107*, 331–339. [[CrossRef](#)]
271. Dong, Z.M.; Chapman, S.M.; Brown, A.A.; Frenette, P.S.; Hynes, R.O.; Wagner, D.D. The combined role of P- and E-selectins in atherosclerosis. *J. Clin. Investig.* **1998**, *102*, 145–152. [[CrossRef](#)] [[PubMed](#)]
272. Collins, R.G.; Velji, R.; Guevara, N.V.; Hicks, M.J.; Chan, L.; Beaudet, A.L. P-selectin or intercellular adhesion molecule (ICAM)-1 deficiency substantially protects against atherosclerosis in apolipoprotein E-deficient mice. *J. Exp. Med.* **2000**, *191*, 189–194. [[CrossRef](#)]
273. An, G.; Wang, H.; Tang, R.; Yago, T.; McDaniel, J.M.; McGee, S.; Huo, Y.; Xia, L. P-selectin glycoprotein ligand-1 is highly expressed on ly-6Chi monocytes and a major determinant for ly-6Chi monocyte recruitment to sites of atherosclerosis in mice. *Circulation* **2008**, *117*, 3227–3237. [[CrossRef](#)]
274. Johnson-Tidey, R.R.; McGregor, J.L.; Taylor, P.R.; Poston, R.N. Increase in the adhesion molecule P-selectin in endothelium overlying atherosclerotic plaques: Coexpression with intercellular adhesion molecule-1. *Am. J. Pathol.* **1994**, *144*, 952–961.

275. Body, R.; Pemberton, P.; Ali, F.; McDowell, G.; Carley, S.; Smith, A.; Mackway-Jones, K. Low soluble P-selectin may facilitate early exclusion of acute myocardial infarction. *Clin. Chim. Acta* **2011**, *412*, 614–618. [[CrossRef](#)]
276. Thomas, M.R.; Wijeyeratne, Y.D.; May, J.A.; Johnson, A.; Heptinstall, S.; Fox, S.C. A platelet P-selectin test predicts adverse cardiovascular events in patients with acute coronary syndromes treated with aspirin and clopidogrel. *Platelets* **2014**, *25*, 612–618. [[CrossRef](#)]
277. Tardif, J.C.; Tanguay, J.F.; Wright, S.S.; Duchatelle, V.; Petroni, T.; Grégoire, J.C.; Ibrahim, R.; Heinonen, T.M.; Robb, S.; Bertrand, O.F.; et al. Effects of the P-selectin antagonist inclacumab on myocardial damage after percutaneous coronary intervention for non-st-segment elevation myocardial infarction: Results of the SELECT-ACS trial. *J. Am. Coll. Cardiol.* **2013**, *61*, 2048–2055. [[CrossRef](#)]
278. Tscharre, M.; Vogel, B.; Tentzeris, I.; Freynhofer, M.K.; Rohla, M.; Wojta, J.; Weiss, T.W.; Ay, C.; Huber, K.; Farhan, S. Prognostic Impact of Soluble P-Selectin on Long-Term Adverse Cardiovascular Outcomes in Patients Undergoing Percutaneous Coronary Intervention. *Thromb. Haemost.* **2019**, *119*, 340–347. [[CrossRef](#)] [[PubMed](#)]
279. Ridker, P.M.; Buring, J.E.; Rifai, N. Soluble P-Selectin and the Risk of Future Cardiovascular Events. *Circulation* **2001**, *103*, 491–495. [[CrossRef](#)] [[PubMed](#)]
280. Parker, B.A.; Augeri, A.L.; Capizzi, J.A.; Ballard, K.D.; Kupchak, B.R.; Volek, J.S.; Troyanos, C.; Kriz, P.; D’Hemeecourt, P.; Thompson, P.D. Effect of marathon run and air travel on pre- and post-run soluble d-dimer, microparticle procoagulant activity, and p-selectin levels. *Am. J. Cardiol.* **2012**, *109*, 1521–1525. [[CrossRef](#)] [[PubMed](#)]
281. Fuhrman, B. The urokinase system in the pathogenesis of atherosclerosis. *Atherosclerosis* **2012**, *222*, 8–14. [[CrossRef](#)] [[PubMed](#)]
282. Blasi, F.; Carmeliet, P. uPAR: A versatile signalling orchestrator. *Nat. Rev. Mol. Cell Biol.* **2002**, *3*, 932–943. [[CrossRef](#)] [[PubMed](#)]
283. Madsen, C.D.; Ferraris, G.M.S.; Andolfo, A.; Cunningham, O.; Sidenius, N. uPAR-induced cell adhesion and migration: Vitronectin provides the key. *J. Cell Biol.* **2007**, *177*, 927–939. [[CrossRef](#)]
284. Madsen, C.D.; Sidenius, N. The interaction between urokinase receptor and vitronectin in cell adhesion and signalling. *Eur. J. Cell Biol.* **2008**, *87*, 617–629. [[CrossRef](#)]
285. Eapen, D.J.; Manocha, P.; Ghasemzadeh, N.; Patel, R.S.; Kassem, H.A.L.; Hammadah, M.; Veledar, E.; Le, N.A.; Pielak, T.; Thorball, C.W.; et al. Soluble urokinase plasminogen activator receptor level is an independent predictor of the presence and severity of coronary artery disease and of future adverse events. *J. Am. Heart Assoc.* **2014**, *3*, e001118. [[CrossRef](#)]
286. Corban, M.T.; Prasad, A.; Nesbitt, L.; Loeffler, D.; Herrmann, J.; Lerman, L.O.; Lerman, A. Local production of soluble urokinase plasminogen activator receptor and plasminogen activator inhibitor-1 in the coronary circulation is associated with coronary endothelial dysfunction in humans. *J. Am. Heart Assoc.* **2018**, *7*, e009881. [[CrossRef](#)]
287. Lyngbæk, S.; Andersson, C.; Marott, J.L.; Møller, D.V.; Christiansen, M.; Iversen, K.K.; Clemmensen, P.; Eugen-Olsen, J.; Hansen, P.R.; Jeppesen, J.L. Soluble urokinase plasminogen activator receptor for risk prediction in patients admitted with acute chest pain. *Clin. Chem.* **2013**, *59*, 1621–1629. [[CrossRef](#)]
288. Jung, R.G.; Motazedian, P.; Ramirez, F.D.; Simard, T.; Di Santo, P.; Visintini, S.; Faraz, M.A.; Labinaz, A.; Jung, Y.; Hibbert, B. Association between plasminogen activator inhibitor-1 and cardiovascular events: A systematic review and meta-analysis. *Thromb. J.* **2018**, *16*, 12. [[CrossRef](#)] [[PubMed](#)]
289. Lyngbæk, S.; Marott, J.L.; Møller, D.V.; Christiansen, M.; Iversen, K.K.; Clemmensen, P.M.; Eugen-Olsen, J.; Jeppesen, J.L.; Hansen, P.R. Usefulness of soluble urokinase plasminogen activator receptor to predict repeat myocardial infarction and mortality in patients with st-segment elevation myocardial infarction undergoing primary percutaneous intervention. *Am. J. Cardiol.* **2012**, *110*, 1756–1763. [[CrossRef](#)] [[PubMed](#)]
290. Marcucci, R.; Brogi, D.; Sofi, F.; Giglioli, C.; Valente, S.; Liotta, A.A.; Lenti, M.; Gori, A.M.; Prisco, D.; Abbate, R.; et al. PAI-1 and homocysteine, but not lipoprotein (a) and thrombophilic polymorphisms, are independently associated with the occurrence of major adverse cardiac events after successful coronary stenting. *Heart* **2006**, *92*, 377–381. [[CrossRef](#)] [[PubMed](#)]
291. Eugen-Olsen, J.; Andersen, O.; Linneberg, A.; Ladelund, S.; Hansen, T.W.; Langkilde, A.; Petersen, J.; Pielak, T.; Møller, L.N.; Jeppesen, J.; et al. Circulating soluble urokinase plasminogen activator receptor predicts cancer, cardiovascular disease, diabetes and mortality in the general population. *J. Intern. Med.* **2010**, *268*, 296–308. [[CrossRef](#)]
292. Lyngbæk, S.; Marott, J.L.; Sehestedt, T.; Hansen, T.W.; Olsen, M.H.; Andersen, O.; Linneberg, A.; Haugaard, S.B.; Eugen-Olsen, J.; Hansen, P.R.; et al. Cardiovascular risk prediction in the general population with use of suPAR, CRP, and Framingham Risk Score. *Int. J. Cardiol.* **2013**, *167*, 2904–2911. [[CrossRef](#)]
293. Cesari, M.; Pahor, M.; Incalzi, R.A. REVIEW: Plasminogen Activator Inhibitor-1 (PAI-1): A Key Factor Linking Fibrinolysis and Age-Related Subclinical and Clinical Conditions. *Cardiovasc. Ther.* **2010**, *28*, e72–e91. [[CrossRef](#)]
294. Ochieng, J.; Furtak, V.; Lukyanov, P. Extracellular functions of galectin-3. *Glycoconj. J.* **2002**, *19*, 527–535. [[CrossRef](#)]
295. Chen, H.Y.; Weng, I.C.; Hong, M.H.; Liu, F.T. Galectins as bacterial sensors in the host innate response. *Curr. Opin. Microbiol.* **2014**, *17*, 75–81. [[CrossRef](#)]
296. Duman, J.; Dabelic, S.; Flögel, M. Galectin-3: An open-ended story. *Biochim. Biophys. Acta Gen. Subj.* **2006**, *1760*, 616–635. [[CrossRef](#)]
297. Yang, R.Y.; Rabinovich, G.A.; Liu, F.T. Galectins: Structure, function and therapeutic potential. *Expert Rev. Mol. Med.* **2008**, *10*, e17. [[CrossRef](#)]
298. Lala, R.I.; Puschita, M.; Darabantu, D.; Pilat, L. Galectin-3 in heart failure pathology—‘Another brick in the wall?’ *Acta Cardiol.* **2015**, *70*, 323–331. [[CrossRef](#)] [[PubMed](#)]

299. Clementy, N.; Garcia, B.; André, C.; Bisson, A.; Benhenda, N.; Pierre, B.; Bernard, A.; Fauchier, L.; Piver, E.; Babuty, D. Galectin-3 level predicts response to ablation and outcomes in patients with persistent atrial fibrillation and systolic heart failure. *PLoS ONE* **2018**, *13*, e0201517. [CrossRef] [PubMed]
300. Andre, C.; Piver, E.; Perault, R.; Bisson, A.; Pucheuex, J.; Vermes, E.; Pierre, B.; Fauchier, L.; Babuty, D.; Clementy, N. Galectin-3 predicts response and outcomes after cardiac resynchronization therapy. *11 Medical and Health Sciences 1102 Cardiorespiratory Medicine and Haematology. J. Transl. Med.* **2018**, *16*, 299.
301. Agnello, L.; Bivona, G.; Lo Sasso, B.; Scazzone, C.; Bazan, V.; Bellia, C.; Ciaccio, M. Galectin-3 in acute coronary syndrome. *Clin. Biochem.* **2017**, *50*, 797–803. [CrossRef]
302. Weir, R.A.P.; Petrie, C.J.; Murphy, C.A.; Clements, S.; Steedman, T.; Miller, A.M.; McInnes, I.B.; Squire, I.B.; Ng, L.L.; Dargie, H.J.; et al. Galectin-3 and cardiac function in survivors of acute myocardial infarction. *Circ. Heart Fail.* **2013**, *6*, 492–498. [CrossRef]
303. Ipek, E.G.; Akin Suljevic, S.; Kafes, H.; Basyigit, F.; Karalok, N.; Guray, Y.; Dinc Asarcikli, L.; Acar, B.; Demirel, H. Evaluation of galectin-levels in acute coronary syndrome. *Ann. Cardiol. Angeiol.* **2016**, *65*, 26–30. [CrossRef]
304. George, M.; Shanmugam, E.; Srivatsan, V.; Rajaram, M.; Jena, A.; Sridhar, A.; Vasanth, K.; Chaudhury, M.; Kaliappan, I.; Ramraj, B. Value of pentraxin-3 and galectin-3 in acute coronary syndrome: A short-term prospective cohort study. *Ther. Adv. Cardiovasc. Dis.* **2015**, *9*, 275–284. [CrossRef]
305. Felker, G.M.; Fiuzat, M.; Shaw, L.K.; Clare, R.; Whellan, D.J.; Bettari, L.; Shirolkar, S.C.; Donahue, M.; Kitzman, D.W.; Zannad, F.; et al. Galectin-3 in ambulatory patients with heart failure results from the HF-ACTION study. *Circ. Heart Fail.* **2012**, *5*, 72–78. [CrossRef]
306. Noguchi, K.; Tomita, H.; Kanayama, T.; Niwa, A.; Hatano, Y.; Hoshi, M.; Sugie, S.; Okada, H.; Niwa, M.; Hara, A. Time-course analysis of cardiac and serum galectin-3 in viral myocarditis after an encephalomyocarditis virus inoculation. *PLoS ONE* **2019**, *14*, e0210971. [CrossRef]
307. de Couto, G.; Ouzounian, M.; Liu, P.P. Early detection of myocardial dysfunction and heart failure. *Nat. Rev. Cardiol.* **2010**, *7*, 334–344. [CrossRef]
308. Zuern, C.S.; Floss, N.; Mueller, I.I.; Eick, C.; Duckheim, M.; Patzelt, J.; Gawaz, M.; May, A.E.; Mueller, K.A.L. Galectin-3 is associated with left ventricular reverse remodeling and outcome after percutaneous mitral valve repair. *Int. J. Cardiol.* **2018**, *263*, 104–110. [CrossRef] [PubMed]
309. Asleh, R.; Enriquez-Sarano, M.; Jaffe, A.S.; Manemann, S.M.; Weston, S.A.; Jiang, R.; Roger, V.L. Galectin-3 Levels and Outcomes After Myocardial Infarction. *J. Am. Coll. Cardiol.* **2019**, *73*, 2286–2295. [CrossRef] [PubMed]
310. Cui, Y.; Qi, X.; Huang, A.; Li, J.; Hou, W.; Liu, K. Differential and predictive value of galectin-3 and soluble suppression of tumorigenicity-2 (sST2) in heart failure with preserved ejection fraction. *Med. Sci. Monit.* **2018**, *24*, 5139–5146. [CrossRef] [PubMed]
311. Ghorbani, A.; Bhambhani, V.; Christenson, R.H.; Meijers, W.C.; de Boer, R.A.; Levy, D.; Larson, M.G.; Ho, J.E. Longitudinal Change in Galectin-3 and Incident Cardiovascular Outcomes. *J. Am. Coll. Cardiol.* **2018**, *72*, 3246–3254. [CrossRef]
312. Dupuy, A.M.; Kuster, N.; Curinier, C.; Huet, F.; Plawecki, M.; Solecki, K.; Roubille, F.; Cristol, J.P. Exploring collagen remodeling and regulation as prognosis biomarkers in stable heart failure. *Clin. Chim. Acta* **2019**, *490*, 167–171. [CrossRef]
313. van Kimmenade, R.R.; Januzzi, J.L.; Ellinor, P.T.; Sharma, U.C.; Bakker, J.A.; Low, A.F.; Martinez, A.; Crijns, H.J.; MacRae, C.A.; Menheere, P.P.; et al. Utility of Amino-Terminal Pro-Brain Natriuretic Peptide, Galectin-3, and Apelin for the Evaluation of Patients with Acute Heart Failure. *J. Am. Coll. Cardiol.* **2006**, *48*, 1217–1224. [CrossRef]
314. de Boer, R.A.; Lok, D.J.A.; Jaarsma, T.; Van Der Meer, P.; Voors, A.A.; Hillege, H.L.; Van Veldhuisen, D.J. Predictive value of plasma galectin-3 levels in heart failure with reduced and preserved ejection fraction. *Ann. Med.* **2011**, *43*, 60–68. [CrossRef]
315. Lopez-Andrés, N.; Rossignol, P.; Iraqi, W.; Fay, R.; Nuée, J.; Ghio, S.; Cleland, J.G.F.; Zannad, F.; Lacolley, P. Association of galectin-3 and fibrosis markers with long-term cardiovascular outcomes in patients with heart failure, left ventricular dysfunction, and dyssynchrony: Insights from the CARE-HF (Cardiac Resynchronization in Heart Failure) trial. *Eur. J. Heart Fail.* **2012**, *14*, 74–81. [CrossRef]
316. van der Velde, A.R.; Gullestad, L.; Ueland, T.; Aukrust, P.; Guo, Y.; Adourian, A.; Muntendam, P.; Van Veldhuisen, D.J.; De Boer, R.A. Prognostic value of changes in galectin-3 levels over time in patients with heart failure data from CORONA and COACH. *Circ. Heart Fail.* **2013**, *6*, 219–226. [CrossRef]
317. Baldenhofer, G.; Baldenhofer, G.; Zhang, K.; Spethmann, S.; Laule, M.; Eilers, B.; Leonhardt, F.; Sanad, W.; Dreger, H.; Sander, M.; et al. Galectin-3 predicts short- and long-term outcome in patients undergoing transcatheter aortic valve implantation (TAVI). *Int. J. Cardiol.* **2014**, *177*, 912–917. [CrossRef]
318. Srivatsan, V.; George, M.; Shanmugam, E. Utility of galectin-3 as a prognostic biomarker in heart failure: Where do we stand? *Eur. J. Prev. Cardiol.* **2015**, *22*, 1096–1110. [CrossRef] [PubMed]
319. Tan, K.C.B.; Cheung, C.L.; Lee, A.C.H.; Lam, J.K.Y.; Wong, Y.; Shiu, S.W.M. Galectin-3 is independently associated with progression of nephropathy in type 2 diabetes mellitus. *Diabetologia* **2018**, *61*, 1212–1219. [CrossRef] [PubMed]
320. Alam, M.L.; Katz, R.; Bellovich, K.A.; Bhat, Z.Y.; Brosius, F.C.; de Boer, I.H.; Gadegbeku, C.A.; Gipson, D.S.; Hawkins, J.J.; Himmelfarb, J.; et al. Soluble ST2 and Galectin-3 and Progression of CKD. *Kidney Int. Rep.* **2019**, *4*, 103–111. [CrossRef] [PubMed]
321. Savoj, J.; Becerra, B.; Kim, J.K.; Fusaro, M.; Gallieni, M.; Lombardo, D.; Lau, W.L. Utility of Cardiac Biomarkers in the Setting of Kidney Disease. *Nephron* **2019**, *141*, 227–235. [CrossRef]

322. Chen, S.C.; Kuo, P.L. The role of galectin-3 in the kidneys. *Int. J. Mol. Sci.* **2016**, *17*, 565. [[CrossRef](#)]
323. Gopal, D.M.; Ayalon, N.; Wang, Y.C.; Siwik, D.; Sverdlov, A.; Donohue, C.; Perez, A.; Downing, J.; Apovian, C.; Silva, V.; et al. Galectin-3 is associated with stage B metabolic heart disease and pulmonary hypertension in young obese patients. *J. Am. Heart Assoc.* **2019**, *8*, e011100. [[CrossRef](#)]
324. Nielsen, C.T.; Østergaard, O.; Rasmussen, N.S.; Jacobsen, S.; Heegaard, N.H.H. A review of studies of the proteomes of circulating microparticles: Key roles for galectin-3-binding protein-expressing microparticles in vascular diseases and systemic lupus erythematosus. *Clin. Proteom.* **2017**, *14*, 11. [[CrossRef](#)]
325. Kobayashi, K.; Niwa, M.; Hoshi, M.; Saito, K.; Hisamatsu, K.; Hatano, Y.; Tomita, H.; Miyazaki, T.; Hara, A. Early microlesion of viral encephalitis confirmed by galectin-3 expression after a virus inoculation. *Neurosci. Lett.* **2015**, *592*, 107–112. [[CrossRef](#)]
326. Sato, S.; Ouellet, M.; St-Pierre, C.; Tremblay, M.J. Glycans, galectins, and HIV-1 infection. *Ann. N. Y. Acad. Sci.* **2012**, *1253*, 133–148. [[CrossRef](#)]
327. de Oliveira, F.L.; Gatto, M.; Bassi, N.; Luisetto, R.; Ghirardello, A.; Punzi, L.; Doria, A. Galectin-3 in autoimmunity and autoimmune diseases. *Exp. Biol. Med.* **2015**, *240*, 1019–1028. [[CrossRef](#)]
328. Dhirapong, A.; Lleo, A.; Leung, P.; Gershwin, M.E.; Liu, F.T. The immunological potential of galectin-1 and -3. *Autoimmun. Rev.* **2009**, *8*, 360–363. [[CrossRef](#)] [[PubMed](#)]
329. Shin, T. The pleiotropic effects of galectin-3 in neuroinflammation: A review. *Acta Histochem.* **2013**, *115*, 407–411. [[CrossRef](#)] [[PubMed](#)]
330. Saccon, F.; Gatto, M.; Ghirardello, A.; Iaccarino, L.; Punzi, L.; Doria, A. Role of galectin-3 in autoimmune and non-autoimmune nephropathies. *Autoimmun. Rev.* **2017**, *16*, 34–47. [[CrossRef](#)] [[PubMed](#)]
331. Satoh, K.; Niwa, M.; Binh, N.H.; Nakashima, M.; Kobayashi, K.; Takamatsu, M.; Hara, A. Increase of galectin-3 expression in microglia by hyperthermia in delayed neuronal death of hippocampal CA1 following transient forebrain ischemia. *Neurosci. Lett.* **2011**, *504*, 199–203. [[CrossRef](#)]
332. Satoh, K.; Niwa, M.; Goda, W.; Binh, N.H.; Nakashima, M.; Takamatsu, M.; Hara, A. Galectin-3 expression in delayed neuronal death of hippocampal CA1 following transient forebrain ischemia, and its inhibition by hypothermia. *Brain Res.* **2011**, *1382*, 266–274. [[CrossRef](#)]
333. Hisamatsu, K.; Niwa, M.; Kobayashi, K.; Miyazaki, T.; Hirata, A.; Hatano, Y.; Tomita, H.; Hara, A. Galectin-3 expression in hippocampal CA2 following transient forebrain ischemia and its inhibition by hypothermia or antiapoptotic agents. *Neuroreport* **2016**, *27*, 311–317. [[CrossRef](#)]
334. Ashraf, G.M.; Baesa, S.S. Investigation of Gal-3 Expression Pattern in Serum and Cerebrospinal Fluid of Patients Suffering From Neurodegenerative Disorders. *Front. Neurosci.* **2018**, *12*, 430. [[CrossRef](#)]
335. Rotshenker, S. The role of Galectin-3/MAC-2 in the activation of the innate-immune function of phagocytosis in microglia in injury and disease. *J. Mol. Neurosci.* **2009**, *39*, 99–103. [[CrossRef](#)]
336. Song, L.; Tang, J.W.; Owusu, L.; Sun, M.Z.; Wu, J.; Zhang, J. Galectin-3 in cancer. *Clin. Chim. Acta* **2014**, *431*, 185–191. [[CrossRef](#)]
337. Fortuna-Costa, A.; Gomes, A.M.; Kozlowski, E.O.; Stelling, M.P.; Pavão, M.S.G. Extracellular galectin-3 in tumor progression and metastasis. *Front. Oncol.* **2014**, *4*, 138. [[CrossRef](#)]
338. Funasaka, T.; Raz, A.; Nangia-Makker, P. Galectin-3 in angiogenesis and metastasis. *Glycobiology* **2014**, *24*, 886–891. [[CrossRef](#)] [[PubMed](#)]
339. Xin, M.; Dong, X.W.; Guo, X.L. Role of the interaction between galectin-3 and cell adhesion molecules in cancer metastasis. *Biomed. Pharmacother.* **2015**, *69*, 179–185. [[CrossRef](#)] [[PubMed](#)]
340. Ruvolo, P.P. Galectin 3 as a guardian of the tumor microenvironment. *Biochim. Biophys. Acta Mol. Cell Res.* **2016**, *1863*, 427–437. [[CrossRef](#)] [[PubMed](#)]
341. Zeinali, M.; Adelinik, A.; Papian, S.; Khorramdelazad, H.; Abedinzadeh, M. Role of galectin-3 in the pathogenesis of bladder transitional cell carcinoma. *Hum. Immunol.* **2015**, *76*, 770–774. [[CrossRef](#)]
342. Wang, L.; Guo, X.L. Molecular regulation of galectin-expression and therapeutic implication in cancer progression. *Biomed. Pharmacother.* **2016**, *78*, 165–171. [[CrossRef](#)]
343. Nangia-Makker, P.; Hogan, V.; Raz, A. Galectin-3 and cancer stemness. *Glycobiology* **2018**, *28*, 172–181. [[CrossRef](#)]
344. Wang, C.; Zhou, X.; Ma, L.; Zhuang, Y.; Wei, Y.; Zhang, L.; Jin, S.; Liang, W.; Shen, X.; Li, C.; et al. Galectin-3 may serve as a marker for poor prognosis in colorectal cancer: A meta-analysis. *Pathol. Res. Pract.* **2019**, *215*, 152612. [[CrossRef](#)]
345. Binh, N.H.; Satoh, K.; Kobayashi, K.; Takamatsu, M.; Hatano, Y.; Hirata, A.; Tomita, H.; Kuno, T.; Hara, A. Galectin-3 in preneoplastic lesions of glioma. *J. Neurooncol.* **2013**, *111*, 123–132. [[CrossRef](#)]
346. Yabluchanskiy, A.; Ma, Y.; Iyer, R.P.; Hall, M.E.; Lindsey, M.L. Matrix metalloproteinase-9: Many shades of function in cardiovascular disease. *Physiology* **2013**, *28*, 391–403. [[CrossRef](#)]
347. Ma, Y.; De Castro Brás, L.E.; Toba, H.; Iyer, R.P.; Hall, M.E.; Winniford, M.D.; Lange, R.A.; Tyagi, S.C.; Lindsey, M.L. Myofibroblasts and the extracellular matrix network in post-myocardial infarction cardiac remodeling. *Pflugers Arch. Eur. J. Physiol.* **2014**, *466*, 1113–1127. [[CrossRef](#)]
348. Ikonomidis, J.S.; Hendrick, J.W.; Parkhurst, A.M.; Herron, A.R.; Escobar, P.G.; Dowdy, K.B.; Stroud, R.E.; Hapke, E.; Zile, M.R.; Spinale, F.G. Accelerated LV remodeling after myocardial infarction in TIMP-1-deficient mice: Effects of exogenous MMP inhibition. *Am. J. Physiol. Heart Circ. Physiol.* **2005**, *288*, 149–158. [[CrossRef](#)] [[PubMed](#)]

349. Rouis, M.; Adamy, C.; Duverger, N.; Lesnik, P.; Horellou, P.; Moreau, M.; Emmanuel, F.; Caillaud, J.M.; Laplaud, P.M.; Dachet, C.; et al. Adenovirus-mediated overexpression of tissue inhibitor of metalloproteinase-1 reduces atherosclerotic lesions in apolipoprotein E-deficient mice. *Circulation* **1999**, *100*, 533–540. [CrossRef] [PubMed]
350. Akahane, T.; Akahane, M.; Shah, A.; Thorgerisson, U.P. TIMP-1 stimulates proliferation of human aortic smooth muscle cells and Ras effector pathways. *Biochem. Biophys. Res. Commun.* **2004**, *324*, 440–445. [CrossRef] [PubMed]
351. Uchinaka, A.; Kawaguchi, N.; Mori, S.; Hamada, Y.; Miyagawa, S.; Saito, A.; Sawa, Y.; Matsuura, N. Tissue inhibitor of metalloproteinase-1 and -3 improves cardiac function in an ischemic cardiomyopathy model rat. *Tissue Eng. Part A* **2014**, *20*, 3073–3084. [CrossRef] [PubMed]
352. Ma, Y.; Chiao, Y.A.; Clark, R.; Flynn, E.R.; Yabluchanskiy, A.; Ghasemi, O.; Zouein, F.; Lindsey, M.L.; Jin, Y.F. Deriving a cardiac ageing signature to reveal MMP-9-dependent inflammatory signalling in senescence. *Cardiovasc. Res.* **2015**, *106*, 421–431. [CrossRef] [PubMed]
353. Zamilpa, R.; Ibarra, J.; de Castro Brás, L.E.; Ramirez, T.A.; Nguyen, N.; Halade, G.V.; Zhang, J.; Dai, Q.; Dayah, T.; Chiao, Y.A.; et al. Transgenic overexpression of matrix metalloproteinase-9 in macrophages attenuates the inflammatory response and improves left ventricular function post-myocardial infarction. *J. Mol. Cell. Cardiol.* **2012**, *53*, 599–608. [CrossRef]
354. Chiao, Y.A.; Ramirez, T.A.; Zamilpa, R.; Okoronkwo, S.M.; Dai, Q.; Zhang, J.; Jin, Y.F.; Lindsey, M.L. Matrix metalloproteinase-9 deletion attenuates myocardial fibrosis and diastolic dysfunction in ageing mice. *Cardiovasc. Res.* **2012**, *96*, 444–455. [CrossRef]
355. Wang, W.; Song, X.; Chen, Y.; Yuan, F.; Xu, F.; Zhang, M.; Tan, K.; Yang, X.; Yu, X.; Lv, S. The Long-Term Influence of Tissue Inhibitor of Matrix Metalloproteinase-1 in Patients with Mild to Moderate Coronary Artery Lesions in a Chinese Population: A 7-Year Follow-Up Study. *Cardiology* **2015**, *132*, 151–158. [CrossRef]
356. Kelly, D.; Squire, I.B.; Khan, S.Q.; Dhillon, O.; Narayan, H.; Ng, K.H.; Quinn, P.; Davies, J.E.; Ng, L.L. Usefulness of plasma tissue inhibitors of metalloproteinases as markers of prognosis after acute myocardial infarction. *Am. J. Cardiol.* **2010**, *106*, 477–482. [CrossRef]
357. Lubos, E.; Schnabel, R.; Rupprecht, H.J.; Bickel, C.; Messow, C.M.; Prigge, S.; Cambien, F.; Tiret, L.; Münzel, T.; Blankenberg, S. Prognostic value of tissue inhibitor of metalloproteinase-1 for cardiovascular death among patients with cardiovascular disease: Results from the AtheroGene study. *Eur. Heart J.* **2006**, *27*, 150–156. [CrossRef]
358. Sundstrom, J. Relations of plasma total TIMP-1 levels to cardiovascular risk factors and echocardiographic measures: The Framingham heart study. *Eur. Heart J.* **2004**, *25*, 1509–1516. [CrossRef] [PubMed]
359. Kormi, I.; Nieminen, M.T.; Havulinna, A.S.; Zeller, T.; Blankenberg, S.; Tervahartiala, T.; Sorsa, T.; Salomaa, V.; Pussinen, P.J. Matrix metalloproteinase-8 and tissue inhibitor of matrix metalloproteinase-1 predict incident cardiovascular disease events and all-cause mortality in a population-based cohort. *Eur. J. Prev. Cardiol.* **2017**, *24*, 1136–1144. [CrossRef] [PubMed]
360. van den Borne, S.W.M.; Cleutjens, J.P.M.; Hanemaaijer, R.; Creemers, E.E.; Smits, J.F.M.; Daemen, M.J.A.P.; Blanksteijn, W.M. Increased matrix metalloproteinase-8 and -9 activity in patients with infarct rupture after myocardial infarction. *Cardiovasc. Pathol.* **2009**, *18*, 37–43. [CrossRef] [PubMed]
361. Blankenberg, S.; Rupprecht, H.J.; Poirier, O.; Bickel, C.; Smieja, M.; Hafner, G.; Meyer, J.; Cambien, F.; Tiret, L. Plasma concentrations and genetic variation of matrix metalloproteinase 9 and prognosis of patients with cardiovascular disease. *Circulation* **2003**, *107*, 1579–1585. [CrossRef]
362. Hamed, G.M.; Fattah, M.F.A. Clinical Relevance of Matrix Metalloproteinase 9 in Patients with Acute Coronary Syndrome. *Clin. Appl. Thromb.* **2015**, *21*, 750–754. [CrossRef]
363. Morishita, T.; Uzui, H.; Mitsu, Y.; Amaya, N.; Kaseno, K.; Ishida, K.; Fukuoka, Y.; Ikeda, H.; Tama, N.; Yamazaki, T.; et al. Association between matrix metalloproteinase-9 and worsening heart failure events in patients with chronic heart failure. *ESC Heart Fail.* **2017**, *4*, 321–330. [CrossRef]
364. Wagner, D.R.; Delagardelle, C.; Ernens, I.; Rouy, D.; Vaillant, M.; Beissel, J. Matrix metalloproteinase-9 is a marker of heart failure after acute myocardial infarction. *J. Card. Fail.* **2006**, *12*, 66–72. [CrossRef]
365. Tan, J.; Hua, Q.; Xing, X.; Wen, J.; Liu, R.; Yang, Z. Impact of the Metalloproteinase-9/Tissue Inhibitor of Metalloproteinase-1 System on Large Arterial Stiffness in Patients with Essential Hypertension. *Hypertens. Res.* **2007**, *30*, 959–963. [CrossRef]
366. Kempf, T.; Eden, M.; Strelau, J.; Naguib, M.; Willenbockel, C.; Tongers, J.; Heineke, J.; Kotlarz, D.; Xu, J.; Molkentin, J.D.; et al. The transforming growth factor- β superfamily member growth-differentiation factor-15 protects the heart from ischemia/reperfusion injury. *Circ. Res.* **2006**, *98*, 351–360. [CrossRef]
367. Xu, J.; Kimball, T.R.; Lorenz, J.N.; Brown, D.A.; Bauskin, A.R.; Klevitsky, R.; Hewett, T.E.; Breit, S.N.; Molkentin, J.D. GDF15/MIC-1 functions as a protective and antihypertrophic factor released from the myocardium in association with SMAD protein activation. *Circ. Res.* **2006**, *98*, 342–350. [CrossRef]
368. de Jager, S.C.A.; Bermúdez, B.; Bot, I.; Koenen, R.R.; Bot, M.; Kavelaars, A.; De Waard, V.; Heijnen, C.J.; Muriana, F.J.G.; Weber, C. Growth differentiation factor 15 deficiency protects against atherosclerosis by attenuating CCR2-mediated macrophage chemotaxis. *J. Exp. Med.* **2011**, *208*, 217–225. [CrossRef] [PubMed]
369. Bonaterra, G.A.; Zügel, S.; Thogersen, J.; Walter, S.A.; Haberkorn, U.; Strelau, J.; Kinscherf, R. Growth differentiation factor-15 deficiency inhibits atherosclerosis progression by regulating interleukin-6-dependent inflammatory response to vascular injury. *J. Am. Heart Assoc.* **2012**, *1*, e002550. [CrossRef] [PubMed]
370. Murielle, M.; Batra, S.K. Divergent molecular mechanisms underlying the pleiotropic functions of macrophage inhibitory cytokine-1 in cancer. *J. Cell. Physiol.* **2010**, *224*, 626–635.

371. Tsai, V.W.W.; Lin, S.; Brown, D.A.; Salis, A.; Breit, S.N. Anorexia-cachexia and obesity treatment may be two sides of the same coin: Role of the TGF- β superfamily cytokine MIC-1/GDF15. *Int. J. Obes.* **2016**, *40*, 193–197. [CrossRef] [PubMed]
372. Kempf, T.; Zarbock, A.; Widera, C.; Butz, S.; Stadtmann, A.; Rossaint, J.; Bolomini-Vittori, M.; Korf-Klingebiel, M.; Napp, L.C.; Hansen, B.; et al. GDF-15 is an inhibitor of leukocyte integrin activation required for survival after myocardial infarction in mice. *Nat. Med.* **2011**, *17*, 581–588. [CrossRef]
373. Wollert, K.C.; Kempf, T.; Peter, T.; Olofsson, S.; James, S.; Johnston, N.; Lindahl, B.; Horn-Wichmann, R.; Brabant, G.; Simoons, M.L.; et al. Prognostic value of growth-differentiation factor-15 in patients with non-ST-elevation acute coronary syndrome. *Circulation* **2007**, *115*, 962–971. [CrossRef]
374. Lok, S.I.; Winkens, B.; Goldschmeding, R.; Van Geffen, A.J.P.; Nous, F.M.A.; Van Kuik, J.; Van Der Weide, P.; Klöpping, C.; Kirkels, J.H.; Lahpor, J.R.; et al. Circulating growth differentiation factor-15 correlates with myocardial fibrosis in patients with non-ischaemic dilated cardiomyopathy and decreases rapidly after left ventricular assist device support. *Eur. J. Heart Fail.* **2012**, *14*, 1249–1256. [CrossRef]
375. Kempf, T.; von Haehling, S.; Peter, T.; Allhoff, T.; Cicora, M.; Doehner, W.; Ponikowski, P.; Filippatos, G.S.; Rozentryt, P.; Drexler, H.; et al. Prognostic Utility of Growth Differentiation Factor-15 in Patients with Chronic Heart Failure. *J. Am. Coll. Cardiol.* **2007**, *50*, 1054–1060. [CrossRef]
376. Brown, D.A.; Breit, S.N.; Buring, J.; Fairlie, W.D.; Bauskin, A.R.; Liu, T.; Ridker, P.M. Concentration in plasma of macrophage inhibitory cytokine-1 and risk of cardiovascular events in women: A nested case-control study. *Lancet* **2002**, *359*, 2159–2163. [CrossRef]
377. Lind, L.; Wallentin, L.; Kempf, T.; Tapken, H.; Quint, A.; Lindahl, B.; Olofsson, S.; Venge, P.; Larsson, A.; Hulthe, J.; et al. Growth-differentiation factor-15 is an independent marker of cardiovascular dysfunction and disease in the elderly: Results from the Prospective Investigation of the Vasculature in Uppsala Seniors (PIVUS) study. *Eur. Heart J.* **2009**, *30*, 2346–2353. [CrossRef]
378. Daniels, L.B.; Clopton, P.; Laughlin, G.A.; Maisel, A.S.; Barrett-Connor, E. Growth-Differentiation Factor-15 Is a Robust, Independent Predictor of 11-Year Mortality Risk in Community-Dwelling Older Adults. *Circulation* **2011**, *123*, 2101–2110. [CrossRef] [PubMed]
379. Ho, J.E.; Mahajan, A.; Chen, M.-H.; Larson, M.G.; McCabe, E.L.; Ghorbani, A.; Cheng, S.; Johnson, A.D.; Lindgren, C.M.; Kempf, T.; et al. Clinical and Genetic Correlates of Growth Differentiation Factor 15 in the Community. *Clin. Chem.* **2012**, *58*, 1582–1591. [CrossRef] [PubMed]
380. Rohatgi, A.; Patel, P.; Das, S.R.; Ayers, C.R.; Khera, A.; Martinez-Rumayor, A.; Berry, J.D.; McGuire, D.K.; de Lemos, J.A. Association of Growth Differentiation Factor-15 with Coronary Atherosclerosis and Mortality in a Young, Multiethnic Population: Observations from the Dallas Heart Study. *Clin. Chem.* **2012**, *58*, 172–182. [CrossRef] [PubMed]
381. Wiklund, F.E.; Bennet, A.M.; Magnusson, P.K.E.; Eriksson, U.K.; Lindmark, F.; Wu, L.; Yaghoutyfam, N.; Marquis, C.P.; Stattin, P.; Pedersen, N.L.; et al. Macrophage inhibitory cytokine-1 (MIC-1/GDF15): A new marker of all-cause mortality. *Aging Cell* **2010**, *9*, 1057–1064. [CrossRef]
382. Andersson, C.; Enserro, D.; Sullivan, L.; Wang, T.J.; Januzzi, J.L.; Benjamin, E.J.; Vita, J.A.; Hamburg, N.M.; Larson, M.G.; Mitchell, G.F.; et al. Relations of circulating GDF-15, soluble ST2, and troponin-I concentrations with vascular function in the community: The Framingham Heart Study. *Atherosclerosis* **2016**, *248*, 245–251. [CrossRef]
383. Xanthakos, V.; Larson, M.G.; Wollert, K.C.; Aragam, J.; Cheng, S.; Ho, J.; Coglianese, E.; Levy, D.; Colucci, W.S.; Michael Felker, G.; et al. Association of novel biomarkers of cardiovascular stress with left ventricular hypertrophy and dysfunction: Implications for screening. *J. Am. Heart Assoc.* **2013**, *2*, e000399. [CrossRef]
384. Kempf, T.; Horn-Wichmann, R.; Brabant, G.; Peter, T.; Allhoff, T.; Klein, G.; Drexler, H.; Johnston, N.; Wallentin, L.; Wollert, K.C. Circulating concentrations of growth-differentiation factor 15 in apparently healthy elderly individuals and patients with chronic heart failure as assessed by a new immunoradiometric sandwich assay. *Clin. Chem.* **2007**, *53*, 284–291. [CrossRef]
385. Wallentin, L.; Zethelius, B.; Berglund, L.; Eggers, K.M.; Lind, L.; Lindahl, B.; Wollert, K.C.; Siegbahn, A. GDF-15 for prognostication of cardiovascular and cancer morbidity and mortality in men. *PLoS ONE* **2013**, *8*, e78797. [CrossRef]
386. Hagström, E.; James, S.K.; Bertilsson, M.; Becker, R.C.; Himmelmann, A.; Husted, S.; Katus, H.A.; Steg, P.G.; Storey, R.F.; Siegbahn, A.; et al. Growth differentiation factor-15 level predicts major bleeding and cardiovascular events in patients with acute coronary syndromes: Results from the PLATO study. *Eur. Heart J.* **2016**, *37*, 1325–1333. [CrossRef]
387. Kempf, T.; Björklund, E.; Olofsson, S.; Lindahl, B.; Allhoff, T.; Peter, T.; Tongers, J.; Wollert, K.C.; Wallentin, L. Growth-differentiation factor-15 improves risk stratification in ST-segment elevation myocardial infarction. *Eur. Heart J.* **2007**, *28*, 2858–2865. [CrossRef]
388. Schopfer, D.W.; Ku, I.A.; Regan, M.; Whooley, M.A. Growth differentiation factor 15 and cardiovascular events in patients with stable ischemic heart disease (The Heart and Soul Study). *Am. Heart J.* **2014**, *167*, 186–192.e1. [CrossRef] [PubMed]
389. Bonaca, M.P.; Morrow, D.A.; Braunwald, E.; Cannon, C.P.; Jiang, S.; Breher, S.; Sabatine, M.S.; Kempf, T.; Wallentin, L.; Wollert, K.C. Growth differentiation factor-15 and risk of recurrent events in patients stabilized after acute coronary syndrome: Observations from PROVE IT-TIMI 22. *Arterioscler. Thromb. Vasc. Biol.* **2011**, *31*, 203–210. [CrossRef] [PubMed]
390. Anand, I.S.; Kempf, T.; Rector, T.S.; Tapken, H.; Allhoff, T.; Jantzen, F.; Kuskowski, M.; Cohn, J.N.; Drexler, H.; Wollert, K.C. Serial measurement of growth-differentiation factor-15 in heart failure: Relation to disease severity and prognosis in the valsartan heart failure trial. *Circulation* **2010**, *122*, 1387–1395. [CrossRef] [PubMed]

391. Chan, M.M.Y.; Santhanakrishnan, R.; Chong, J.P.C.; Chen, Z.; Tai, B.C.; Liew, O.W.; Ng, T.P.; Ling, L.H.; Sim, D.; Leong, K.T.G.; et al. Growth differentiation factor 15 in heart failure with preserved vs. reduced ejection fraction. *Eur. J. Heart Fail.* **2016**, *18*, 81–88. [[CrossRef](#)] [[PubMed](#)]
392. Cotter, G.; Voors, A.A.; Prescott, M.F.; Felker, G.M.; Filippatos, G.; Greenberg, B.H.; Pang, P.S.; Ponikowski, P.; Milo, O.; Hua, T.A.; et al. Growth differentiation factor 15 (GDF-15) in patients admitted for acute heart failure: Results from the RELAX-AHF study. *Eur. J. Heart Fail.* **2015**, *17*, 1133–1143. [[CrossRef](#)]
393. Hijazi, Z.; Oldgren, J.; Lindbäck, J.; Alexander, J.H.; Connolly, S.J.; Eikelboom, J.W.; Ezekowitz, M.D.; Held, C.; Hylek, E.M.; Lopes, R.D.; et al. The novel biomarker-based ABC (age, biomarkers, clinical history)-bleeding risk score for patients with atrial fibrillation: A derivation and validation study. *Lancet* **2016**, *387*, 2302–2311. [[CrossRef](#)]
394. Hijazi, Z.; Lindbäck, J.; Alexander, J.H.; Hanna, M.; Held, C.; Hylek, E.M.; Lopes, R.D.; Oldgren, J.; Siegbahn, A.; Stewart, R.A.H.; et al. The ABC (age, biomarkers, clinical history) stroke risk score: A biomarker-based risk score for predicting stroke in atrial fibrillation. *Eur. Heart J.* **2016**, *37*, 1582–1590. [[CrossRef](#)]
395. Hijazi, Z.; Oldgren, J.; Lindbäck, J.; Alexander, J.H.; Connolly, S.J.; Eikelboom, J.W.; Ezekowitz, M.D.; Held, C.; Hylek, E.M.; Lopes, R.D.; et al. A biomarker-based risk score to predict death in patients with atrial fibrillation: The ABC (age, biomarkers, clinical history) death risk score. *Eur. Heart J.* **2018**, *39*, 477–485. [[CrossRef](#)]
396. Wallentin, L.; Hijazi, Z.; Andersson, U.; Alexander, J.H.; De Caterina, R.; Hanna, M.; Horowitz, J.D.; Hylek, E.M.; Lopes, R.D. Growth differentiation factor 15, a marker of oxidative stress and inflammation, for risk assessment in patients with atrial fibrillation: Insights from the Apixaban for reduction in stroke and other thromboembolic events in atrial fibrillation (ARISTOTLE) trial. *Circulation* **2014**, *130*, 1847–1858.
397. Hijazi, Z.; Oldgren, J.; Andersson, U.; Connolly, S.J.; Eikelboom, J.W.; Ezekowitz, M.D.; Reilly, P.A.; Yusuf, S.; Siegbahn, A.; Wallentin, L. Growth-differentiation factor 15 and risk of major bleeding in atrial fibrillation: Insights from the Randomized Evaluation of Long-Term Anticoagulation Therapy (RE-LY) trial. *Am. Heart J.* **2017**, *190*, 94–103. [[CrossRef](#)]
398. Widera, C.; Pencina, M.J.; Bobadilla, M.; Reimann, I.; Guba-Quint, A.; Marquardt, I.; Bethmann, K.; Korf-Klingebiel, M.; Kempf, T.; Lichtinghagen, R.; et al. Incremental prognostic value of biomarkers beyond the GRACE (Global Registry of Acute Coronary Events) score and high-sensitivity cardiac troponin T in non-ST-elevation acute coronary syndrome. *Clin. Chem.* **2013**, *59*, 1497–1505. [[CrossRef](#)] [[PubMed](#)]
399. Widera, C.; Pencina, M.J.; Meisner, A.; Kempf, T.; Bethmann, K.; Marquardt, I.; Katus, H.A.; Giannitsis, E.; Wollert, K.C. Adjustment of the GRACE score by growth differentiation factor 15 enables a more accurate appreciation of risk in non-ST-elevation acute coronary syndrome. *Eur. Heart J.* **2012**, *33*, 1095–1104. [[CrossRef](#)] [[PubMed](#)]
400. Ho, J.E.; Hwang, S.J.; Wollert, K.C.; Larson, M.G.; Cheng, S.; Kempf, T.; Vasan, R.S.; Januzzi, J.L.; Wang, T.J.; Fox, C.S. Biomarkers of cardiovascular stress and incident chronic kidney disease. *Clin. Chem.* **2013**, *59*, 1613–1620. [[CrossRef](#)] [[PubMed](#)]
401. Saucerman, J.J.; Tan, P.M.; Buchholz, K.S.; McCulloch, A.D.; Omens, J.H. Mechanical regulation of gene expression in cardiac myocytes and fibroblasts. *Nat. Rev. Cardiol.* **2019**, *16*, 361–378. [[CrossRef](#)] [[PubMed](#)]
402. Vila, G.; Riedl, M.; Anderwald, C.; Resl, M.; Handisurya, A.; Clodi, M.; Prager, G.; Ludvik, B.; Krebs, M.; Luger, A. The relationship between insulin resistance and the cardiovascular biomarker growth differentiation factor-15 in obese patients. *Clin. Chem.* **2011**, *57*, 309–316. [[CrossRef](#)]
403. Farhan, S.; Freynhofer, M.K.; Brozovic, I.; Bruno, V.; Vogel, B.; Tentzeris, I.; Baumgartner-Parzer, S.; Huber, K.; Kautzky-Willer, A. Determinants of growth differentiation factor 15 in patients with stable and acute coronary artery disease. A prospective observational study. *Cardiovasc. Diabetol.* **2016**, *15*, 60. [[CrossRef](#)]
404. Schaub, N.; Reichlin, T.; Twerenbold, R.; Reiter, M.; Steuer, S.; Bassetti, S.; Stelzig, C.; Wolf, C.; Winkler, K.; Haaf, P.; et al. Growth differentiation factor-15 in the early diagnosis and risk stratification of patients with acute chest pain. *Clin. Chem.* **2012**, *58*, 441–449. [[CrossRef](#)]
405. Mueller, T.; Leitner, I.; Egger, M.; Haltmayer, M.; Dieplinger, B. Association of the biomarkers soluble ST2, galectin-3 and growth-differentiation factor-15 with heart failure and other non-cardiac diseases. *Clin. Chim. Acta* **2015**, *445*, 155–160. [[CrossRef](#)]
406. Dhaun, N.; Webb, D.J. Endothelins in cardiovascular biology and therapeutics. *Nat. Rev. Cardiol.* **2019**, *16*, 491–502. [[CrossRef](#)]
407. Yanagisawa, M.; Kurihara, H.; Kimura, S.; Tomobe, Y.; Kobayashi, M.; Mitsui, Y.; Yazaki, Y.; Goto, K.; Masaki, T. A novel potent vasoconstrictor peptide produced by vascular endothelial cells. *Nature* **1988**, *332*, 411–415. [[CrossRef](#)]
408. Pollock, D.M. 2013 Dahl Lecture American heart association council for high blood pressure research clarifying the physiology of endothelin. *Hypertension* **2014**, *63*, 110–117. [[CrossRef](#)] [[PubMed](#)]
409. Hunter, R.W.; Moorhouse, R.; Farrah, T.E.; MacIntyre, I.M.; Asai, T.; Gallacher, P.J.; Kerr, D.; Melville, V.; Czopek, A.; Morrison, E.E.; et al. First-in-Man Demonstration of Direct Endothelin-Mediated Natriuresis and Diuresis. *Hypertension* **2017**, *70*, 192–200. [[CrossRef](#)] [[PubMed](#)]
410. Haynes, W.G.; Webb, D.J. Contribution of endogenous generation of endothelin-1 to basal vascular tone. *Lancet* **1994**, *344*, 852–854. [[CrossRef](#)]
411. Morrell, N.; Suntharalingam, J. Endothelins. *Encycl. Respir. Med. Four-Volume Set* **2006**, *95499*, 75–79.
412. van Wamel, A.J.E.T.; Ruwhof, C.; van der Valk-Kokshoorn, L.E.J.M.; Schriern, P.I.; van der Laarse, A. The role of angiotensin II, endothelin-1 and transforming growth factor- β as autocrine/paracrine mediators of stretch-induced cardiomyocyte hypertrophy. *Mol. Cell. Biochem.* **2001**, *218*, 113–124. [[CrossRef](#)] [[PubMed](#)]

413. Bhandari, S.S.; Davies, J.E.; Struck, J.; Ng, L.L. Plasma C-terminal proEndothelin-1 (CTproET-1) is affected by age, renal function, left atrial size and diastolic blood pressure in healthy subjects. *Peptides* **2014**, *52*, 53–57. [[CrossRef](#)]
414. Jankowich, M.D.; Wu, W.C.; Choudhary, G. Association of elevated plasma endothelin-1 levels with pulmonary hypertension, mortality, and heart failure in African American Individuals: The jackson heart study. *JAMA Cardiol.* **2016**, *1*, 461–469. [[CrossRef](#)]
415. Hirai, Y.; Adachi, H.; Fujiura, Y.; Hiratsuka, A.; Enomoto, M.; Imaizumi, T. Plasma endothelin-1 level is related to renal function and smoking status but not to blood pressure: An epidemiological study. *J. Hypertens.* **2004**, *22*, 713–718. [[CrossRef](#)]
416. Yokoi, K.; Adachi, H.; Hirai, Y.; Enomoto, M.; Fukami, A.; Ogata, K.; Tsukagawa, E.; Kasahara, A.; Imaizumi, T. Plasma endothelin-1 level is a predictor of 10-year mortality in a general population: The tanushimaru study. *Circ. J.* **2012**, *76*, 2779–2784. [[CrossRef](#)]
417. Papassotiriou, J.; Morgenthaler, N.G.; Struck, J.; Alonso, C.; Bergmann, A. Immunoluminometric assay for measurement of the C-terminal endothelin-I precursor fragment in human plasma. *Clin. Chem.* **2006**, *52*, 1144–1151. [[CrossRef](#)]
418. Jankowich, M.; Elston, B.; Liu, Q.; Abbasi, S.; Wu, W.C.; Blackshear, C.; Godfrey, M.; Choudhary, G. Restrictive spirometry pattern, cardiac structure and function, and incident heart failure in African Americans. The Jackson heart study. *Ann. Am. Thorac. Soc.* **2018**, *15*, 1186–1196. [[CrossRef](#)] [[PubMed](#)]
419. Oelsner, E.C.; Pottinger, T.D.; Burkart, K.M.; Allison, M.; Buxbaum, S.G.; Hansel, N.N.; Kumar, R.; Larkin, E.K.; Lange, L.A.; Loehr, L.R.; et al. Adhesion molecules, endothelin-1 and lung function in seven population-based cohorts. *Biomarkers* **2013**, *18*, 196–203. [[CrossRef](#)] [[PubMed](#)]
420. Maeder, M.T.; Brutsche, M.H.; Arenja, N.; Socrates, T.; Reiter, M.; Meissner, J.; Staub, D.; Morgenthaler, N.G.; Bergmann, A.; Mueller, C. Biomarkers and peak oxygen uptake in patients with chronic lung disease. *Respiration* **2010**, *80*, 543–552. [[CrossRef](#)] [[PubMed](#)]
421. Dhaun, N.; Yuzugulen, J.; Kimmitt, R.A.; Wood, E.G.; Chariyavilaskul, P.; MacIntyre, I.M.; Goddard, J.; Webb, D.J.; Corder, R. Plasma pro-endothelin-1 peptide concentrations rise in chronic kidney disease and following selective endothelin A receptor antagonism. *J. Am. Heart Assoc.* **2015**, *4*, e001624. [[CrossRef](#)]
422. Pavo, N.; Hülsmann, M.; Neuhold, S.; Adlbrecht, C.; Strunk, G.; Goliasch, G.; Gisslinger, H.; Steger, G.G.; Hejna, M.; Köstler, W.; et al. Cardiovascular biomarkers in patients with cancer and their association with all-cause mortality. *Heart* **2015**, *101*, 1874–1880. [[CrossRef](#)]
423. Wang, Y.; Tang, Y.; Zou, Y.; Wang, D.; Zhu, L.; Tian, T.; Wang, J.; Bao, J.; Hui, R.; Kang, L.; et al. Plasma level of big endothelin-1 predicts the prognosis in patients with hypertrophic cardiomyopathy. *Int. J. Cardiol.* **2017**, *243*, 283–289. [[CrossRef](#)]
424. Gupta, R.M.; Hadaya, J.; Trehan, A.; Zekavat, S.M.; Roselli, C.; Klarin, D.; Emdin, C.A.; Hilvering, C.R.E.; Bianchi, V.; Mueller, C.; et al. A Genetic Variant Associated with Five Vascular Diseases Is a Distal Regulator of Endothelin-1 Gene Expression. *Cell* **2017**, *170*, 522–533.e15. [[CrossRef](#)]
425. Perez, A.L.; Grodin, J.L.; Wu, Y.; Hernandez, A.F.; Butler, J.; Metra, M.; Felker, G.M.; Voors, A.A.; McMurray, J.J.; Armstrong, P.W.; et al. Increased mortality with elevated plasma endothelin-1 in acute heart failure: An ASCEND-HF biomarker substudy. *Eur. J. Heart Fail.* **2016**, *18*, 290–297. [[CrossRef](#)]
426. Weitzberg, E.; Ahlborg, G.; Lundberg, J.M. Long-lasting vasoconstriction and efficient regional extraction of endothelin-1 in human splanchnic and renal tissues. *Biochem. Biophys. Res. Commun.* **1991**, *180*, 1298–1303. [[CrossRef](#)]
427. Vierhapper, H.; Wagner, O.; Nowotny, P.; Waldhäusl, W. Effect of endothelin-1 in man. *Circulation* **1990**, *81*, 1415–1418. [[CrossRef](#)]
428. Parker, J.D.; Thiessen, J.J.; Reilly, R.; Tong, J.H.; Stewart, D.J.; Pandey, A.S. Human endothelin-1 clearance kinetics revealed by a radiotracer technique. *J. Pharmacol. Exp. Ther.* **1999**, *289*, 261–265. [[PubMed](#)]
429. Khan, S.Q.; Dhillon, O.; Struck, J.; Quinn, P.; Morgenthaler, N.G.; Squire, I.B.; Davies, J.E.; Bergmann, A.; Ng, L.L. C-terminal pro-endothelin-1 offers additional prognostic information in patients after acute myocardial infarction. Leicester Acute Myocardial Infarction Peptide (LAMP) Study. *Am. Heart J.* **2007**, *154*, 736–742. [[CrossRef](#)] [[PubMed](#)]
430. Adlbrecht, C.; Hülsmann, M.; Strunk, G.; Berger, R.; Mörtl, D.; Struck, J.; Morgenthaler, N.G.; Bergmann, A.; Jakowitsch, J.; Maurer, G.; et al. Prognostic value of plasma midregional pro-adrenomedullin and C-terminal-pro-endothelin-1 in chronic heart failure outpatients. *Eur. J. Heart Fail.* **2009**, *11*, 361–366. [[CrossRef](#)] [[PubMed](#)]
431. Kakkar, R.; Lee, R.T. The IL-33/ST2 pathway: Therapeutic target and novel biomarker. *Nat. Rev. Drug Discov.* **2008**, *7*, 827–840. [[CrossRef](#)]
432. Pusceddu, I.; Dieplinger, B.; Mueller, T. ST2 and the ST2/IL-33 signalling pathway—biochemistry and pathophysiology in animal models and humans. *Clin. Chim. Acta* **2019**, *495*, 493–500. [[CrossRef](#)]
433. Pascual-Figal, D.A.; Januzzi, J.L. The biology of ST2: The international ST2 consensus panel. *Am. J. Cardiol.* **2015**, *115*, n3B–7B. [[CrossRef](#)]
434. Ciccone, M.M.; Cortese, F.; Gesualdo, M.; Riccardi, R.; Di Nunzio, D.; Moncelli, M.; Iacoviello, M.; Scicchitano, P. A novel cardiac bio-marker: ST2: A review. *Molecules* **2013**, *18*, 15314–15328. [[CrossRef](#)]
435. Aimo, A.; Vergaro, G.; Ripoli, A.; Bayes-Genis, A.; Pascual Figal, D.A.; de Boer, R.A.; Lassus, J.; Mebazaa, A.; Gayat, E.; Breidthardt, T.; et al. Meta-Analysis of Soluble Suppression of Tumorigenicity-2 and Prognosis in Acute Heart Failure. *JACC Heart Fail.* **2017**, *5*, 287–296. [[CrossRef](#)]
436. Lupón, J.; Simpson, J.; McMurray, J.J.V.; de Antonio, M.; Vila, J.; Subirana, I.; Barallat, J.; Moliner, P.; Domingo, M.; Zamora, E.; et al. Barcelona Bio-HF Calculator Version 2.0: Incorporation of angiotensin II receptor blocker neprilysin inhibitor (ARNI) and risk for heart failure hospitalization. *Eur. J. Heart Fail.* **2018**, *20*, 938–940. [[CrossRef](#)]

437. Bayes-Genis, A.; Zhang, Y.; Ky, B. ST2 and patient prognosis in chronic heart failure. *Am. J. Cardiol.* **2015**, *115*, 64B–69B. [CrossRef]
438. Weinberg, E.O.; Shimpo, M.; Hurwitz, S.; Tominaga, S.i.; Rouleau, J.L.; Lee, R.T. Identification of serum soluble ST2 receptor as a novel heart failure biomarker. *Circulation* **2003**, *107*, 721–726. [CrossRef] [PubMed]
439. Weinberg, E.O.; Shimpo, M.; De Keulenaer, G.W.; MacGillivray, C.; Tominaga, S.; Ichi Solomon, S.D.; Rouleau, J.L.; Lee, R.T. Expression and regulation of ST2, an interleukin-1 receptor family member, in cardiomyocytes and myocardial infarction. *Circulation* **2002**, *106*, 2961–2966. [CrossRef] [PubMed]
440. Aimo, A.; Vergaro, G.; Passino, C.; Ripoli, A.; Ky, B.; Miller, W.L.; Bayes-Genis, A.; Anand, I.; Januzzi, J.L.; Emdin, M. Prognostic Value of Soluble Suppression of Tumorigenicity-2 in Chronic Heart Failure. *JACC Heart Fail.* **2017**, *5*, 280–286. [CrossRef] [PubMed]
441. Pérez-Martínez, M.T.; Vergaro, G.; Passino, C.; Ripoli, A.; Ky, B.; Miller, W.L.; Bayes-Genis, A.; Anand, I.; Januzzi, J.L.; Emdin, M. Noncardiac Production of Soluble ST2 in ST-Segment Elevation Myocardial Infarction. *J. Am. Coll. Cardiol.* **2018**, *72*, 1429–1430. [CrossRef] [PubMed]
442. Tseng, C.A.; Lee, C.P.; Huang, Y.J.; Pang, H.W.; Ho, K.C.; Chen, Y.T. One-step synthesis of graphene hollow nanoballs with various nitrogen-doped states for electrocatalysis in dye-sensitized solar cells. *Mater. Today Energy* **2018**, *8*, 15–21. [CrossRef]
443. Tseng, C.C.S.; Huibers, M.M.H.; van Kuik, J.; de Weger, R.A.; Vink, A.; de Jonge, N. The Interleukin-33/ST2 Pathway Is Expressed in the Failing Human Heart and Associated with Pro-fibrotic Remodeling of the Myocardium. *J. Cardiovasc. Transl. Res.* **2018**, *11*, 15–21. [CrossRef]
444. Bartunek, J.; Delrue, L.; Van Durme, F.; Muller, O.; Casselman, F.; De Wiest, B.; Croes, R.; Verstreken, S.; Goethals, M.; de Raedt, H.; et al. Nonmyocardial Production of ST2 Protein in Human Hypertrophy and Failure Is Related to Diastolic Load. *J. Am. Coll. Cardiol.* **2008**, *52*, 2166–2174. [CrossRef]
445. Vittos, O.; Toana, B.; Vittos, A.; Moldoveanu, E. Lipoprotein-associated phospholipase A2 (Lp-PLA2): A review of its role and significance as a cardiovascular biomarker. *Biomarkers* **2012**, *17*, 289–302. [CrossRef]
446. Caslake, M.J.; Packard, C.J.; Suckling, K.E.; Holmes, S.D.; Chamberlain, P.; Macphee, C.H. Lipoprotein-associated phospholipase A2, platelet-activating factor acetylhydrolase: A potential new risk factor for coronary artery disease. *Atherosclerosis* **2000**, *150*, 413–419. [CrossRef]
447. Packard, C.J.; O'Reilly, D.S.J.; Caslake, M.J.; McMahon, A.D.; Ford, I.; Cooney, J.; Macphee, C.H.; Suckling, K.E.; Krishna, M.; Wilkinson, F.E.; et al. Lipoprotein-Associated Phospholipase A 2 as an Independent Predictor of Coronary Heart Disease. *N. Engl. J. Med.* **2000**, *343*, 1148–1155. [CrossRef]
448. Oei, H.H.S.; Van Der Meer, I.M.; Hofman, A.; Koudstaal, P.J.; Stijnen, T.; Breteler, M.M.B.; Witteman, J.C.M. Lipoprotein-associated phospholipase A2 activity is associated with risk of coronary heart disease and ischemic stroke: The Rotterdam Study. *Circulation* **2005**, *111*, 570–575. [CrossRef] [PubMed]
449. Gerber, Y.; McConnell, J.P.; Jaffe, A.S.; Weston, S.A.; Killian, J.M.; Roger, V.L. Lipoprotein-associated phospholipase A2 and prognosis after myocardial infarction in the community. *Arterioscler. Thromb. Vasc. Biol.* **2006**, *26*, 2517–2522. [CrossRef] [PubMed]
450. André, P.; Nannizzi-Alaimo, L.; Prasad, S.K.; Phillips, D.R. Platelet-derived CD40L: The switch-hitting player of cardiovascular disease. *Circulation* **2002**, *106*, 896–899. [CrossRef]
451. Antoniades, C.; Bakogiannis, C.; Tousoulis, D.; Antonopoulos, A.S.; Stefanidis, C. The CD40/CD40 Ligand System. Linking Inflammation with Atherothrombosis. *J. Am. Coll. Cardiol.* **2009**, *54*, 669–677. [CrossRef] [PubMed]
452. Lutgens, E.; Gorelik, L.; Daemen, M.J.A.P.; De Muinck, E.D.; Grewal, I.S.; Koteliansky, V.E.; Flavell, R.A. Requirement for CD154 in the progression of atherosclerosis. *Nat. Med.* **1999**, *5*, 1313–1316. [CrossRef] [PubMed]
453. Varo, N.; De Lemos, J.A.; Libby, P.; Morrow, D.A.; Murphy, S.A.; Nuzzo, R.; Gibson, C.M.; Cannon, C.P.; Braunwald, E.; Schönbeck, U. Soluble CD40L: Risk prediction after acute coronary syndromes. *Circulation* **2003**, *108*, 1049–1052. [CrossRef] [PubMed]
454. Pereira-da-Silva, T.; Napoleao, P.; Pinheiro, T.; Selas, M.; Silva, F.; Ferreira, R.C.; Carmo, M.M. Inflammation is associated with the presence and severity of chronic coronary syndrome through soluble CD40 ligand. *Am. J. Cardiovasc. Dis.* **2020**, *10*, 329–339.
455. Romaine, S.P.R.; Tomaszewski, M.; Condorelli, G.; Samani, N.J. MicroRNAs in cardiovascular disease: An introduction for clinicians. *Heart* **2015**, *101*, 921–928. [CrossRef]
456. Bargiel, W.; Cierpiszewska, K.; Maruszczak, K.; Pakuła, A.; Szwankowska, D.; Wrzesińska, A.; Gutowski, Ł.; Formanowicz, D. Recognized and potentially new biomarkers—their role in diagnosis and prognosis of cardiovascular disease. *Medicina* **2021**, *57*, 701. [CrossRef]
457. Wang, K.; Gao, X.Q.; Wang, T.; Zhou, L.Y. The Function and Therapeutic Potential of Circular RNA in Cardiovascular Diseases. *Cardiovasc. Drugs Ther.* **2021**. [CrossRef]
458. McAloon, C.J.; Ali, D.; Hamborg, T.; Banerjee, P.; O'Hare, P.; Randeva, H.; Osman, F. Extracellular cardiac matrix biomarkers in patients with reduced ejection fraction heart failure as predictors of response to cardiac resynchronisation therapy: A systematic review. *Open Heart* **2017**, *4*, e000639. [CrossRef] [PubMed]
459. McAloon, C.J.; Barwari, T.; Hu, J.; Hamborg, T.; Nevill, A.; Hyndman, S.; Ansell, V.; Musa, A.; Jones, J.; Goodby, J.; et al. Characterisation of circulating biomarkers before and after cardiac resynchronisation therapy and their role in predicting CRT response: The COVERT-HF study. *Open Heart* **2018**, *5*, e000899. [CrossRef] [PubMed]
460. Sack, G.H., Jr. Serum amyloid A—A review. *Mol. Med.* **2018**, *24*, 46. [CrossRef] [PubMed]

461. Ciccone, M.M.; Scicchitano, P.; Gesualdo, M.; Zito, A.; Carbonara, R.; Locorotondo, M.; Mandurino, C.; Masi, F.; Boccalini, F.; Lepera, M.E. Serum osteoprotegerin and carotid intima-media thickness in acute/chronic coronary artery diseases. *J. Cardiovasc Med.* **2013**, *14*, 43–48. [CrossRef] [PubMed]
462. Montagnana, M.; Lippi, G.; Danese, E.; Guidi, G.C. The role of osteoprotegerin in cardiovascular disease. *Ann. Med.* **2013**, *45*, 254–264. [CrossRef] [PubMed]
463. Silvestre-Roig, C.; Braster, Q.; Ortega-Gomez, A.; Soehnlein, O. Neutrophils as regulators of cardiovascular inflammation. *Nat. Rev. Cardiol.* **2020**, *17*, 327–340. [CrossRef]
464. NICE. Diagnosis and management of chronic heart failure in adults. *Natl. Inst. Health Care Excell.* **2018**, *75*, 742–753.
465. Yancy, C.W.; Jessup, M.; Bozkurt, B.; Butler, J.; Casey, D.E.; Colvin, M.M.; Drazner, M.H.; Filippatos, G.S.; Fonarow, G.C.; Givertz, M.M.; et al. 2017 ACC/AHA/HFSA Focused Update of the 2013 ACCF/AHA Guideline for the Management of Heart Failure: A Report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines and the Heart Failure Society of America. *J. Am. Coll. Cardiol.* **2017**, *70*, 776–803.
466. Lazzerini, P.E.; Capecchi, P.L.; Laghi-Pasini, F. Systemic inflammation and arrhythmic risk: Lessons from rheumatoid arthritis. *Eur. Heart J.* **2017**, *38*, 1717–1727. [CrossRef]
467. Issac, T.T.; Dokainish, H.; Lakkis, N.M. Role of Inflammation in Initiation and Perpetuation of Atrial Fibrillation. A Systematic Review of the Published Data. *J. Am. Coll. Cardiol.* **2007**, *50*, 2021–2028. [CrossRef]
468. Scott, L.; Li, N.; Dobrev, D. Role of inflammatory signaling in atrial fibrillation. *Int. J. Cardiol.* **2019**, *287*, 195–200. [CrossRef] [PubMed]
469. Melenovsky, V.; Lip, G.Y.H. Interleukin-8 and atrial fibrillation. *Europace* **2008**, *10*, 784–785. [CrossRef] [PubMed]
470. Guan, H.; Liu, J.; Ding, J.; Liu, W.; Feng, Y.; Bao, Y.; Li, H.; Wang, X.; Zhou, Z.; Chen, Z. Arrhythmias in patients with coronavirus disease 2019 (COVID-19) in Wuhan, China: Incidences and implications. *J. Electrocardiol.* **2021**, *65*, 96–101. [CrossRef] [PubMed]
471. Shah, A.S.V.; Anand, A.; Sandoval, Y.; Lee, K.K.; Smith, S.W.; Adamson, P.D.; Chapman, A.R.; Langdon, T.; Sandeman, D.; Vaswani, A.; et al. High-sensitivity cardiac troponin i at presentation in patients with suspected acute coronary syndrome: A cohort study. *Lancet* **2015**, *386*, 2481–2488. [CrossRef]
472. Roos, A.; Bandstein, N.; Lundbäck, M.; Hammarsten, O.; Ljung, R.; Holzmann, M.J. Stable High-Sensitivity Cardiac Troponin T Levels and Outcomes in Patients with Chest Pain. *J. Am. Coll. Cardiol.* **2017**, *70*, 2226–2236. [CrossRef]
473. Kvistvik, B.; Mørkrid, L.; Røsjø, H.; Cvancarova, M.; Rowe, A.D.; Eek, C.; Bendz, B.; Edvardsen, T.; Gravning, J. High-sensitivity troponin T vs i in acute coronary syndrome: Prediction of significant coronary lesions and long-term prognosis. *Clin. Chem.* **2017**, *63*, 552–562. [CrossRef]
474. Tahhan, A.S.; Sandesara, P.; Hayek, S.S.; Hammada, M.; Alkhoder, A.; Kelli, H.M.; Topel, M.; O’Neal, W.T.; Ghasemzadeh, N.; Ko, Y.A.; et al. High-sensitivity troponin I levels and coronary artery disease severity, progression, and long-term outcomes. *J. Am. Heart Assoc.* **2018**, *7*, e007914. [CrossRef]
475. Omland, T.; de Lemos, J.A.; Sabatine, M.S.; Christophi, C.A.; Rice, M.M.; Jablonski, K.A.; Tjora, S.; Domanski, M.J.; Gersh, B.J.; Rouleau, J.L.; et al. A Sensitive Cardiac Troponin T Assay in Stable Coronary Artery Disease. *N. Engl. J. Med.* **2009**, *361*, 2538–2547. [CrossRef]
476. Vavik, V.; Pedersen, E.K.R.; Svingen, G.F.T.; Tell, G.S.; Schartum-Hansen, H.; Aakre, K.M.; Nygård, O.; Vikenes, K. Usefulness of Higher Levels of Cardiac Troponin T in Patients with Stable Angina Pectoris to Predict Risk of Acute Myocardial Infarction. *Am. J. Cardiol.* **2018**, *122*, 1142–1147. [CrossRef]
477. McQueen, M.J.; Kavsak, P.A.; Xu, L.; Shestakovska, O.; Yusuf, S. Predicting myocardial infarction and other serious cardiac outcomes using high-sensitivity cardiac troponin T in a high-risk stable population. *Clin. Biochem.* **2013**, *46*, 5–9. [CrossRef]
478. Saunders, J.T.; Nambi, V.; de Lemos, J.A.; Chambliss, L.E.; Virani, S.S.; Boerwinkle, E.; Hoogeveen, R.C.; Liu, X.; Astor, B.C.; Mosley, T.H.; et al. Cardiac Troponin T Measured by a Highly Sensitive Assay Predicts Coronary Heart Disease, Heart Failure, and Mortality in the Atherosclerosis Risk in Communities Study. *Circulation* **2011**, *123*, 1367–1376. [CrossRef] [PubMed]
479. De Filippi, C.R.; de Lemos, J.A.; Tkaczuk, A.T.; Christenson, R.H.; Carnethon, M.R.; Siscovick, D.S.; Gottdiener, J.S.; Seliger, S.L. Physical Activity, Change in Biomarkers of Myocardial Stress and Injury, and Subsequent Heart Failure Risk in Older Adults. *J. Am. Coll. Cardiol.* **2012**, *60*, 2539–2547. [CrossRef] [PubMed]
480. Ndumele, C.E.; Coresh, J.; Lazo, M.; Hoogeveen, R.C.; Blumenthal, R.S.; Folsom, A.R.; Selvin, E.; Ballantyne, C.M.; Nambi, V. Obesity, Subclinical Myocardial Injury, and Incident Heart Failure. *JACC Heart Fail.* **2014**, *2*, 600–607. [CrossRef] [PubMed]
481. McEvoy, J.W.; Chen, Y.; Ndumele, C.E.; Solomon, S.D.; Nambi, V.; Ballantyne, C.M.; Blumenthal, R.S.; Coresh, J.; Selvin, E. Six-Year Change in High-Sensitivity Cardiac Troponin T and Risk of Subsequent Coronary Heart Disease, Heart Failure, and Death. *JAMA Cardiol.* **2016**, *1*, 519. [CrossRef]
482. Evans, J.D.W.; Dobbin, S.J.H.; Pettit, S.J.; di Angelantonio, E.; Willeit, P. High-Sensitivity Cardiac Troponin and New-Onset Heart Failure: A Systematic Review and Meta-Analysis of 67,063 Patients with 4,165 Incident Heart Failure Events. *JACC Heart Fail.* **2018**, *6*, 187–197. [CrossRef]
483. Horwich, T.B.; Patel, J.; MacLellan, W.R.; Fonarow, G.C. Cardiac troponin I is associated with impaired hemodynamics, progressive left ventricular dysfunction, and increased mortality rates in advanced heart failure. *Circulation* **2003**, *108*, 833–838. [CrossRef]
484. Rienstra, M.; Yin, X.; Larson, M.G.; Fontes, J.D.; Magnani, J.W.; McManus, D.D.; McCabe, E.L.; Coglianese, E.E.; Amponsah, M.; Ho, J.E.; et al. Relation between soluble ST2, growth differentiation factor-15, and high-sensitivity troponin I and incident atrial fibrillation. *Am. Heart J.* **2014**, *167*, 109–115.e2. [CrossRef]

485. Filion, K.B.; Agarwal, S.K.; Ballantyne, C.M.; Eberg, M.; Hoogeveen, R.C.; Huxley, R.R.; Loehr, L.R.; Nambi, V.; Soliman, E.Z.; Alonso, A. High-sensitivity cardiac troponin T and the risk of incident atrial fibrillation: The Atherosclerosis Risk in Communities (ARIC) study. *Am. Heart J.* **2015**, *169*, 31–38.e3. [[CrossRef](#)]
486. Hijazi, Z.; Siegbahn, A.; Andersson, U.; Granger, C.B.; Alexander, J.H.; Atar, D.; Gersh, B.J.; Mohan, P.; Harjola, V.P.; Horowitz, J.; et al. High-sensitivity troponin I for risk assessment in patients with atrial fibrillation: Insights from the apixaban for reduction in stroke and other thromboembolic events in atrial fibrillation (ARISTOTLE) trial. *Circulation* **2014**, *129*, 625–634. [[CrossRef](#)]
487. Hijazi, Z.; Wallentin, L.; Siegbahn, A.; Andersson, U.; Alexander, J.H.; Atar, D.; Gersh, B.J.; Hanna, M.; Harjola, V.P.; Horowitz, J.D.; et al. High-sensitivity troponin T and risk stratification in patients with atrial fibrillation during treatment with apixaban or warfarin. *J. Am. Coll. Cardiol.* **2014**, *63*, 52–61. [[CrossRef](#)]
488. Zeller, T.; Tunstall-Pedoe, H.; Saarela, O.; Ojeda, F.; Schnabel, R.B.; Tuovinen, T.; Woodward, M.; Struthers, A.; Hughes, M.; Kee, F.; et al. High population prevalence of cardiac troponin I measured by a high-sensitivity assay and cardiovascular risk estimation: The MORGAM Biomarker Project Scottish Cohort. *Eur. Heart J.* **2014**, *35*, 271–281. [[CrossRef](#)] [[PubMed](#)]
489. Luers, C.; Sutcliffe, A.; Binder, L.; Irle, S.; Pieske, B. NT-proANP and NT-proBNP as prognostic markers in patients with acute decompensated heart failure of different etiologies. *Clin. Biochem.* **2013**, *46*, 1013–1019. [[CrossRef](#)] [[PubMed](#)]
490. Maisel, A.S.; Krishnaswamy, P.; Nowak, R.M.; McCord, J.; Hollander, J.E.; Duc, P.; Omland, T.; Storrow, A.B.; Abraham, W.T.; Wu, A.H.B.; et al. Rapid Measurement of B-Type Natriuretic Peptide in the Emergency Diagnosis of Heart Failure. *N. Engl. J. Med.* **2002**, *347*, 161–167. [[CrossRef](#)] [[PubMed](#)]
491. Omland, T.; Sabatine, M.S.; Jablonski, K.A.; Rice, M.M.; Hsia, J.; Wergeland, R.; Landaas, S.; Rouleau, J.L.; Domanski, M.J.; Hall, C.; et al. Prognostic Value of B-Type Natriuretic Peptides in Patients with Stable Coronary Artery Disease. The PEACE Trial. *J. Am. Coll. Cardiol.* **2007**, *50*, 205–214. [[CrossRef](#)]
492. Hall, C.; Rouleau, J.L.; Moye, L.; De Champlain, J.; Bichet, D.; Klein, M.; Sussex, B.; Packer, M.; Rouleau, J.; Arnold, M.O.; et al. N-terminal proatrial natriuretic factor: An independent predictor of long-term prognosis after myocardial infarction. *Circulation* **1994**, *89*, 1934–1942. [[CrossRef](#)]
493. Anand, I.S.; Fisher, L.D.; Chiang, Y.T.; Latini, R.; Masson, S.; Maggioni, A.P.; Glazer, R.D.; Tognoni, G.; Cohn, J.N. Changes in brain natriuretic peptide and norepinephrine over time and mortality and morbidity in the Valsartan Heart Failure Trial (Val-HeFT). *Circulation* **2003**, *107*, 1278–1283. [[CrossRef](#)]
494. Maisel, A.; Hollander, J.E.; Guss, D.; McCullough, P.; Nowak, R.; Green, G.; Saltzberg, M.; Ellison, S.R.; Bhalla, M.A.; Bhalla, V.; et al. Primary results of the Rapid Emergency Department Heart Failure Outpatient Trial (REDHOT): A multi-center study of B-type natriuretic peptide levels, emergency department decision making, and outcomes in patients presenting with shortness of breath. *J. Am. Coll. Cardiol.* **2004**, *44*, 1328–1333. [[CrossRef](#)]
495. Doust, J.A.; Pietrzak, E.; Dobson, A.; Glasziou, P.P. How well does B-type natriuretic peptide predict death and cardiac events in patients with heart failure: Systematic review. *Br. Med. J.* **2005**, *330*, 625–627. [[CrossRef](#)]
496. Kragelund, C.; Grønning, B.; Køber, L.; Hildebrandt, P.; Steffensen, R. N-Terminal Pro-B-Type Natriuretic Peptide and Long-Term Mortality in Stable Coronary Heart Disease. *N. Engl. J. Med.* **2005**, *352*, 666–675. [[CrossRef](#)]
497. Bibbins-Domingo, K.; Gupta, R.; Na, B.; Wu, A.H.B.; Schiller, N.B.; Whooley, M.A. N-terminal fragment of the prohormone brain-type natriuretic peptide (NT-proBNP), cardiovascular events, and mortality in patients with stable coronary heart disease. *J. Am. Med. Assoc.* **2007**, *297*, 169–176. [[CrossRef](#)]
498. Khan, S.Q.; Dhillon, O.; Kelly, D.; Squire, I.B.; Struck, J.; Quinn, P.; Morgenthaler, N.G.; Bergmann, A.; Davies, J.E.; Ng, L.L. Plasma N-Terminal B-Type Natriuretic Peptide as an Indicator of Long-Term Survival after Acute Myocardial Infarction: Comparison with Plasma Midregional Pro-Atrial Natriuretic Peptide. The LAMP (Leicester Acute Myocardial Infarction Peptide) Study. *J. Am. Coll. Cardiol.* **2008**, *51*, 1857–1864. [[CrossRef](#)] [[PubMed](#)]
499. Richards, A.M.; Nicholls, M.G.; Yandle, T.G.; Frampton, C.; Espiner, E.A.; Turner, J.G.; Buttimore, R.C.; Lainchbury, J.G.; Elliott, J.M.; Ikram, H.; et al. Plasma N-Terminal Pro-Brain Natriuretic Peptide and Adrenomedullin. *Circulation* **1998**, *97*, 1921–1929. [[CrossRef](#)] [[PubMed](#)]
500. Richards, A.M.; Nicholls, M.G.; Espiner, E.A.; Lainchbury, J.G.; Troughton, R.W.; Elliott, J.; Frampton, C.; Turner, J.; Crozier, I.G.; Yandle, T.G. B-type natriuretic peptides and ejection fraction for prognosis after myocardial infarction. *Circulation* **2003**, *107*, 2786–2792. [[CrossRef](#)] [[PubMed](#)]
501. Patton, K.K.; Ellinor, P.T.; Heckbert, S.R.; Christenson, R.H.; DeFilippi, C.; Gottdiener, J.S.; Kronmal, R.A. N-Terminal Pro-B-Type Natriuretic Peptide Is a Major Predictor of the Development of Atrial Fibrillation. *Circulation* **2009**, *120*, 1768–1774. [[CrossRef](#)]
502. Hijazi, Z.; Oldgren, J.; Andersson, U.; Connolly, S.J.; Ezekowitz, M.D.; Hohnloser, S.H.; Reilly, P.A.; Vinereanu, D.; Siegbahn, A.; Yusuf, S.; et al. Cardiac Biomarkers Are Associated with an Increased Risk of Stroke and Death in Patients with Atrial Fibrillation. *Circulation* **2012**, *125*, 1605–1616. [[CrossRef](#)] [[PubMed](#)]
503. Folsom, A.R.; Nambi, V.; Bell, E.J.; Oluleye, O.W.; Gottesman, R.F.; Lutsey, P.L.; Huxley, R.R.; Ballantyne, C.M. Troponin T, N-Terminal Pro-B-Type Natriuretic Peptide, and Incidence of Stroke. *Stroke* **2013**, *44*, 961–967. [[CrossRef](#)] [[PubMed](#)]
504. McCann, C.J.; Glover, B.M.; Menown, I.B.A.; Moore, M.J.; McEneny, J.; Owens, C.G.; Smith, B.; Sharpe, P.C.; Young, I.S.; Adey, J.A. Novel biomarkers in early diagnosis of acute myocardial infarction compared with cardiac troponin T. *Eur. Heart J.* **2008**, *29*, 2843–2850. [[CrossRef](#)]

505. Ishii, J.; Ozaki, Y.; Lu, J.; Kitagawa, F.; Kuno, T.; Nakano, T.; Nakamura, Y.; Naruse, H.; Mori, Y.; Matsui, S.; et al. Prognostic value of serum concentration of heart-type fatty acid-binding protein relative to cardiac troponin T on admission in the early hours of acute coronary syndrome. *Clin. Chem.* **2005**, *51*, 1397–1404. [[CrossRef](#)]
506. O'Donoghue, M.; De Lemos, J.A.; Morrow, D.A.; Murphy, S.A.; Buros, J.L.; Cannon, C.P.; Sabatine, M.S. Prognostic utility of heart-type fatty acid binding protein in patients with acute coronary syndromes. *Circulation* **2006**, *114*, 550–557. [[CrossRef](#)]
507. Viswanathan, K.; Kilcullen, N.; Morrell, C.; Thistlethwaite, S.J.; Sivananthan, M.U.; Hassan, T.B.; Barth, J.H.; Hall, A.S. Heart-Type Fatty Acid-Binding Protein Predicts Long-Term Mortality and Re-Infarction in Consecutive Patients with Suspected Acute Coronary Syndrome Who Are Troponin-Negative. *J. Am. Coll. Cardiol.* **2010**, *55*, 2590–2598. [[CrossRef](#)]
508. Arimoto, T.; Takeishi, Y.; Shiga, R.; Fukui, A.; Tachibana, H.; Nozaki, N.; Hiroto, J.; Miyamoto, T.; Hoit, B.D.; et al. Prognostic value of elevated circulating heart-type fatty acid binding protein in patients with congestive heart failure. *J. Card. Fail.* **2005**, *11*, 56–60. [[CrossRef](#)] [[PubMed](#)]
509. Maisel, A.; Mueller, C.; Neath, S.X.; Christenson, R.H.; Morgenthaler, N.G.; McCord, J.; Nowak, R.M.; Vilke, G.; Daniels, L.B.; Hollander, J.E.; et al. Copeptin helps in the early detection of patients with acute myocardial infarction: Primary results of the CHOPIN trial (Copeptin Helps in the early detection of Patients with acute myocardial INfarction). *J. Am. Coll. Cardiol.* **2013**, *62*, 150–160. [[CrossRef](#)] [[PubMed](#)]
510. Voors, A.A.; Von Haehling, S.; Anker, S.D.; Hillege, H.L.; Struck, J.; Hartmann, O.; Bergmann, A.; Squire, I.; Van Veldhuisen, D.J.; Dickstein, K. C-terminal provasopressin (copeptin) is a strong prognostic marker in patients with heart failure after an acute myocardial infarction: Results from the OPTIMAAL study. *Eur. Heart J.* **2009**, *30*, 1187–1194. [[CrossRef](#)] [[PubMed](#)]
511. Tu, W.J.; Dong, X.; Zhao, S.J.; Yang, D.G.; Chen, H. Prognostic value of plasma neuroendocrine biomarkers in patients with acute ischaemic stroke. *J. Neuroendocrinol.* **2013**, *25*, 771–778. [[CrossRef](#)]
512. de Marchis, G.M.; Katan, M.; Weck, A.; Fluri, F.; Foerch, C.; Findling, O.; Schuetz, P.; Buhl, D.; El-Koussy, M.; Gensicke, H.; et al. Copeptin adds prognostic information after ischemic stroke: Results from the CoRisk study. *Neurology* **2013**, *80*, 1278–1286. [[CrossRef](#)] [[PubMed](#)]
513. Katan, M.; Fluri, F.; Morgenthaler, N.G.; Schuetz, P.; Zweifel, C.; Bingisser, R.; Müller, K.; Meckel, S.; Gass, A.; Kappos, L.; et al. Copeptin: A novel, independent prognostic marker in patients with ischemic stroke. *Ann. Neurol.* **2009**, *66*, 799–808. [[CrossRef](#)]
514. Dhillon, O.S.; Khan, S.Q.; Narayan, H.K.; Ng, K.H.; Struck, J.; Quinn, P.A.; Morgenthaler, N.G.; Squire, I.B.; Davies, J.E.; Bergmann, A.; et al. Prognostic Value of Mid-Regional Pro-Adrenomedullin Levels Taken on Admission and Discharge in Non-ST-Elevation Myocardial Infarction. The LAMP (Leicester Acute Myocardial Infarction Peptide) II Study. *J. Am. Coll. Cardiol.* **2010**, *56*, 125–133. [[CrossRef](#)]
515. Bar-Or, D.; Winkler, J.V.; VanBenthuyzen, K.; Harris, L.; Lau, E.; Hetzel, F.W. Reduced albumin-cobalt binding with transient myocardial ischemia after elective percutaneous transluminal coronary angioplasty: A preliminary comparison to creatine kinase-MB, myoglobin, and troponin I. *J. Am. Heart J.* **2001**, *141*, 985–991. [[CrossRef](#)]
516. Christenson, R.H.; Show Hong, D.; Sanhai, W.R.; Wu, A.H.B.; Holtman, V.; Painter, P.; Branham, E.; Apple, F.S.; Murakami, M.; Morris, D.L. Characteristics of an albumin cobalt binding test for assessment of acute coronary syndrome patients: A multicenter study. *Clin. Chem.* **2001**, *47*, 464–470. [[CrossRef](#)] [[PubMed](#)]
517. Bar-Or, D.; Lau, E.; Winkler, J.V. A novel assay for cobalt-albumin binding and its potential as a marker for myocardial ischemia—A preliminary report. *J. Emerg. Med.* **2000**, *19*, 311–315. [[CrossRef](#)]
518. Bhagavan, N.V.; Lai, E.M.; Rios, P.A.; Yang, J.; Ortega-Lopez, A.M.; Shinoda, H.; Honda, S.A.A.; Rios, C.N.; Sugiyama, C.E.; Ha, C.E. Evaluation of human serum albumin cobalt binding assay for the assessment of myocardial ischemia and myocardial infarction. *Clin. Chem.* **2003**, *49*, 581–585. [[CrossRef](#)] [[PubMed](#)]
519. Shah, R.V.; Chen-Tournoux, A.A.; Picard, M.H.; van Kimmenade, R.R.J.; Januzzi, J.L. Galectin-3, cardiac structure and function, and long-term mortality in patients with acutely decompensated heart failure. *Eur. J. Heart Fail.* **2010**, *12*, 826–832. [[CrossRef](#)] [[PubMed](#)]
520. Lok, D.J.A.; Van Der Meer, P.; De La Porte, P.W.B.A.; Lipsic, E.; Van Wijngaarden, J.; Hillege, H.L.; Van Veldhuisen, D.J. Prognostic value of galectin-3, a novel marker of fibrosis, in patients with chronic heart failure: Data from the DEAL-HF study. *Clin. Res. Cardiol.* **2010**, *99*, 323–328. [[CrossRef](#)] [[PubMed](#)]
521. Caforio, A.L.P.; Pankuweit, S.; Arbustini, E.; Bassi, C.; Gimeno-Blanes, J.; Felix, S.B.; Fu, M.; Heliö, T.; Heymans, S.; Jahns, R.; et al. Current state of knowledge on aetiology, diagnosis, management, and therapy of myocarditis: A position statement of the European Society of Cardiology Working Group on Myocardial and Pericardial Diseases. *Eur. Heart J.* **2013**, *34*, 2636–2648. [[CrossRef](#)] [[PubMed](#)]
522. Jefferis, B.J.; Whincup, P.; Welsh, P.; Wannameethee, G.; Rumley, A.; Lennon, L.; Thomson, A.; Lawlor, D.; Carson, C.; Ebrahim, S.; et al. Prospective study of matrix metalloproteinase-9 and risk of myocardial infarction and stroke in older men and women. *Atherosclerosis* **2010**, *208*, 557–563. [[CrossRef](#)]
523. Welsh, P.; Whincup, P.H.; Papacosta, O.; Wannameethee, S.G.; Lennon, L.; Thomson, A.; Rumley, A.; Lowe, G.D.O. Serum matrix metalloproteinase-9 and coronary heart disease: A prospective study in middle-aged men. *Qjm* **2008**, *101*, 785–791. [[CrossRef](#)]
524. Jenkins, W.S.; Roger, V.L.; Jaffe, A.S.; Weston, S.A.; AbouEzzeddine, O.F.; Jiang, R.; Manemann, S.M.; Enriquez-Sarano, M. Prognostic Value of Soluble ST2 After Myocardial Infarction: A Community Perspective. *Am. J. Med.* **2017**, *130*, 1112.e9–1112.e15. [[CrossRef](#)]

525. Kohli, P.; Bonaca, M.P.; Kakkar, R.; Kudinova, A.Y.; Scirica, B.M.; Sabatine, M.S.; Murphy, S.A.; Braunwald, E.; Lee, R.T.; Morrow, D.A. Role of ST2 in Non-ST-Elevation Acute Coronary Syndrome in the MERLIN-TIMI 36 Trial. *Clin. Chem.* **2012**, *58*, 257–266. [CrossRef]
526. Pascual-Figal, D.A.; Ordoñez-Llanos, J.; Tornel, P.L.; Vázquez, R.; Puig, T.; Valdés, M.; Cinca, J.; de Luna, A.B.; Bayes-Genis, A.; MUSIC Investigators. Soluble ST2 for Predicting Sudden Cardiac Death in Patients with Chronic Heart Failure and Left Ventricular Systolic Dysfunction. *J. Am. Coll. Cardiol.* **2009**, *54*, 2174–2179. [CrossRef]
527. Januzzi, J.L.; Peacock, W.F.; Maisel, A.S.; Chae, C.U.; Jesse, R.L.; Baggish, A.L.; O'Donoghue, M.; Sakhija, R.; Chen, A.A.; van Kimmenade, R.R.J.; et al. Measurement of the Interleukin Family Member ST2 in Patients with Acute Dyspnea. Results from the PRIDE (Pro-Brain Natriuretic Peptide Investigation of Dyspnea in the Emergency Department) Study. *J. Am. Coll. Cardiol.* **2007**, *50*, 607–613. [CrossRef]
528. Pascual-Figal, D.A.; Manzano-Fernández, S.; Boronat, M.; Casas, T.; Garrido, I.P.; Bonaque, J.C.; Pastor-Perez, F.; Valdés, M.; Januzzi, J.L. Soluble ST2, high-sensitivity troponin T- and N-terminal pro-B-type natriuretic peptide: Complementary role for risk stratification in acutely decompensated heart failure. *Eur. J. Heart Fail.* **2011**, *13*, 718–725. [CrossRef] [PubMed]
529. Dieplinger, B.; Egger, M.; Haltmayer, M.; Kleber, M.E.; Scharnagl, H.; Silbernagel, G.; De Boer, R.A.; Maerz, W.; Mueller, T. Increased soluble ST2 predicts long-term mortality in patients with stable coronary artery disease: Results from the ludwigshafen risk and cardiovascular health study. *Clin. Chem.* **2014**, *60*, 530–540. [CrossRef] [PubMed]
530. Kaptoge, S.; Seshasai, S.R.K.; Gao, P.; Freitag, D.F.; Butterworth, A.S.; Borglykke, A.; Di Angelantonio, E.; Gudnason, V.; Rumley, A.; Lowe, G.D.O.; et al. Inflammatory cytokines and risk of coronary heart disease: New prospective study and updated meta-analysis. *Eur. Heart J.* **2014**, *35*, 578–589. [CrossRef]
531. Ridker, P.M.; Everett, B.M.; Thuren, T.; MacFadyen, J.G.; Chang, W.H.; Ballantyne, C.; Fonseca, F.; Nicolau, J.; Koenig, W.; Anker, S.D.; et al. Antiinflammatory Therapy with Canakinumab for Atherosclerotic Disease. *N. Engl. J. Med.* **2017**, *377*, 1119–1131. [CrossRef] [PubMed]
532. Ridker, P.M.; Rifai, N.; Stampfer, M.J.; Hennekens, C.H. Plasma concentration of interleukin-6 and the risk of future myocardial infarction among apparently healthy men. *Circulation* **2000**, *101*, 1767–1772. [CrossRef] [PubMed]
533. Kiran, B.S.R.; Mohanalakshmi, T.; Sri Kumar, R.; Prabhakar, E. Reddy C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in diabetes. *Int. J. Res. Pharm. Sci.* **2017**, *8*, 476–479.
534. The Lp-PLA2 Studies Collaboration. Lipoprotein-associated phospholipase A2 and risk of coronary disease, stroke, and mortality: Collaborative analysis of 32 prospective studies. *Lancet* **2010**, *375*, 1536–1544. [CrossRef]
535. Davì, G.; Tuttolomondo, A.; Santilli, F.; Basili, S.; Ferrante, E.; Di Raimondo, D.; Pinto, A.; Licata, G. CD40 ligand and MCP-1 as predictors of cardiovascular events in diabetic patients with stroke. *J. Atheroscler. Thromb.* **2009**, *16*, 707–713. [CrossRef]
536. Schönbeck, U.; Varo, N.; Libby, P.; Buring, J.; Ridker, P.M. Soluble CD40L and cardiovascular risk in women. *Circulation* **2001**, *104*, 2266–2268. [CrossRef]
537. di Napoli, M.; Papa, F.; Bocola, V. C-Reactive Protein in Ischemic Stroke. *Stroke* **2001**, *32*, 917–924. [CrossRef]
538. Gottdiener, J.S.; Arnold, A.M.; Aurigemma, G.P.; Polak, J.F.; Tracy, R.P.; Kitzman, D.W.; Gardin, J.M.; Rutledge, J.E.; Boineau, R.C. Predictors of congestive heart failure in the elderly: The cardiovascular health study. *J. Am. Coll. Cardiol.* **2000**, *35*, 1628–1637. [CrossRef]
539. Vasan, R.S.; Sullivan, L.M.; Roubenoff, R.; Dinarello, C.A.; Harris, T.; Benjamin, E.J.; Sawyer, D.B.; Levy, D.; Wilson, P.W.F.; D'Agostino, R.B. Inflammatory markers and risk of heart failure in elderly subjects without prior myocardial infarction: The Framingham Heart Study. *Circulation* **2003**, *107*, 1486–1491. [CrossRef] [PubMed]
540. Park, D.W.; Lee, S.W.; Yun, S.C.; Song, H.G.; Ahn, J.M.; Lee, J.Y.; Kim, W.J.; Kang, S.J.; Kim, Y.H.; Lee, C.W.; et al. A point-of-care platelet function assay and C-reactive protein for prediction of major cardiovascular events after drug-eluting stent implantation. *J. Am. Coll. Cardiol.* **2011**, *58*, 2630–2639. [CrossRef] [PubMed]
541. Chew, D.P.; Bhatt, D.L.; Robbins, M.A.; Penn, M.S.; Schneider, J.P.; Lauer, M.S.; Topol, E.J.; Ellis, S.G. Incremental prognostic value of elevated baseline C-reactive protein among established markers of risk in percutaneous coronary intervention. *Circulation* **2001**, *104*, 992–997. [CrossRef]
542. Rallidis, L.S.; Zolindaki, M.G.; Manioudaki, H.S.; Laoutaris, N.P.; Velissaridou, A.H.; Papasteriadis, E.G. Prognostic value of C-reactive protein, fibrinogen, interleukin-6, and macrophage colony stimulating factor in severe unstable angina. *Clin. Cardiol.* **2002**, *25*, 505–510. [CrossRef]
543. Ridker, P.M.; Cushman, M.; Stampfer, M.J.; Tracy, R.P.; Hennekens, C.H. Inflammation, Aspirin, and the Risk of Cardiovascular Disease in Apparently Healthy Men. *N. Engl. J. Med.* **1997**, *336*, 973–979. [CrossRef]
544. The Emerging Risk Factors Collaboration. C-reactive protein concentration and risk of coronary heart disease, stroke, and mortality: An individual participant meta-analysis. *Lancet* **2010**, *375*, 132–140. [CrossRef]
545. Lindahl, B.; Toss, H.; Siegbahn, A.; Venge, P.; Wallentin, L. Markers of Myocardial Damage and Inflammation in Relation to Long-Term Mortality in Unstable Coronary Artery Disease. *N. Engl. J. Med.* **2000**, *343*, 1139–1147. [CrossRef]
546. di Napoli, M.; Schwaninger, M.; Cappelli, R.; Ceccarelli, E.; Di Gianfilippo, G.; Donati, C.; Emsley, H.C.A.; Forconi, S.; Hopkins, S.J.; Masotti, L.; et al. Evaluation of C-Reactive Protein Measurement for Assessing the Risk and Prognosis in Ischemic Stroke. *Stroke* **2005**, *36*, 1316–1329. [CrossRef]
547. Aviles, R.J.; Martin, D.O.; Apperson-Hansen, C.; Houghtaling, P.L.; Rautaharju, P.; Kronmal, R.A.; Tracy, R.P.; Van Wagoner, D.R.; Psaty, B.M.; Lauer, M.S.; et al. Inflammation as a Risk Factor for Atrial Fibrillation. *Circulation* **2003**, *108*, 3006–3010. [CrossRef]

548. Anand, I.S.; Latini, R.; Florea, V.G.; Kuskowski, M.A.; Rector, T.; Masson, S.; Signorini, S.; Mocarelli, P.; Hester, A.; Glazer, R.; et al. C-reactive protein in heart failure: Prognostic value and the effect of Valsartan. *Circulation* **2005**, *112*, 1428–1434. [[CrossRef](#)] [[PubMed](#)]
549. Toker, A.; Karatas, Z.; Altin, H.; Karaarslan, S.; Cicekler, H.; Alp, H. Evaluation of serum ischemia modified albumin levels in acute rheumatic fever before and after therapy. *Indian J. Pediatr.* **2014**, *81*, 120–125. [[CrossRef](#)] [[PubMed](#)]
550. Yang, X.; Lin, Q.; Li, X.; Wu, L.; Xu, W.; Zhu, Y.; Deng, H.; Zhang, Y.; Yao, B. Cystatin C is an important biomarker for cardiovascular autonomic dysfunction in Chinese type 2 diabetic patients. *J. Diabetes Res.* **2019**, *2019*, 1706964. [[CrossRef](#)] [[PubMed](#)]
551. van der Laan, S.W.; Fall, T.; Soumaré, A.; Teumer, A.; Sedaghat, S.; Baumert, J.; Zabaneh, D.; van Setten, J.; Isgum, I.; Galesloot, T.E.; et al. Cystatin C and Cardiovascular Disease. *J. Am. Coll. Cardiol.* **2016**, *68*, 934–945. [[CrossRef](#)]
552. Oh, J.-Y. Serum Cystatin C as a Biomarker for Predicting Coronary Artery Disease in Diabetes. *Korean Diabetes J.* **2010**, *34*, 84. [[CrossRef](#)]